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Research Article

Asiatic acid Improves Physical Fatigue through Regulating Inflammatory Reaction, Oxidative Stress and Energy Metabolism in Mice

¹Liu Xianchu, ¹Cheng Changhao and ²Liu Ming

Abstract

Background and Objective: Fatigue is a complex physiological phenomenon in daily life, which can easily evoke physical dysfunction and serious illness. Asiatic acid with multifaceted biological activities has been used in food and medicinal fields for a long period. The purpose of this study was to explore the potential anti-fatigue effect and mechanisms of asiatic acid in high-intensity exercise. **Materials and Methods:** The exercise-fatigue model was constituted via a forced swimming test. The experiment mice were continuously fed with asiatic acid for 28 days. The exhaustive time, fatigue relevant biochemical indices, inflammatory cytokines, oxidation parameters and energy metabolism indicators were detected to explore evidence of asiatic acid in high-intensity exercise. **Results:** Current researches showed that asiatic acid significantly prolonged swimming exhaustive time. In high-intensity exercise, asiatic acid dramatically alleviated fatigue relevant biochemical indices, including CK, LDH, AST, ALT, SUN and BLA to protect against organ injury. Moreover, asiatic acid relieved TNF-α and IL-6 levels to restrain inflammatory reaction. In muscle tissue, asiatic acid markedly elevated SOD, CAT and GSH-Px activities to promote antioxidant ability and observably alleviated MDA level to eliminate oxidative injury. Asiatic acid remarkably increased SDH and Na+-K+-ATPase activities to ameliorate energy metabolism. In addition, asiatic acid markedly activated PGC-1α, TFAM, NRF-2 and HO-1 expression to increase energy metabolism and suppress oxidative stress in excessive exercise. **Conclusion:** These results suggested asiatic acid could improve exercise-induced fatigue via inhibition of inflammatory response, prevention of oxidative stress damage and improvement of energy metabolism in mice. Asiatic acid could serve as a novel potential candidate to postpone fatigue.

Key words: Asiatic acid, fatigue, inflammatory reaction, oxidative stress, energy metabolism

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Corresponding Author: Liu Ming, Faculty of Science, College of Furong, Hunan University of Arts and Science, 415000, Changde, Hunan, China

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Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

¹Institute of Physical Culture, Hunan University of Arts and Science, 415000, Changde, Hunan, China

²Faculty of Science, College of Furong, Hunan University of Arts and Science, 415000, Changde, Hunan, China

INTRODUCTION

Fatigue is a common and complex phenomenon in the general population, which influences mood, causes myalgia and affects physical performance to lower the quality of life, even evoking serious illness¹. High-intensity exercise is one of the underlying factors of physical fatigue. The pathophysiological mechanism of physical fatigue is complicated and there are some theories to illuminate the possible potential physiological process after high-intensity exercise. Firstly, extra and excessive intermediate and end metabolites, such as LA and BUN, can be produced in highintensity exercise. These biochemical parameters were confirmed to be linked to fatigue². Secondly, over-production of reactive oxygen species leads to oxidation stress in highintensity exercise. Then, disruption of oxidative balance was involved in the pathophysiology of fatigue³. Thirdly, intensive muscle contraction is the cause of inflammation reaction, evidenced by accumulation of inflammatory cytokine, which has a detrimental effect on organs, such as liver and muscle⁴. Last but not least, energy is not only to maintain normal physical activity but also play a crucial role in high-intensity exercise-induced fatigue. SDH and ATPase are closely involved in the production of ATP, which are related elements of material and energy metabolism⁵. Currently, the research focus of anti-fatigue is to enhance physical performance and mitigate recovery time after exercise-induced fatigue.

Asiatic acid is one of the triterpenes, which are naturally biosynthesized by cyclization. Asiatic acid is widely cultivated in Asian and African countries and has been regarded as a vegetable, spice and herb in food and medicinal fields. Nowadays, Asiatic acid has been confirmed as a functional food and therapeutic drug that possesses plentiful pharmacological activities in tissues and organs. In addition, heart and pulmonary function play a vital position in highintensity exercise. Previous research reported asiatic acid alleviated doxorubicin-induced heart, liver and kidney toxicities through enhancing NRF-2 protein expression⁶. Asiatic acid was also shown to ameliorate hypochlorous acid-induced pulmonary fibrosis via regulating advanced oxidation protein and TGF-β1/Smad2/3 signaling⁷. What is more, asiatic acid is involved in energy metabolism, which is closely associated with exercise-induced physical fatigue. In a hypoxia environment, asiatic acid attenuated cardiomyocytes apoptosis by miR-1290/HIF3A/HIF-1α signaling⁸. In glucose uptake, asiatic acid improved diabetic-induced glucose response and promoted glucose homeostasis by antioxidant defense9. In the lactate signalling cascade, asiatic acid mitigated lactate-induced apoptosis via suppression of oxidative injury¹⁰. This study investigated the asiatic acid exerted improvement on physical fatigue through regulating inflammatory reaction, oxidative stress and energy metabolism.

MATERIALS AND METHODS

Study area: The study was carried out at the Human Movement Science Laboratory, Hunan University of Arts and Science in May-September, 2021.

Chemicals and reagents: Asiatic acid was purchased from Sigma (St. Louis, MO, United States). The purity of asiatic acid surpassed 95% by HPLC. The assay kits of BLA, CK, LDH, BUN, AST, ALT, SOD, CAT, GPX, MDA, SDH and Na⁺-K⁺-ATPase were purchased from Nanjing Jiancheng Biotechnology Institute (Nanjing, China). The detection kits of TNF- α and IL-6 were purchased from BOSTER Biological Technology Co., Ltd., (Wuhan, China).

