ISSN 1682-8356 ansinet.org/ijps



# POULTRY SCIENCE



308 Lasani Town, Sargodha Road, Faisalabad - Pakistan Mob: +92 300 3008585, Fax: +92 41 8815544 E-mail: editorijps@gmail.com

#### International Journal of Poultry Science

ISSN 1682-8356 DOI: 10.3923/ijps.2018.605.609



## Research Article Effect of Fermentation using *Trichoderma harzianum* and *Saccharomyces cerevisiae* on Crude Protein, Crude fibre and Zinc Content of Duckweed

Hendi Setiyatwan, E. Harlia, D. Rusmana, Tubagus Benito and Lovita Adriani

Faculty of Animal Husbandry, University of Padjadjaran, Bandung-Sumedang Street KM. 21, Sumedang 45363, Indonesia

### Abstract

**Background and Objectives:** Duckweed has a potential to be used as poultry feed, however due to its high crude fibre content, its inclusion at high levels is limited. This could be overcome by means of fermentation with microbial strains like *Trichoderma harzianum* and *Saccharomyces cerevisiae*. Therefore, the present study was designed in order to improve the nutritive value of duckweed by fermentation with *Trichoderma harzianum* and *Saccharomyces cerevisiae*. **Materials and Methods:** There were five treatments, each having 4 replicates, thus making it a total of 20 experimental units. The treatments consisted of P1: (Fermentation using *Trichoderma harzianum* for 1 day followed by *Saccharomyces cerevisiae* fermentation for 9 days), P2: (*Trichoderma harzianum* for 3 days followed by *Saccharomyces cerevisiae* fermentation for 5 days), P4: (*Trichoderma harzianum* for 7 days), P3: (*Trichoderma harzianum* for 5 days followed by *Saccharomyces cerevisiae* fermentation of a days followed by *Saccharomyces cerevisiae* fermentation for 1 day). **Results:** The fermentation of 2 days and P5: (*Trichoderma harzianum* for 9 days followed by *Saccharomyces cerevisiae* fermentation of a days) and P5: (*Trichoderma harzianum* for 9 days followed by *Saccharomyces cerevisiae* fermentation of 1 day). **Results:** The fermentation of Duckweed. The best combination that improved the nutritive value of duckweed in terms of increasing the crude protein (33.88%) and zinc (88.6%) content and decreasing the crude fibre (8.16%) content was P2 in which duckweed was fermented with *Trichoderma harzianum* for 3 days followed by *Saccharomyces cerevisiae* fermentation for 7 days. **Conclusion:** Fermentation with *Trichoderma harzianum* for 3 days followed by *Saccharomyces cerevisiae* fermentation for 7 days. **Conclusion:** Fermentation with *Trichoderma harzianum* for 3 days followed by *Saccharomyces cerevisiae* fermentation for 7 days. **Conclusion:** Fermentation with *Trichoderma harzianum* for 3 days followed by *Saccharomyces ce* 

Key words: Duckweed, fermentation, nutrient quality, Saccharomyces cerevisiae, Trichoderma harzianum

Citation: Hendi Setiyatwan, E. Harlia, D. Rusmana, Tubagus Benito and Lovita Adriani, 2018. Effect of fermentation using *Trichoderma harzianum* and *Saccharomyces cerevisiae* on crude protein, crude fibre and zinc content of duckweed. Int. J. Poult. Sci., 17: 605-609.

