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# Biochemical Effects of Short-term Cadmium Exposure on the Freshwater Fish, *Oreochromis niloticus*

Atef M. Al-Attar

Department of Biology, Teachers College, P.O. Box 2375, Dammam 31451, Saudi Arabia

Abstract: The influences of cadmium, a well known environmental pollutant, on different biochemical parameters in fresh water fish, *Oreochromis niloticus* exposed to sublethal concentration of 9.3 ppm, 0.5 of 96 h LC<sub>50</sub>, for 1, 4 and 7 days were investigated. Significant increases in the levels of blood glucose and the activities of glutamic pyruvic acid transaminase, glutamic oxaloacetic acid transaminase and alkaline phosphatase were observed in the fish treated with cadmium during all experimental periods. The levels of blood triglycerides and total protein were statistically elevated at second and last periods. Insignificant alterations in the levels of blood cholesterol were noted. The observed hyperglycaemia induced by cadmium might be explained in part by increasing rate of glycogenolysis or gluconeogenesis. The observed hypertriglyceridaemia and the elevations of glutamic pyruvic acid transaminase, glutamic oxaloacetic acid transaminase and alkaline phosphatase may be due to liver dysfunction. The observed hyperproteinaemia in the fish, *Oreochromis niloticus* following cadmium administration is possibly attribute to disorder of protein metabolism. However, the significance of the toxic effects of sublethal concentration of cadmium on the various biochemical parameters studied is discussed.

Key words: Biochemical parameters, blood, cadmium, Oreochromis niloticus, toxicity

#### INTRODUCTION

Heavy metals are recognized as one of the most hazardous environmental pollutants and they toxic to many living organisms. Environmental exposure to heavy metals has been reported to cause disease in human and other mammals[1]. In recent years there has been a growing interest about the effects of heavy metals on the health of fish. This aspect is of great importance with development of fisheries, which are often located in rivers, ponds or estuaries subjected to industrialization or mining practices<sup>[2]</sup>. Contamination of freshwater with heavy metals causes devastating effects on ecological balance of the aquatic environments. The diversity of aquatic organisms becomes limited with the extent of pollution. Cadmium is among the heavy metals most toxic to many aquatic organisms<sup>[3-5]</sup>. Also, cadmium is a carcinogenic metal<sup>[6-8]</sup>. Cadmium effects have been already recorded as affecting different metabolic and physiological variables[9-14].

The aim of this study was to investigate the effect of exposure to a sublethal concentration of cadmium on the blood of adult Tilapia, *Oreochromis niloticus* during different time periods from 1 to 7 days. The Tilapia was

chosen as an experimental animal on the basis of some important criteria. The organism is representative of an ecologically important group. Moreover, it is widely edible and available, is amenable to laboratory testing, easily maintained and genetically stable so uniform populations can be tested. In addition, other purposes including type of test, sensitivity to the pollutants and consistency in response to the pollutants were taken into consideration.

## MATERIALS AND METHODS

**Experimental animals:** Adult healthy specimens of *Oreochromis niloticus* (200 $\pm$ 10.2 g weight) were obtained from a local hatchery. They were adapted to laboratory conditions for at least 2 weeks in several 200 L glass aquaria containing well aerated tap water (pH = 7.1 $\pm$ 0.1, hardness as CaCO $_3$  = 94 $\pm$ 2 ppm, oxygen concentration=8.9 $\pm$ 0.1 ppm and temperature = 22 $\pm$ 1 °C). Thereafter, groups of 10 fish each were also adapted for 1 week in 100 L experimental glass aquaria. During the adaptation period they were fed 3 times daily on commercial fish pellets (35% protein) and they were fasted for 24 h prior to any experiment.

Corresponding Author: Dr. Atef M. Al-Attar, Department of Biology, Teachers College, P.O. Box 2375,

Dammam 31451, Saudi Arabia

Tel: +096 650 4629915 E-mail: atef a 2000@yahoo.com

**Toxicity bioassay:** After adaptation period, preliminary experiments were conducted to determine the median lethal concentration at 96 h (96 h LC<sub>50</sub>) for cadmium. Various concentrations of cadmium as Cd (NO<sub>3</sub>)<sub>2</sub>.4H<sub>2</sub>O (10, 12, 14, 16, 18, 20, 22 and 24 ppm Cd) were prepared in eight 100 L glass aquaria. Groups of 10 fish were exposed to each test concentration of cadmium for 96 h. The ninth group served as control. The exposure and control tests were replicated twice. The median lethal concentration at 96 h (96 h LC<sub>50</sub>) was calculated according to the method of the American Public Health Association<sup>[15]</sup> and was found to be 18.6 ppm.