Animals and treatment: A total of 80 male ICR mice (7 weeks old, 18-22 g) were purchased from Hunan SJA Laboratory Animal Co., Ltd. (Changsha, China). Mice lived in specific pathogen-free conditions. After 7 days of acclimatization, mice were randomly assigned into 4 groups: (1) Control group (Con), (2) 1 mg kg⁻¹ asiatic acid (AA-L), (3) 10 mg kg⁻¹ asiatic acid (AA-M) and (4) 20 mg kg⁻¹ asiatic acid (AA-H). Asiatic acid was treated by gavage for 28 successive days. The control group was given an equivalent dose of distilled water. The body weight, liver, lung, kidney and heart were weighed by scales to calculate organ index. The form of organ index is 100 g G⁻¹. All animal experiments were inspected according to the Ethics Committee of Hunan University of Arts and Science (No. HUAS-2021-TY-179).

Forced swimming test: Weight-loaded forced swimming test was used to measure exhaust time. Briefly, 30 min after the last treatment, the mouse with a lead (5% of body weight) swam individually in the pool (24-26°C, 30 cm deep). Swimming time was defined when the mouse did not return to the surface for greater than 10 sec.

Biochemical assessment: After 28 days of intervention, mice underwent a 90 min swimming test without any loads. After resting for 1 hr, mice were anaesthetized. Blood was withdrawn by removing the eyeball. Then blood was centrifuged (1500 g, 4°C, 10 min) to prepare serum. The fatigue-associated biochemical indicators of LA, CK, LDH, AST, ALT and SUN were tested to evaluate fatigue and injury of muscle, liver and kidney.

Inflammatory variables determination: The parameters of TNF- α and IL-6 as inflammatory cytokines, were analyzed by the ELISA method. In 96-well plates, the biotin-labelled anti-mouse antibody of TNF- α and IL-6 was incubated (37°C, 90 min). After washing 3 times, avidin-peroxidase was added into plates and reacted (37°C, 30 min). The plate was again washed 5 times and TMB substrate was combined (37°C, 20 min). The wavelength of absorbance was 405 nm.

Oxidant stress, energy metabolism detection: After anesthetization, muscles were immediately excised from each group, frozen by liquid nitrogen and collected at -80°C. The tissue was weighed, ground and centrifuged (4000 g, 4°C, 15 min) to detect SOD, CAT, GSH-PX and MDA. At the end of experiments, the content of SDH and Na⁺-K⁺-ATPase were determined in muscle to evaluate the efficiency of energy metabolism.

mRNA gene expression: RNA from muscle was extracted by TRIzol and transformed into cDNA. mRNA expression was detected with SYBR by Bio-Rad CFX real-time PCR. Primers for PGC-1 α , TFAM, NRF-2, HO-1 and β -actin were designed by Sangon Biotech (Shanghai) Co., Ltd., (Table 1). Gene expression was calculated via standardization to β -actin.

Statistical analysis: All results were expressed as Mean \pm SD. The significance was demonstrated by a one-way ANOVA. p<0.05 were deemed significant.

RESULTS

Properties of AA on exhaustive exercise time: The swimming endurance was shown in Fig. 1. The exhaustive swimming time of control, AA-L, AA-M and AA-H groups were 9.22, 10.83, 15.43 and 21.08 min, respectively. Comparison to the control group, exhaustive time in AA-M and AA-H groups was markedly prolonged by 67, 129%, respectively (p<0.01), which demonstrated AA had prominent properties on movement and endurance in high-intensity exercise.

Properties of AA on CK, LDH, ALT, AST, SUN and BLA: The biochemical parameters were tested to evaluate the anti-fatigue efficacy of AA. Figure 2a showed CK levels of control, AA-L, AA-M and AA-H groups were 1.96, 1.87, 1.37 and 1.20 U mL⁻¹, respectively. Figure 2b showed LDH levels of control, AA-L, AA-M and AA-H groups were 1.74, 1.60, 1.27 and 1.19 U mL⁻¹, respectively. Figure 2c showed ALT levels of control, AA-L, AA-M and AA-H groups were 53.22, 50.19, 41.26

and 39.51 U L $^{-1}$, respectively. Figure 2d showed AST levels of control, AA-L, AA-M and AA-H groups were 79.98, 71.97, 60.76 and 55.67 U L $^{-1}$, respectively. Figure 2e showed SUN levels of control, AA-L, AA-M and AA-H groups were 13.55, 12.53, 9.48 and 8.30 mmol L $^{-1}$, respectively. Figure 2f showed BLA levels of control, AA-L, AA-M and AA-H groups were 18.68, 17.67 L, 13.48 and 11.59 mmol L $^{-1}$, respectively. The levels of biochemical parameters in AA-M and AA-H groups were visibly reduced compared to the control group (p<0.01), which demonstrated AA prevented muscle damage, liver damage and kidney damage to made positive contribution in high-intensity exercise.

Properties of AA on inflammatory response in serum: The proinflammatory cytokines, including TNF- α and IL-6, were examined to evaluate the anti-inflammatory function of AA. Figure 3a showed TNF- α levels of control, AA-L, AA-M and AA-H groups were 98.04, 92.11, 82.83 and 74.62 pg mL⁻¹, respectively. Figure 3b showed IL-6 levels of control, AA-L, AA-M and AA-H groups were 54.82, 49.98, 43.71 and 36.89 pg mL⁻¹, respectively. TNF- α and IL-6 levels in AA-M and AA-H groups were relieved compared to the control group (p<0.05, p<0.01), which demonstrated AA delayed physical fatigue by regulating inflammatory mediators in high-intensity exercise.