Corresponding Author: Hendi Setiyatwan, Faculty of Animal Husbandry, University of Padjadjaran, Bandung-Sumedang Street KM. 21 Sumedang 45363, Indonesia Tel: +6222 7798241 Fax: +6222 7798212

Copyright: © 2018 Hendi Setiyatwan *et al.* This is an open access article distributed under the terms of the creative commons attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

#### INTRODUCTION

Duckweed has been known for a long period of time as a potential source of food for humans and animals. It is a water plant that lives floating on the surface of the water. It is proliferated vegetatively<sup>1</sup> with the total biomass production reaching 1.76-2.34%<sup>2,3</sup>. It contains 22.4% crude protein and 10.16% crude fiber<sup>2</sup>. Moreover, it has amino acids {such as lysine (6.9%), methionine (1.4%) and histidine (2.7%)}<sup>4</sup> and high content of minerals and vitamin A<sup>5</sup>. It has been used as an animal feed<sup>6-8</sup>. The use of Duckweed in poultry nutrition is limited to 5% level due to high crude fiber content, unbalanced essential amino acids and low Zn content.

To overcome the problem of limited use of Duckweed, fermentation is a viable option. Fermentation can increase the nutrient availability and reduce the amount of anti-nutrients<sup>6</sup>. Fermentation also increases the content of proteins, essential amino acids, vitamins and bioactive compounds<sup>9</sup>. Thapa and Tamang<sup>10</sup> also reported that fermentation process increased the content of vitamin B12, vitamin B complex and C, lysine and tryptophan and various minerals. Furthermore, Astuti *et al.*<sup>11</sup> reported that fermentation improves the content of folic acid, niacin, riboflavin, nicotinamide and pyridoxine.

The use of Trichoderma harzianum for fermentation has been reported to have positive effect on lowering of crude fiber. Ghosh and Chattopadhyay<sup>12</sup> reported that the crude fiber of palm kernel flour decreased after fermentation because of the use of Trichoderma harzianum. Moreover, Setiyatwan<sup>7</sup> indicated that the crude fiber content of duckweed decreased by 76.06% after fermentation by Trichoderma harzianum for 6-9 days. Saccharomyces *cerevisiae* is a yeast that can produce heterologous proteins<sup>13</sup>. The fermentation using Saccharomyces cerevisiae yeast can enrich the material with amino acid dl-methionine and inorganic Zinc (Zn) mineral. Zn is an important element in the growth and metabolism of Saccharomyces cerevisiae. In addition, zinc is involved in the structure and function of protein and nucleic acids<sup>14</sup>. Saccharomyces cerevisiae can combine Zn ions of origin substrate. The amount of Zn absorbed depends on the Zn source, cell physiology, specific surface properties of the organism and the physicochemical influence of the environment<sup>15</sup>. The concentration of Zn in the substrate affects the ability of Saccharomyces cerevisiae to form metal bonds<sup>16</sup>.

Duckweed quality can be improved by fermentation process using *Trichoderma harzianum* and *Saccharomyces cerevisiae*. The secondary metabolic pathway of *Trichoderma harzianum* and *Saccharomyces cerevisiae* are synergistic. *Trichoderma harzianum* can provide nitrogen for the growth of *Saccharomyces cerevisiae* and subsequently used as a raw material for the formation of metallic bonds and amino acids. The activity of both microbes produces complementary effects for better results, thus imitates a natural microbial growth medium. Duckweed fermentation process using two microbes (co-culture) is expected to produce functional complementary enzymes due to the mutual expression of metabolic pathways in substrate utilization. However, the research on the quality improvement of duckweed through fermentation using *Trichoderma harzianum* and *Saccharomyces cerevisiae* has not been carried out. Therefore, the purpose of this research was to enhance the nutritive value of Duckweed by fermentation process using two types of microbes.

#### **MATERIALS AND METHODS**

**Materials and research tools:** The research material used was duckweed, potato dextrose, PDA agar, Aquadest, ZnCO<sub>3</sub>, dl-methionine, rice, tripton, cotton, *Trichoderma harzianum* and *Saccharomyces cerevisiae*. The equipments consisted of analytical scales, technical scales, measuring flask 1000 mL, Erlenmeyer 1000 mL, stirrer mugs, reaction tubes, Oase needles, spirtus lights, Erlenmeyer glasses, sterilisator, beaker glass, etiquette paper, electric oven, refrigerator, autoclave and fermentor.