Cadmium exposure: To trace the effects of cadmium on blood, three groups of 10 fish each were exposed to 9.3 ppm of cadmium, 0.5 of the calculated 96 h LC<sub>50</sub>, in 100 L glass aquaria. Fish were sampled for biochemical investigations at 1, 4 and 7 days following exposure to cadmium. Three groups of control fish in tap water were sampled for each specified period. During the experimental periods the test medium was daily renewed to sustain nominal concentration of cadmium.

Biochemical measurements: For blood analysis, the caudal peduncle was cut, blood was collected in non-heparinized tubes. The blood immediately centrifuged at 1500 rpm for 10 min. Serum was then removed and stored at 4°C prior to immediate determination of biochemical parameters, glucose, cholesterol, triglycerides, total protein, glutamic pyruvic acid transaminase (GPT), glutamic oxaloacetic acid transaminase (GOT) and alkaline phosphatase (ALP). Blood glucose was estimated using the method of Trinder<sup>[16]</sup>. Blood cholesterol was measured according to the procedure of Pearson *et al.*<sup>[17]</sup>. Blood triglycerides was determined using the method of Rice<sup>[18]</sup>. The method of

Lowry *et al.*<sup>[19]</sup> was carried out to determine the value of total protein. The activities of blood GPT and GOT were estimated according to the methods of Reitman and Frankel<sup>[20]</sup>. To determine the activity of blood ALP, Bessey *et al.*<sup>[21]</sup> method was used.

**Statistical analysis:** The significance of differences between control and experimental data was statistically analyzed using the Student's t-test.  $p \le 0.05$  was taken as the level of significance.

### RESULTS AND DISCUSSION

The levels of blood glucose and the activities of GPT, GOT and ALP were significantly increased from the corresponding control values throughout the periods of exposure (Table 1). The blood triglycerides and total protein values were markedly elevated at the second and last periods. Insignificant changes in the levels of blood cholesterol were noted during the periods of cadmium treatment (Table 1).

The present study demonstrated that the fish niloticus sublethal Oreochromis exposed to concentration of cadmium displayed a significant elevation in the level of blood glucose after one day till the end of the experimental period. Similar observations have been reported by Benson et al.[22], Early et al.[23], Fu et al.[24], Partap and Bonga[25], Hontela et al.[26] in fish and rat treated with cadmium. Additionally, significant hyperglycaemia were noted by Nath and Kumar<sup>[27]</sup> in the catfish, Heteropneustes fossilis exposed to nickel, Radhakrishaniah et al.[28] and Van Vuren et al.[29] in the fish, Labeo rohita and Claries gariepinus subjected to copper and James et al.[30] in the teleost, Oreochromis mossambicus exposed to lead.

Table 1: Levels of biochemical parameters (X±SD)in the serum of control (n = 5) and in fish (n = 6) exposed to 9.3 ppm cadmium

Parameters	Treatment	Exposure period (day)		
		1	4	 7
Glucose (mg 100 mL <sup>-1</sup> )	С	82.80±1.68	85.26±1.44	100.26±1.35
	T	347.76±2.46*	$158.94\pm1.30^{*}$	155.17±2.23*
Cholesterol (mg 100 mL <sup>-1</sup> )	C	162.24±1.18	169.65±1.85	184.32±1.53
	Т	161.85±1.13	170.82±1.84	186.03±1.75
Trigly cerides (mg 100 mL <sup>-1</sup> )	Ĉ	197.67±2.07	204.16±1.52	190.96±1.60
	T	196.85±1.11	336.16±1.69*	217.04±1.52*
Total protein (g 100 mL <sup>-1</sup> )	C	2.73±0.20	2.81±0.06	2.53±0.07
	T	2.84±0.06	$3.11\pm0.13^*$	3.05±0.07*
GPT (U L <sup>-1</sup> )	C	51.40±2.30	57.60±3.54	48.60±2.79
	T	63.33±2.42*	85.67±2.25*	152.13±3.31*
GOT (U L <sup>-1</sup> )	C	189.40±3.89	175.80±4.64	160.60±6.11
	T	$320.83\pm4.31^*$	342.50±3.08*	355.33±3.62*
$ALP (U L^{-1})$	C	11.60±1.82	12.80±0.84	16.80±3.58
	T	$14.33\pm1.07^*$	$16.17\pm1.60^*$	26.17±1.47*

 $C \ indicates \ Controls, \ T \ indicates \ Treated, \ * \ Statistically \ significant \ at \ p \leq 0.05, \ Student's \ t-test$ 