Properties of AA on oxidation stress in muscle: The oxidation indices, including SOD, CAT, GSH-Px and MDA, were measured to estimate the anti-oxidant function of AA. Figure 4a showed SOD activities of control, AA-L, AA-M and AA-H groups were 86.65, 95.39, 106.53 and 111.09 U mg⁻¹ pro, respectively.

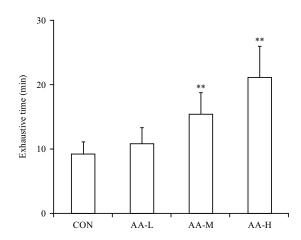


Fig. 1: Properties of AA on weight-loaded swimming time (n = 10 mice/group)

*p<0.05 and **p<0.01 versus control group

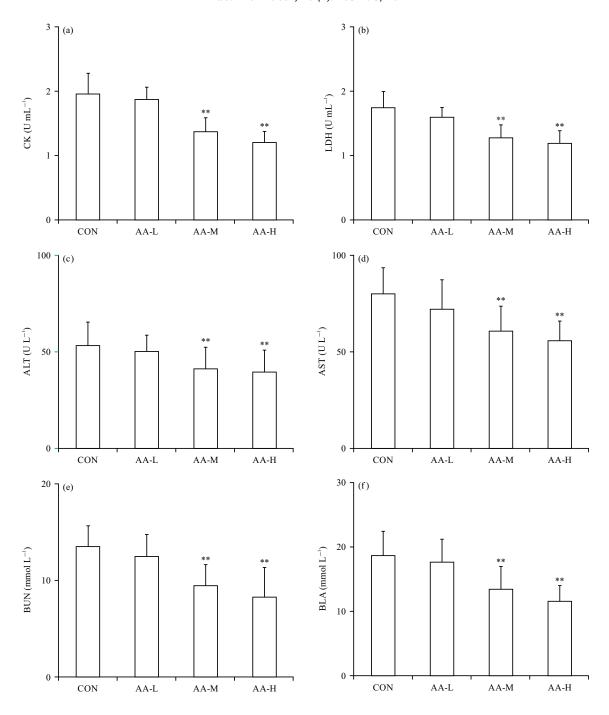


Fig. 2(a-f): Properties of AA on biochemical parameters relevant to fatigue (n = 10 mice/group), (a) Creatine kinase (CK), (b) Lactate dehydrogenase (LDH), (c) Alanine aminotransferase (ALT), (d) Aspartate aminotransferase (AST), (e) Serum urea nitrogen (SUN) and (f) Blood lactic acid (BLA)

*p<0.05 and **p<0.01 versus control group

Figure 4b showed CAT activities of control, AA-L, AA-M and AA-H groups were 26.92, 30.19, 34.14 and 36.47 U mg⁻¹ pro, respectively. Figure 4c showed GSH-Px activities of control, AA-L, AA-M and AA-H groups were 41.15, 43.51, 51.79 and 55.03 U mg⁻¹ pro, respectively. Figure 4d showed MDA levels

of control, AA-L, AA-M and AA-H groups were 3.56, 3.31, 3.09 and 2.48 nmol mg⁻¹ pro, respectively. The activities of SOD, CAT and GSH-Px in AA-M and AA-H groups were markedly elevated, while MDA levels in AA-M and AA-H groups were significantly alleviated compared to the control group

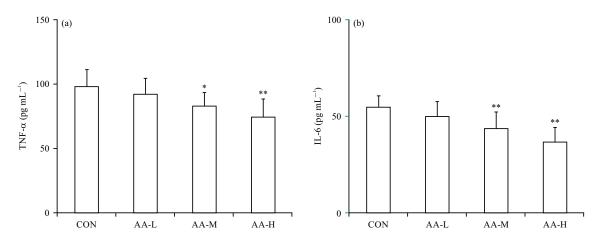


Fig. 3(a-b): Properties of AA on the inflammatory response (n = 10 mice/group), (a) Tumor necrosis factor (TNF- α) and (b) Interleukin 6 (IL-6)

*p<0.05 and **p<0.01 versus control group

Table 1: Primers were used in this study¹¹⁻¹³

Gene	Accession numbers	Primer sequences
PGC-1α	NM_008904	
	Forward primer	5'-TATGGAGTGACATAGAGTGTGCT-3'
	Reverse primer	5'-CCACTTCAATCCACCCAGAAAG-3'
TFAM	NM_009360	
	Forward primer	5'-CCTGAGGAAAAGCAGGCATA-3'
	Reverse primer	5'-TCACTTCGTCCAACTTCAGC-3'
NRF-2	NM_010902	
	Forward primer	5'-TCCGCTGCCATCAGTCAGTC-3'
	Reverse primer	5'-ATTGTGCCTTCAGCGTGCTTC-3'
HO-1	NM_010442	
	Forward primer	5'-TGCAGGTGATGCTGACAGAGG-3'
	Reverse primer	5'-GGGATGAGCTAGTGCTGATCTGG-3'
β-actin	NM_007393	
	Forward primer	5'-GATTACTGCTCTGGCTCCTAGC-3'
	Reverse primer	5'-GACTCATCGTACTCCTGCTTGC-3'

(p<0.05, p<0.01). These results implied AA exerted a positive role in physical fatigue by increasing antioxidant ability and eliminating oxidative injury in high-intensity exercise.