**Research procedure:** Substrate consisted of a mixture of duckweed and selective medium. The mixture was boiled in the water for 60 min at a temperature of 115 °C and pressure of 1.1 atmospheres. *Trichoderma harzianum* microbes were incubated in each treatment @  $3 \times 10^7$  spores/100 g of the substrate for 1, 3, 5, 7 and 9 days at room temperature. After first stage fermentation, ZnCO<sub>3</sub> (186 ppm) and dl-methionine (286 ppm) were added. Afterwards, substrate was fermented using *Saccharomyces cerevisiae* @  $3 \times 10^7$  spores/100 g of substrate for 9, 7, 5, 3 and 1 day. Thereafter, the product was dried in the oven.

**Experimental design:** The experimental design used was Completely Randomized Design (CRD). There were five treatments, each having 4 replicates, thus making it a total of 20 experimental units. The treatments consisted of:

- **P1:** Fermentation using *Trichoderma harzianum* for 1day followed by *Saccharomyces cerevisiae* fermentation for 9 days
- P2: Trichoderma harzianum for 3 days followed by Saccharomyces cerevisiae fermentation for 7 days

- **P1:** *Trichoderma harzianum* for 5 days followed by *Saccharomyces cerevisiae* fermentation for 5 days
- **P4:** *Trichoderma harzianum* for 7 days followed by *Saccharomyces cerevisiae* fermentation for 3 days
- **P5:** *Trichoderma harzianum* for 9 days followed by *Saccharomyces cerevisiae* fermentation for 1 day

#### **Parameters recorded**

- **Crude protein:** Crude protein content (based on Dry Matter) was determined by Wendee method. The proximate analysis was carried out by Kjeldahl method. It consisted of destruction, distillation and titration stages
- **Crude fiber:** The analytical method used to determine crude fiber (based on Dry Matter) was by proximate analysis using Wendee method. It involves dissolving the soluble ingredients in strong acids and bases. The material is then filtered out leaving the remaining residue. This residue is crude fiber comprising of cellulose and slight lignin and pentosan
- **Zinc content:** The Zn content was analyzed by Atomic Absorption Spectrophotometer (AAS)

**Statistical analysis:** Data collected were subjected to one-way analysis of variance (ANOVA) as per Steel and Torrie. If there were differences in means, they were compared using Duncan's multiple range test with 5% significant level<sup>17</sup>.

#### **RESULTS AND DISCUSSION**

**Crude protein content of fermented duckweed:** The fermentation of Duckweed using *Trichoderma harzianum* and *Saccharomyces cerevisiae* was helpful in increasing the protein content of duckweed (Table 1). The results of the present study are in-line with other studies, such as Omwango *et al.*<sup>18</sup> and Ezekiel and Aworh<sup>19</sup> who improved the protein content of Pineapple waste (up to 45%) and Cassava peel by various microbes.

The highest Duckweed protein content was observed in the group in which Duckweed was fermented using *Trichoderma harzianum* for 3 day followed by *Saccharomyces cerevisiae* fermentation for 7 days (P2). This was followed by the groups P1, P3 and P4. The significantly (p<0.05) lowest Duckweed protein content was observed in the group in which Duckweed was fermented using *Trichoderma harzianum* for 9 day followed by *Saccharomyces cerevisiae* fermentation for 1 day (P5). This is in line with the study of Kuswardani and Wijajaseputra<sup>20</sup> who stated that the short duration fermentation using *Saccharomyces cerevisiae* decreases the protein content. The longer fermentation with *Saccharomyces cerevisiae* was found to be more useful in increasing the Duckweed protein content. This is in agreement with the results of Kustyawati *et al.*<sup>21</sup> who reported that the addition of *Saccharomyces cerevisiae* can be used as a duckweed modification agent to increase its protein content and solubility.