Wedemeyer and McLeay<sup>[31]</sup> stated that high levels of blood glucose are caused by disorders in carbohydrate metabolism appearing in the condition of physical and chemical stresses. A variety of stressors stimulate the adrenal tissue, resulting in increased level of circulating glucocorticoids<sup>[25,26,32-34]</sup> and catecholamines <sup>[35-37]</sup>Both of these groups of hormones produce hyperglycaemic. Several authors [30,38-42] observed significant decreases in the levels of muscle and/or liver glycogen with significant hyperglycaemia in fish exposed to cadmium, lead and chromium. Förlin et al.[43] demonstrated that the apparent glycogen depletion in trout hepatocytes following exposure to cadmium may reflect an accelerate rate of glycolysis, which would deplete the liver glycogen stores. The decrease in liver glycogen is due to glycogenolysis and normally the decrease of tissue glycogen content runs parallel to a elevation of glucose in serum<sup>[26,30,32]</sup>. Additionally, Kozaric et al.[44] reported that the liver glycogen content of carp (Cyprinus carpio) proved significantly decreased, but the activities of hexokinase and glucose phosphate isomerase were elevated. They suggested an intensive glycogenolysis in cadmiumintoxicated carp. Also, Sastry and Shukla<sup>[45]</sup> stated that the activities of hexokinase, glucose-6-phosphataes, lactate dehydrogenase, succinate and malate dehydrogenases decreased in the liver of teleost fish, Channa punctatus exposed to cadmium. They suggested that glycolysis and gluconeogenesis were impaired in liver. It can not excluded that the possibility that cadmium exert a reducer effect on the content of liver and muscle glycogen in Oreochromis niloticus. However, the elevation of blood glucose is a response to the increased rate of glycogenolysis or gluconeogenesis.

Exposure of cadmium induced a significant rise in blood triglycerides level in the experimental animals, *Oreochromis niloticus*. Several studies demonstrated that the levels of blood triglycerides were significantly increased in the animals treated with different metals<sup>[46,47]</sup>. There are a number of reports indicating that pollutants (e.g., cadmium, lead and mercury)influence thyroid function<sup>[26,34,48-50]</sup>. Since decreased thyroid secretion (hypothyroidism) greatly increases triglycerides level in the blood, the observed hypertriglyceridaemia may be due to hypothyroidism induced by cadmium and/or liver dysfunction because the liver is the principle center of lipid metabolism<sup>[51]</sup>.

The observed hyperproteinaemia in the fish, *Oreochromis niloticus* following cadmium administration is possibly due either to (1) water loss in the serum, (2) the relative changes in the mobilization of blood protein, or (3) elevated *de novo* synthesis. These findings and

explanation are comparable to those considered by Hilmy *et al.*<sup>[52]</sup>, Ruparelia *et al.*<sup>[53]</sup> and Gopal *et al.*<sup>[54]</sup> on toad and fish exposed to cadmium, lead, mercury, copper and nickel.

The data of the present study showed that the exposure of cadmium caused significant elevations in the activities of blood GPT, GOT and ALP. Several investigations showed that these blood enzymes were highly increased in the fish treated with cadmium, zinc and copper<sup>[22,46,55,56]</sup>. Shakoori et al. [57] reported that the increase of blood enzymatic activity is either due to (1) leakage of these enzymes from hepatic cells and thus raising levels in blood, (2) increased synthesis and (3) enzyme induction of these enzymes. Also, Tietz<sup>[51]</sup> and Campbell et al.[58] reported that these enzymes liberate to the blood stream when the hepatic parenchyma cells are damaged. Förlin et al.[43] and Thophon et al.[59] found structural and ultrastructural damage in the liver of rainbow trout and white seabass following cadmium exposure. Moreover, Shimizu and Morita[60], Habeebu-Sultan et al.[61] demonstrated that cadmium induced hepatic necrosis in rats and mice. Finally, De Smet and Bulst<sup>[62]</sup> reported that the concentrations of free amino acids and the activities of proteases were increased in gill, liver and kidney of carp, Cyprinus carpio exposed to cadmium. Also, they observed an increase in the activities of GOT and GPT and they suggested that the observed proteolysis is intended to increase the role of protein in the energy production during cadmium stress.

The results indicated that cadmium produces severe toxic effects in fish blood. Also, *O. niloticus* is incapable of adapting to changes in the water quality caused by the addition of sublethal concentration of cadmium. Further studies are required to investigate the influences of chronic exposure to cadmium.