Properties of AA on energy metabolism in muscle: The SDH and Na⁺-K⁺-ATPase, as vital regulatory enzymes in anabolism and catabolism, were detected to estimate the ATP generation rate of AA. Figure 5a showed SDH activities of control, AA-L, AA-M and AA-H groups were 0.82, 0.85, 0.97 and 0.99 U mg⁻¹ pro, respectively. Figure 5b showed Na⁺-K⁺-ATPase activities of control, AA-L, AA-M and AA-H groups were 0.95, 1.04, 1.13 and 1.22 U mg⁻¹ pro, respectively. SDH and Na⁺-K⁺-ATPase activities in AA-M and AA-H groups were remarkably increased compared to the control group (p<0.05, p<0.01), which manifested AA possessed anti-fatigue properties by improving energy metabolism in high-intensity exercise.

Properties of AA on the expression of PGC-1 α , TFAM, NRF-2 and HO-1 in muscle: The PGC-1α, TFAM, NRF-2 and HO-1 as crucial regulatory factors in energy metabolism and antioxidation in muscle, were detected to evaluate the antifatigue function of AA. Figure 6a showed PGC-1α relative expression of control, AA-L, AA-M and AA-H groups were 1, 1.09, 1.57 and 1.74, respectively. Figure 6b showed TFAM relative expression of control, AA-L, AA-M and AA-H groups were 1, 1.12, 1.34 and 1.55, respectively. Figure 6c showed NRF-2 relative expression of control, AA-L, AA-M and AA-H groups were 1, 1.12, 1.50 and 1.64, respectively. Figure 6d showed HO-1 relative expression of control, AA-L, AA-M and AA-H groups were 1, 1.11, 1.71 and 2.06, respectively. The relative expressions of PGC-1α, TFAM, NRF-2 and HO-1 in AA-M and AA-H groups were significantly activated compared to the control group (p<0.05, p<0.01), which demonstrated AA was effective in anti-fatigue by regulating energy metabolism and oxidative injury in excessive exercise.

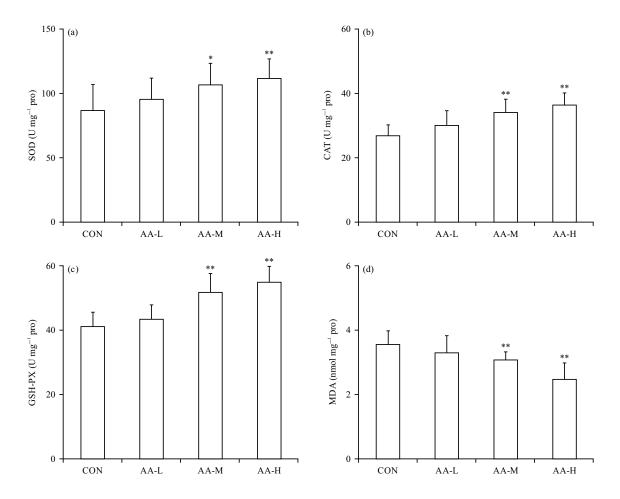


Fig. 4(a-d): Properties of AA on oxidation parameters (n = 10 mice/group), (a) Superoxide dismutase (SOD), (b) Catalase (CAT) (c) Glutathione peroxidase (GSH-Px) and (d) Malondialdehyde (MDA)

*p<0.05 and **p<0.01 versus control group

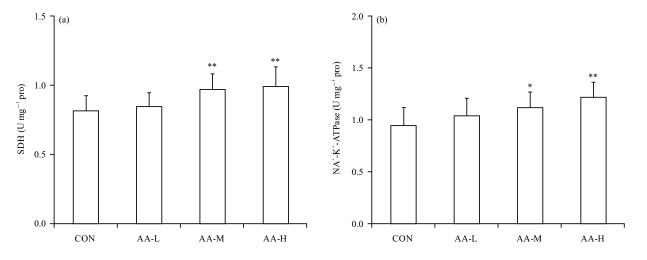


Fig. 5(a-b): Properties of AA on energy metabolism (n = 10 mice/group), (a) Saccharopine dehydrogenase (SDH) and (b) Na^+-K^+-ATP ase *p<0.05 and **p<0.01 versus control group

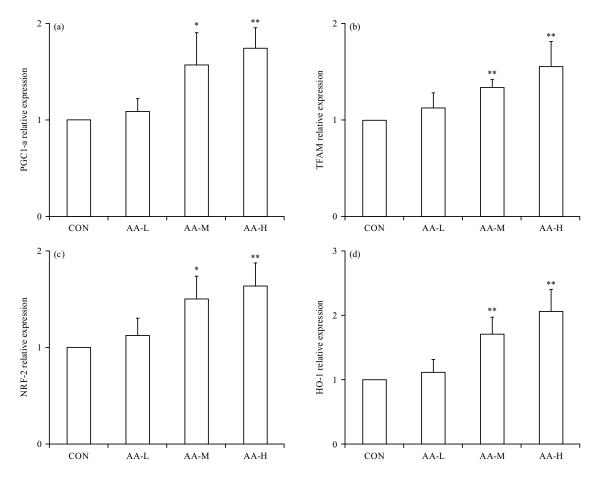


Fig. 6(a-d): Properties of AA on modulating PGC-1 α , TFAM, NRF-2 and HO-1 in muscle (n = 3 mice/group), (a) Peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC-1 α), (b) Transcription factor A, mitochondrial (TFAM), (c) Nuclear factor (erythroid-derived 2)-like 2 (NRF-2) and (d) Heme oxygenase-1 (HO-1) *p<0.05 and **p<0.01 versus control group

DISCUSSION

In this experiment, 1st monitored the roles of asiatic acid by forced swimming test which was approved as a well-founded animal model to assess anti-fatigue activity in present researches¹⁴. The indicator of exhausting time was used to manifest movement endurance and physical fatigue susceptibility. In weight-loaded forced swimming test, treatment with asiatic acid to mice over 28 days increased exhausted exercise time in dose-dependent relations. The middle and high dosages of asiatic acid concentration increased exercise capacity by 67 and 129%, respectively. These results indicated that asiatic acid was a safe and valid anti-fatigue agent.