**Crude fiber content of fermented duckweed:** The effect of fermentation on crude fibre content in duck weed is given in Table 1. The lowest crude fibre content in duckweed was observed in the group in which duckweed was fermented using *Trichoderma harzianum* for 3 days followed by *Saccharomyces cerevisiae* fermentation for 7 days (P2). This was followed by the groups P3, P4 and P5. Highest crude fibre content in duckweed was fermented using *Trichoderma harzianum* for 1 days followed by *Saccharomyces cerevisiae* fermentation for 7 days (P2). This of the groups P3, P4 and P5. Highest crude fibre content in duckweed was observed in the group in which duckweed was fermented using *Trichoderma harzianum* for 1 day followed by *Saccharomyces cerevisiae* fermentation for 9 days (P1).

Dietary fibre is the part of plant material consisting mainly of cellulosic and non-cellulosic polysaccharides and a non-carbohydrate component, lignin. These components are highly resistant to hydrolysis by alimentary enzymes and therefore cannot be digested or absorbed in the blood stream. So, high crude fibre is not desirable. Hatta *et al.*<sup>22</sup> stated that *Trichoderma* sp. can reduce lignocellulose. Furthermore, Nsereko *et al.*<sup>23</sup> suggested that *Trichoderma harzianum* can break the bonds of lignin and cellulose because it produces cellulase, xylosidase and xylanase enzymes. Shaikh *et al.*<sup>24</sup> reported *Saccharomyces cerevisiae* decreased the crude fibre content of cottonseed cake.

**Zinc content of fermented duckweed:** The effect of fermentation on zinc content in duck weed is given in Table 1. The highest zinc content in duck weed was observed in the

Table 1: Crude protein, crude fiber and Zn content of duckweed fermented with Trichoderma harzianum followed by Saccharomyces cerevisiae

Parameters (%)	Treatments				
	 P1	P2	P3	P4	P5
Crude protein	31.84±0.76 <sup>b</sup>	33.88±0.85°	31.90±0.82 <sup>b</sup>	31.01±0.82 <sup>b</sup>	29.73±0.89ª
Crude fiber	16.27±0.82°	8.16±0.87ª	10.27±0.84 <sup>b</sup>	10.25±0.82 <sup>b</sup>	10.33±0.83 <sup>b</sup>
Mineral Zn	83.08±1.43ª	88.60±0.96°	86.60±2.46 <sup>b</sup>	$84.60 \pm 0.80^{ab}$	84.60±1.72 <sup>ab</sup>

Means in the same column with different superscript differ significantly (p<0.05)

group in which duck weed was fermented using *Trichoderma harzianum* for 3 days followed by *Saccharomyces cerevisiae* fermentation for 7 days (P2). This was followed by the groups P3 in which duck weed was fermented using *Trichoderma harzianum* for 5 days followed by *Saccharomyces cerevisiae* fermentation for 5 days. The lowest zinc content in duck weed was observed in the group in which duck weed was fermented using *Trichoderma harzianum* for 1 day followed by *Saccharomyces cerevisiae* fermented using *Trichoderma harzianum* for 1 day followed by *Saccharomyces cerevisiae* fermentation for 9 days (P1).

The difference in the zinc content of duckweed among various treatments was observed due to the fact that the nutrient content of duckweed depends on the environment where it is grown<sup>25</sup>. Zinc in the feed is in the form of inorganic mineral salts, such as Zn oxide<sup>26</sup>. Schneider *et al.*<sup>27</sup> stated that Zn is the essential element and acts as a co-factor for many proteins and enzymes.

#### CONCLUSION

Fermentation with *Trichoderma harzianum* and *Saccharomyces cerevisiae* was found effective in modifying the nutrient composition of duckweed. The best combination that improved the nutritive value of duckweed in terms of increasing the crude protein and zinc content and decreasing the crude fibre content was *Trichoderma harzianum* fermentation for 3 days followed by *Saccharomyces cerevisiae* fermentation for 7 days. Hence, this combination is recommended for improving the nutritive value of duckweed for poultry feeding.