BLA is an important indicator of fatigue. In high-intensity exercise, a large quantity of energy was generally formed via aerobic and anaerobic respiration to furnish tissues needs. In this case, lactic acid is mainly produced in skeletal muscles

during the aerobic respiration process. What is more, an excess of BLA aggravates the degree of fatigue and propels fatigue development¹⁵. Extra lactic acid can further dissociate to manufacture hydrogen ions in the blood, which lowers pH level to affect internal environment homeostasis. More seriously, excess lactic acid derived from muscles, in turn, bring about suppression of muscle contraction and muscle damage^{16,17}. Hence, bioaccumulation of BLA is a high correlation with fatigue. Moreover, treatment with asiatic acid ameliorated lactate concentration and glucose homeostasis to inhibit hyperlactatemia in malaria¹⁸. In this study, results demonstrated asiatic acid exerted the effect of alleviating BLA levels in high-intensity exercise.

CK and LDH, as cytosolic enzymes, principally consist in muscle tissue. CK and LDH are biomarkers of muscle injury. CK is a critical kinase and increased CK level was clinically associated with muscle-relevant diseases, such as muscular dystrophy, muscle contraction, muscle disruption, muscle

necrosis and myocardial infarction¹⁹. LDH is involved in eliminating BLA²⁰. In a long period of train, excess and detrimental intermediate metabolites could result in myocyte damage. Then, CK and LDH are over-produced and permeate into serum²¹. Hence, excessive CK and LDH activity could play a critical role in decreasing exercise capacity and increasing the degree of movement endurance. What is more, asiatic acid was demonstrated to reduce CK vitalities and suppress apoptosis in MI/R injury²². In PC12 cells, asiatic acid relieved leakage and inhibit LD content in OGD/R injury²³. In this study, results demonstrated asiatic acid ameliorated CK and LDH activities to attenuate muscle damage and physical fatigue in high-intensity exercise.

SUN is not only clinically applied to assess renal function, but also positively correlated with fatigue. SUN is a metabolite end-product that is generated in protein metabolism²⁴. In strenuous exercise, energy generated from carbohydrates and fats cannot meet the requirement of the body. Then, protein as an energy substance is consumed in catabolic metabolism to supply energy. Generally, SUN is formed in the liver and excreted through the kidney. However, supernormal SUN increases the burden of renal function and a part of BUN is released into the blood. Therefore, the vitality of SUN plays a crucial role in the anti-fatigue effect. In addition, treatment with asiatic acid improved carbohydrate metabolic disorder and decreased urea level to protect renal function in diabetic rats²⁵. In this study, asiatic acid was able to dramatically alleviate the bioaccumulation of SUN in high-intensity exercise, indicating that asiatic acid could strengthen movement endurance and exercise ability by reducing protein metabolism-provided energy.

AST and ALT in serum are authorized as important parameters to evaluate liver injury²⁶. Under normal circumstances, the activities of AST and ALT are low. Once their activities are increased, which indicates liver injury is occurring or has occurred. Previous research proved that strenuous exercise is harmful to liver function^{27,28}. Therefore, AST and ALT are biochemical parameters relevant to fatigue. In addition, asiatic acid was reported to improve liver damage by reducing the release of AST in ischemia/reperfusion rat²⁹. In this study, asiatic acid notably mitigated AST and ALT activities in high-intensity exercise, indicating that the anti-fatigue effect of asiatic acid was associated with its protection on liver damage.

Strenuous exercise evokes uncoordinated activation of systemic cytokine balance. Inflammatory cytokines were unconventionally altered by heavy exertion, such as long-term endurance training, intense exercise and marathon³⁰. As a secondary reaction, oxidative stress also triggers an

inflammatory response in the living body³¹. What is more, the inflammatory reaction was involved in the pathophysiology of muscle damage³². In return, muscle injury with elevated inflammatory factors, such as TNF- α and IL-6, on physiological fatigue may induce muscle pain and efficacy deficits³³. Hence, the inflammatory response may be a causative factor in physiological fatigue. Although physiology researchers have reported a potential connection between asiatic acid and inflammatory cytokines, little is known concerning the effect of asiatic acid on inflammatory reaction after high-intensity exercise. Therefore, inflammatory cytokines, including TNF- α and IL-6, were detected to assess the anti-fatigue properties of asiatic acid. In this study, results notarized that asiatic acid had anti-fatigue effects via suppression of inflammatory reaction.

Oxidation stress is now believed to participate in the progression of fatigue. In high-intensity exercise, the process of energy production is accompanied by large amounts of free radical, which is harmful for antioxidation and systemic redox balance. Generally, SOD, CAT and GSH-Px are vital antioxidants to exclude ROS, while MDA is a metabolite of lipid peroxidation to aggravate oxidative reaction. Previous studies have demonstrated inhibition of oxidative stress is an effective mean to enhance exercise performance and delay the degree of fatigue³⁴. What is more, asiatic acid increased the activity of SOD and reduced the level of MDA to inhibit cardiac fibrosis in spontaneous hypertension rats³⁵. In this study, asiatic acid observably prevented and reduced oxidative injury against fatigue by enhancing SOD, CAT and GSH-Px activities and lowering MDA content.

The mitochondrion is a crucial venue of ATP generation. ATP is regarded as direct energy in normal physiological performance, especially in high-intensity exercise³⁶. Mitochondrial function is used as a vital indicator to evaluate physiological fatigue in high-intensity exercise³⁷. In energy metabolism, various biological enzymes, such as SDH and Na+-K+-ATPase, play a regulatory role in anabolism and catabolism. SDH as a rate-limiting enzyme, participates in the TCA cycle to regulate ATP synthesis³⁸. SDH can affect the electron transport chain in energy metabolism³⁹. Na⁺-K⁺-ATPase as a degradation enzyme, performs its function via hydrolyzing ATP to replenish energy⁴⁰. Under normal conditions, SDH and Na+-K+-ATPase are maintained in normal physiological standards. The previous result showed that small molecule oligopeptides as a potent anti-fatique candidate, enhanced movement endurance and exercise capacity by increasing the activity of SDH and Na+-K+-ATPase⁴¹. What is more, Pre-treatments of asiatic acid might retain SDH and Na⁺-K⁺-ATPase activity in high-glucose-induced damage⁴². In this study, asiatic acid markedly maintained SDH and Na⁺-K⁺-ATPase activity to improve energy metabolism and delay fatigue.

PGC-1 α is known as a regulator to improve transcription in mitochondria 43 . PGC-1 α is also vitally associated with energy metabolism to modulate muscle function. PGC-1α promotes mitochondrial biogenesis partly by regulating TFAM and NRF-2, which is closely associated with antioxidant capacity⁴⁴. Previous results reported anyulignan enhanced muscle function to mitigate fatigue via enhancement expression of PGC-1 α and NRF-2 in high-intensity exercise⁴⁵. In addition, NRF-2 is an important transcription factor connected with protection against oxidative injury. NRF-2 controls antioxidant enzyme gene expression, such as HO-1, Cu, Zn-SOD and Gsr, to resistant body's oxidative response⁴⁶. By enhancing NRF-2 expression, it exhibited cellular oxidation resistance function by augmenting SOD and GSH-Px activities and decreasing ROS and MDA contents, which was closely related to anti-fatigue⁴⁷. In this study, asiatic acid-activated expression of PGC-1α, TFAM, NRF-2 and HO-1 in excessive exercise, which demonstrated that asiatic acid increased energy metabolism and restrained oxidative stress to protect against fatigue.

CONCLUSION

In summary, asiatic acid defended against exercise-induced fatigue in mice. The mechanism of asiatic acid on anti-fatigue efficacy is relevant to inhibition of inflammatory response, prevention of oxidative stress damage and improvement of energy metabolism. These researches affirm that asiatic acid has potential as a protection and therapy agent for exercise-induced fatigue.

SIGNIFICANCE STATEMENT

This experiment proved asiatic acid observably prolonged swimming exhaustive time. In high-intensity exercise, the fatigue relevant indicators, including CK, LDH, AST, ALT, SUN and BLA, were mitigated by treatment of asiatic acid. The anti-fatigue efficacy of asiatic acid is partly due to its effects on restraining inflammatory reaction, as represented by lower TNF- α and IL-6 levels in serum. Especially in muscle tissue, asiatic acid observably alleviated oxidative injury by enhancing SOD and CAT, GSH-Px activities and reducing MDA levels. In addition, asiatic acid remarkably ameliorated energy metabolism by increasing SDH and Na⁺-K⁺-ATPase activities. Lastly, asiatic acid significantly activated PGC-1 α , TFAM, NRF-2 and HO-1 expression in excessive exercise. In all, results

showed asiatic acid possessed anti-fatigue efficacy through regulating inflammatory reaction, oxidative stress and energy metabolism in mice. Asiatic acid can be developed as a beneficial agent to postpone fatigue.

ACKNOWLEDGMENTS

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REFERENCES

- Induruwa, I., C.S. Constantinescu and B. Gran, 2012. Fatigue in multiple sclerosis-a brief review. J. Neurol. Sci., 323: 9-15.
- 2. Pedersen, T.H., O.B. Nielsen, G.D. Lamb and D.G. Stephenson, 2004. Intracellular acidosis enhances the excitability of working muscle. Science, 305: 1144-1147.
- 3. Wang, J.J., M.J. Shieh, S.L. Kuo, C.L. Lee and T.M. Pan, 2006. Effect of red mold rice on antifatigue and exercise-related changes in lipid peroxidation in endurance exercise. Applied Microbiol. Biotechnol., 70: 247-253.
- 4. Bernecker, C., J. Scherr, S. Schinner, S. Braun, W.A. Scherbaum and M. Halle, 2013. Evidence for an exercise induced increase of TNF- α and IL-6 in marathon runners. Scand. J. Med. Sci. Sports, 23: 207-214.
- Xu, C., J. Lv, Y.M. Lo, S.W. Cui, X. Hu and M. Fan, 2013. Effects of oat β-glucan on endurance exercise and its anti-fatigue properties in trained rats. Carbohydr. Polym., 92: 1159-1165.
- Kamble, S.M. and C.R. Patil, 2018. Asiatic acid ameliorates doxorubicin-induced cardiac and hepato-renal toxicities with Nrf2 transcriptional factor activation in rats. Cardiovasc. Toxicol., 18: 131-141.
- 7. Xia, X., C. Dai, H. Yu, X. Huang, A. Chen, Y. Tan and L. Wang, 2018. Asiatic acid prevents the development of interstitial lung disease in a hypochlorous acid-induced mouse model of scleroderma. Oncol. Lett., 15: 8711-8716.
- 8. Wu, K., M. Hu, Z. Chen, F. Xiang and G. Chen *et al.*, 2017. Asiatic acid enhances survival of human AC16 cardiomyocytes under hypoxia by upregulating miR-1290. IUBMB Life, 69: 660-667.
- Ramachandran, V. and R. Saravanan, 2015. Glucose uptake through translocation and activation of GLUT4 in PI3K/Akt signaling pathway by asiatic acid in diabetic rats. Hum. Exp. Toxicol., 34: 884-893.
- 10. Gao, C., F. Wang, Z. Wang, J. Zhang and X. Yang, 2016. Asiatic acid inhibits lactate-induced cardiomyocyte apoptosis through the regulation of the lactate signaling cascade. Int. J. Mol. Med., 38: 1823-1830.