#### SIGNIFICANCE STATEMENT

This study discovers the methods to modify the nutrient quality of duckweed for its incorporation in poultry feed. This study will help the researchers to study the effect of fermentation in improving the nutritive value of duckweed by using various microbial strains which many researchers were not able to explore earlier. Thus, a new theory on the improvement of quality of duckweed through fermentation using a combination of microbes may be arrived at.

#### ACKNOWLEDGMENT

The authors would like to thank the honorable Rector of Padjadjaran University, Ministry of Research, Technology and Higher Education of Padjadjaran University who supported this research. The research was carried out with the financial support from DIPA Fund of Universitas Padjadjaran no. 855/UN6.3.1/PL/2017 dated March 8th, 2017.

#### REFERENCES

- Kittiwongwattana, C. and S. Vuttipongchaikij, 2013. Effects of nutrient media on vegetative growth of *Lemna minor*and *Landoltia punctata* during *in vitro* and *ex vitro* cultivation. Maejo Int. J. Sci. Technol., 7: 60-69.
- Nopriani, U., P.D.M.H. Karti and I. Prihantoro, 2014. Productivity of duckweed (*Lemna* sp. *minor*) as a forage alternative feed of livestock at different light intensities. JITV., 19: 272-286.
- Tanuwiria, U.H. and Febrianto, 2017. Long drying plant lemna sp against dry matter content, crude protein and crude fiber. Proceedings of the National Biology Seminar: Utilization of Biodiversity Based on Local Wisdom, UIN Sunan Gunung Djati Bandung, Aprir 13, 2017, Department of Faculty Biology Science and Technology UIN Sunan Gunung Djati., pp: 300-309.
- Akter, M., S.D. Chowdhury, Y. Akter and M.A. Khatun, 2011. Effect of duckweed (*Lemna minor*) meal in the diet of laying hen and their performance. Bangladesh Res. Publ. J., 5: 252-261.
- Gwaze, F.R. and M. Mwale, 2015. The prospect of duckweed in pig nutrition: A review. J. Agric. Sci., Vol. 7. 10.5539/jas.v7n11p189
- Haustein, A.T., R.H. Gillman, P.W. Skillicorn, V. Vergara, V. Guevara and A. Gastanaduy, 1990. Duckweed, a useful strategy for feeding chickens: Performance of layers fed with sewage-grown *Lemnacea* species. Poult. Sci., 69: 1835-1844.
- 7. Setiyatwan, H., 2007. Quality improvement of duckweed nutrition through fermentation using *Trichoderma harzianum*. J. Anim. Sci. Padjadjaran Univ., 7: 111-116.
- 8. Ngamsaeng, A., S. Thy and T.R. Preston, 2004. Duckweed (*Lemna minor*) and water spinach (*Ipomoea aquatica*) as protein supplements for ducks fed broken rice as the basal diet. Livestock Res. Rural Dev., 16: 18-24.
- Tamang, J.P., K. Watanabe and W.H. Holzapfel, 2016. Review: Diversity of microorganisms in global fermented foods and beverages. Front. Microbiol., Vol. 7. 10.3389/fmicb.2016.00377
- 10. Thapa, N. and J.P. Tamang, 2015. Functionality and Therapeutic Values of Fermented Foods. In: Health Benefits of Fermented Food and Beverages, Tamang, J.P. (Ed.)., CRC Press, USA., pp: 111-168.
- 11. Astuti, M., A. Meliala, F.S. Dalais and M.L. Wahlqvist, 2000. Tempe, a nutritious and healthy food from Indonesia. Asia Paci. J. Clin. Nutr., 9: 322-325.
- 12. Ghosh, D. and P. Chattopadhyay, 2011. Preparation of idli batter, its properties and nutritional improvement during fermentation. J. Food Sci. Technol., 48: 610-615.
- Ginting, S.P. and R. Krisnan, 2006. The Effect of fermentation using several strains of trichoderma and different incubation period to the chemical composition of palm kernel. Ministry of Agriculture Indonesia, Livestock Research Center Indonesia, September 5-6, 2016, pp: 944.