- Choi, W.H., H.J. Son, Y.J. Jang, J. Ahn, C.H. Jung and T.Y. Ha, 2017. Apigenin ameliorates the obesity induced skeletal muscle atrophy by attenuating mitochondrial dysfunction in the muscle of obese mice. Mol. Nutr. Food Res., Vol. 61. 10.1002/mnfr.201700218.
- Bao, L., X. Cai, J. Wang, Y. Zhang, B. Sun and Y. Li, 2016. Anti-fatigue effects of small molecule oligopeptides isolated from *Panax ginseng* C. A. meyer in mice. Nutrients, Vol. 8. 10.3390/nu8120807.
- 13. Xu, X., Y. Ding, Y. Yang, Y. Gao and Q. Sun *et al.*, 2018. β-glucan salecan improves exercise performance and displays anti-fatigue effects through regulating energy metabolism and oxidative stress in mice. Nutrients, Vol.10. 10.3390/nu10070858.
- 14. Wang, X., C. Niu, J. Lu, N. Li and J. Li, 2014. Hydrolyzed protein supplementation improves protein content and peroxidation of skeletal muscle by adjusting the plasma amino acid spectrums in rats after exhaustive swimming exercise: A pilot study. J. Int. Soc. Sports Nutr., Vol. 11. 10.1186/1550-2783-11-5.
- 15. Xianchu, L., L. Ming, L. Xiangbin and Z. Lan, 2018. Grape seed proanthocyanidin extract supplementation affects exhaustive exercise-induced fatigue in mice. Food Nutr. Res., Vol. 62. 10.29219/fnr.v62.1421.
- Chen, X., D. Liang, Z. Huang, G. Jia, H. Zhao and G. Liu, 2021.
 Anti fatigue effect of quercetin on enhancing muscle function and antioxidant capacity. J. Food Biochem., Vol. 45. 10.1111/jfbc.13968.
- 17. Harahap, N.S. and R. Amelia, 2019. Red dragon fruit (*Hylocereus polyrhizus*) extract decreases lactic acid level and creatine kinase activity in rats receiving heavy physical exercise. Open Access Maced. J. Med. Sci., 7: 2232-2235.
- Alfred, M.G., M.B. Nkazimulo, M.M. Vuyisile and M.C. Tagumirwa, 2016. Asiatic acid influences glucose homeostasis in *P. berghei* murine malaria infected sprague-dawley rats. Afr. J. Tradit Complement Altern. Med., 13: 91-101.
- Daab, W., M.A. Bouzid, M. Lajri, M. Bouchiba, M.A. Saafi and H. Rebai, 2020. Chronic beetroot juice supplementation accelerates recovery kinetics following simulated match play in soccer players. J. Am. Coll. Nutr., 40: 61-69.
- 20. Zhao, H.P., Y. Zhang, Z. Liu, J.Y. Chen, S.Y. Zhang, X.D. Yang and H.L. Zhou, 2017. Acute toxicity and anti-fatigue activity of polysaccharide-rich extract from corn silk. Biomed. Pharmacother., 90: 686-693.
- Herrlinger, K.A., D.M. Chirouzes and M.A. Ceddia, 2015. Supplementation with a polyphenolic blend improves postexercise strength recovery and muscle soreness. Food Nutr. Res., Vol. 59. 10.3402/fnr.v59.30034.

- 22. Dai, Y., Z. Wang, M. Quan, Y. Lv, Y. Li, H.B. Xin and Y. Qian, 2018. Asiatic acid protests against myocardial ischemia/reperfusion injury via modulation of glycometabolism in rat cardiomyocyte. Drug Des., Dev. Ther., 12: 3573-3582.
- 23. Yuan, J.P., J.M. Lu and Y. Lu, 2013. The protective effect of asiatic acid against oxygen-glucose deprivation/reoxygenation injury of PC12 cells. Yao Xue Xue Bao, 48: 1738-1742.
- Anand, T., G.P. Kumar, M.D. Pandareesh, M.S.L. Swamy, F. Khanum and A.S. Bawa, 2012. Effect of bacoside extract from *Bacopa monniera* on physical fatigue induced by forced swimming. Phytother. Res., 26: 587-593.
- Kalidhindi, S., V.V.S. Uddandrao, V. Sasikumar, N. Raveendran and S. Ganapathy, 2020. Mitigating perspectives of asiatic acid in the renal derangements of streptozotocinnicotinamide induced diabetic rats. Cardiovasc. Hematol. Agents Medic. Chem., 18: 37-44.
- 26. Anderson, F.H., L. Zeng, N.R. Rock and E.M. Yoshida, 2000. An assessment of the clinical utility of serum ALT and AST in chronic hepatitis C. Hepatol. Res., 18: 63-71.
- 27. Vale, A.F., H.H. Ferreira, E.J. Benetti, A.C.S. Rebelo, A.C.R. Figueiredo, E.C. Barbosa and K. Simões, 2019. Antioxidant effect of the pequi oil (*Caryocar brasiliense*) on the hepatic tissue of rats trained by exhaustive swimming exercises. Braz. J. Biol., 79: 257-262.
- Deng, Y., Q. Tang, Y. Zhang, R. Zhang, Z. Wei, X. Tang and M. Zhang, 2017. Protective effect of *Momordica charantia* water extract against liver injury in restraint-stressed mice and the underlying mechanism. Food Nutr. Res., Vol. 61. 10.1080/16546628.2017.1348864.
- Lu, Y., H. Kan, Y. Wang, D. Wang, X. Wang, J. Gao and L. Zhu, 2018. Asiatic acid ameliorates hepatic ischemia/ reperfusion injury in rats via mitochondria-targeted protective mechanism. Toxicol. Appl. Pharmacol., 338: 214-223.
- Liu, L., X. Wu, B. Zhang, W. Yang and D. Li et al., 2017. Protective effects of tea polyphenols on exhaustive exercise-induced fatigue, inflammation and tissue damage. Food Nutr. Res., Vol. 61. 10.1080/16546628.2017.1333390.
- 31. Pedersen, B.K. and M.A. Febbraio, 2008. Muscle as an endocrine organ: Focus on muscle-derived interleukin-6. Physiol. Rev., 88: 1379-1406.
- 32. Gholamnezhad, Z., M.H. Boskabady and M. Hosseini, 2014. Effect of *Nigella sativa* on immune response in treadmill exercised rat. BMC Complement. Altern. Med., Vol. 14. 10.1186/1472-6882-14-437.
- 33. Lanier, A.B., 2003. Use of nonsteroidal anti-inflammatory drugs following exercise-induced muscle injury. Sports Med., 33: 177-186.

- Martins, N.O., I.M. de Brito, S.S.O. Araújo, G. Negri, E. de Araújo Carlini and F.R. Mendes, 2018. Antioxidant, anticholinesterase and antifatigue effects of *Trichilia catigua* (Catuaba). BMC Compl. Alt. Med., Vol. 18. 10.1186/s12906-018-2222-9.
- 35. Meng, Z., H.Y. Li, C.Y. Si, Y.Z. Liu and S. Teng, 2019. Asiatic acid inhibits cardiac fibrosis through Nrf2/HO-1 and TGF-β1/Smads signaling pathways in spontaneous hypertension rats. Int. Immunopharmacol., Vol. 74. 10.1016/j.intimp.2019.105712.
- Manfredi, G., L. Yang, C.D. Gajewski and M. Mattiazzi, 2002. Measurements of ATP in mammalian cells. Methods, 26: 317-326.
- Aoi, W., Y. Ogaya, M. Takami, T. Konishi and Y. Sauchi et al., 2015. Glutathione supplementation suppresses muscle fatigue induced by prolonged exercise via improved aerobic metabolism. J. Int. Soc. Sports Nutr., Vol. 12. 10.1186/s12970-015-0067-x.
- 38. Tan, S.J., N. Li, F. Zhou, Q.T. Dong, X.D. Zhang, B.C. Chen and Z. Yu, 2014. Ginsenoside Rb1 improves energy metabolism in the skeletal muscle of an animal model of postoperative fatigue syndrome. J. Surg. Res., 191: 344-349.
- 39. Zhou, Q., C. Zhang, S. Cheng, B. Wei, X. Liu and S. Ji, 2014. Changes in energy metabolism accompanying pitting in blueberries stored at low temperature. Food Chem., 164: 493-501.
- 40. Huang, X.P., H. Tan, B.Y. Chen and C.Q. Deng, 2012. *Astragalus* extract alleviates nerve injury after cerebral ischemia by improving energy metabolism and inhibiting apoptosis. Bio. Pharm. Bull., 35: 449-454.

- 41. Li, D., J.W. Ren, T. Zhang, R. Liu, L. Wu, Q. Du and Y. Li, 2018. Anti-fatigue effects of small-molecule oligopeptides isolated from *Panax quinquefolium* L. in mice. Food Funct., 9: 4266-4273.
- 42. Chan, C.Y., M.C. Mong, W.H. Liu, C.Y. Huang and M.C. Yin, 2014. Three pentacyclic triterpenes protect H9c2 cardiomyoblast cells against high-glucose-induced injury. Free Radical Res., 48: 402-411.
- 43. Olesen, J., K. Kiilerich and H. Pilegaard, 2010. PGC-1 α -mediated adaptations in skeletal muscle. Pflügers Archiv Eur. J. Physiol., 460: 153-162.
- 44. Lin, J., C. Handschin and B.M. Spiegelman, 2005. Metabolic control through the PGC-1 family of transcription coactivators. Cell Metab., 1: 361-370.
- 45. Zhang, X., S. Jing, H. Lin, W. Sun and W. Jiang *et al.*, 2019. Anti-fatigue effect of anwulignan via the Nrf2 and PGC-1 α signaling pathway in mice. Food Funct., 10: 7755-7766.
- 46. Yao, J., S. Peng, J. Xu and J. Fang, 2019. Reversing ROS mediated neurotoxicity by chlorogenic acid involves its direct antioxidant activity and activation of Nrf2 ARE signaling pathway. BioFactors, 45: 616-626.
- Wang, X., Y. Qu, Y. Zhang, S. Li and Y. Sun *et al.*, 2018. Antifatigue potential activity of *Sarcodon imbricatus* in acute excise-treated and chronic fatigue syndrome in mice via regulation of Nrf2-mediated oxidative stress. Oxidative Med. Cell. Longevity, Vol. 2018. 10.1155/2018/9140896.