- 14. Hensing, M.C.M., R.J. Rouwenhorst, J.J. Heijnen, J.P. van Dijken and J.T. Pronk, 1995. Physiological and technological aspects of large-scale heterologous-protein production with yeasts. Antonie Leeuwenhoek, 67: 261-279.
- Castro, C.E. and J.S. Sevall, 1993. Zinc Deficiency, Chromatin Structure and Gene Expression. In: Nutrient Modulation of The Immune Response, Cunningham-Rundles, S. (Ed.)., Marcel Dekker, New York, pp: 141-150.
- 16. De Nicola, R., N. Hall, T. Bollag, G. Thermogiannis and G.M. Walker, 2009. Zinc accumulation and utilization by wine yeasts. Int. J. Wine Res., 1: 85-94.
- 17. Steel, R.G.D. and J.H. Torrie, 1991. Principles and Procedures of Statistics: A Biometrical Approach. 2nd Edn., Scholastic Press, Jakarta.
- Omwango, E.O., E.N.M. Njagi, G.O. Orinda and R.N. Wanjau, 2013. Nutrient enrichment of pineapple waste using *Aspergillus niger* and *Trichoderma viride* by solid state fermentation. Afr. J. Biotechnol., 12: 6193-6196.
- 19. Ezekiel, O.O. and O.C. Aworh, 2013. Solid state fermentation of cassava peel with *Trichoderma viride* (ATCC 36316) for protein enrichment. Int. J. Biol. Food Vet. Agric. Eng., 7:667-674.
- Kuswardani, I. and A.I. Wijajaseputra, 1998. Production of single-cell phanerochaete chrysosporium proteins on tofu-enriched wastewater media: Study on optimizing harvest time. Proceedings of the National Seminar on Food and Nutrition Technology, (FNT'98), Yogyakarta, Indonesia.

- 21. Kustyawati, M.E., M. Sari and T. Haryati, 2013. Effect of fermentation using *Saccharomyces cerevisiae* on the biochemical properties of Tapioca. Agritech, 33: 281-287.
- 22. Hatta, U., O. Sjofjan, I. Subagiyo and B. Sundu, 2014. Effects of fermentation by *Trichoderma viride* on nutritive value of copra meal, cellulase activity and performance of broiler chickens. Livest. Res. Dev., Vol. 26, No. 4.
- 23. Nsereko, V.L., K.A. Beauchemin, D.P. Morgavi, L.M. Rode and A.F. Furtado *et al.*, 2002. Effect of a fibrolytic enzyme preparation from *Trichoderma longibrachiatum* on the rumen microbial population of dairy cows. Can. J. Microbiol., 48: 14-20.
- 24. Shaikh, A., A.A. Kathe and V. Mageshwaran, 2014. Reduction of gossypol and increase in crude protein level of cottonseed cake using mixed culture fermentation. Asia-Pac. J. Sci. Technol., 19: 67-73.
- 25. Azia, A., R. Levy, R. Unger, M. Edelman and V. Sobolev, 2015. Genome wide computational determination of the human metalloproteome. Proteins: Struct. Funct. Bioinform., 83: 931-939.
- Strnadova, P., V. Svobodova, L. Pavlata, L. Misurova and R. Dvorak, 2011. Effect of inorganic and organic zinc supplementation on coccidial infections in goat kids. Acta Vet. Brno, 80: 131-137.
- 27. Schneider, T., D.P. Persson, S. Husted, M. Schellenberg and P. Gehrig *et al.*, 2013. A proteomics approach to investigate the process of Zn hyperaccumulation in *Noccaea caerulescens* (J and C. Presl) F.K. Meyer. Plant J., 73: 131-142.