#### REFERENCES

- Friberg, L.T., M. Piscator, G. Nordberg and T. Kjellstrom, 1986. Handbook on the Toxicology of Metal. Vol. II. Amsterdam.
- Allen, P.C., 1993. Effects of acute exposure to cadmium (II) chloride and lead (II) chloride on the haematological profile of *Oreochromis aureus* (Steindachner). Comp. Biochem. Physiol., 105: 213-217.
- Jackim, E., J. M. Hamlin and S. Sonis, 1970. Effects of metal poisoning on five liver enzymes in the killifish (*Fundulus heteroclitus*). J. Fish. Res. Bd. Can., 27: 383-390.

- Waldichuck, M., 1974. Some Biological Concerns in Heavy Metals Pollution. In: Pollution and Physiology of Marine Organisms. Edited by Vernberg, F.J., Vernberg, W.B., Academic Press, New York, pp: 1-57.
- Ruparelia, S.G., Y. Verma, S.R. Saiyed and U.M. Rawal, 1990. Effect of cadmium on blood of Tilapia, *Oreochromis mossambicus* (Peters), during prolonged exposure. Bull. Environ. Toxicol., 45: 305-312.
- Shiraishi, N., J.F. Hochadel, T.P. Coogan, J. Koropatnick and M.P. Waalkes, 1995. Sensitivity to cadmium-induced genotoxicity in rat testicular cells is associated with minimal expression of the metallothionein gene. Toxicol. Applied Pharmacol., 130: 229-236.
- Ding, W. and D.M. Templeton, 2000. Activation of parallel mitogen-activated protein kinase cascades and induction of c-fos by cadmuim. Toxicol. Applied Pharmacol., 162: 93-99.
- Faverney, C. R-d., A. Devaux, M. Lafauire, J.P. Girad, B. Bailly and R. Rahmani, 2001. Cadmium induceds apoptosis and genotoxicity in rainbow trout hepatocytes through generation of reactive oxygen species. Aquat. Toxicol., 53: 65-76.
- Houston, A.H. and J.T. Keen, 1984. Cadmium inhibition erythropoisis in gold fish *Carassius* auratus. Can. J. Fish Aquat. Sci., 41: 1829-1834.
- Karlsson-Norrgren, L. and P. Runn, 1985. Cadmium dynamics in fish: Pulse studies with 109 Cd in female zebrafish *Brachidanio rerio*. J. Fish Biol., 27: 577-581.
- Hollis, L., J.C. McGeer, D.G. McDonald and C.M. Wood, 1999. Cadmium accumulation, gill Cd binding, acclimation and physiological effects during long term sublethal Cd exposure in rainbow trout. Aquat. Toxicol., 46: 101-119.
- Lionetto, M.G., M.E. Giordano, S. Vilella and T. Schettino, 2000. Inhibition of eel enzymatic activities by cadmium. Aquat. Toxicol., 48: 561-571.
- Belyaeva, E.A. and S.M. Korotkov, 2003. Mechanism of primary Cd<sup>2+</sup>-induced rat liver mitochondria dysfunction: discrete modes of Cd<sup>2+</sup> action on calcium and thiol-dependent domanis. Toxicol. Applied Pharmacol., 192: 56-68.
- Santos, F.W., T. Oro, G. Zeni, J.B. Rocha, P.C. do Nascimento and C.W. Nogueira, 2004. Cadmium induced testicular damage and its response to administration of succimer and diphenyl diselenide in mice. Toxicol. Lett., 152: 255-263.

- American Public Health Association.1971. Standard Methods for the Examination of Water and Waste Water. New York, pp. 874.
- 16. Trinder, P., 1969. Determination of blood glucose using 4-aminophenazone. J. Clin. Pathol., 22: 246.
- Pearson, S., S. Stern and T.H. Mcgavack, 1953. A rapid procedure for the determination of serum cholesterol. J. Clin. Endocrinol., 25: 8-13.
- Rice, E., 1970. Triglycerides (Neutral Fat) in Serum. Stand. Meth. Clin. Chem. Vol. 6, Academic Press, New York, pp. 215.
- Lowery, O.H., N.J. Rosebrough, A.L. Farr and R.J. Randall, 1951. Protein measurement with folin phenol reagent. J. Biol. Chem., 193: 265-275.
- Reitman, P.M. and S.A. Frankel, 1957. Colorimetric method for the determination of serum glutamic oxaloacetic and glutamic pyruvic transaminase. Am. J. Clin. Pathol., 28: 56-63.
- Bessey, O.A., O.H. Lowery and M.J. Brock, 1946. A method for the rapid determination of alkaline phosphatase with five cubic millimeters of serum. J. Biol. Chem., 164: 321-329.
- Benson, W.H., K.N. Baer, R.A. Stac Khouse and C.F. Watson, 1987. Influence of cadmium exposure on selected hematological parameters in freshwater teleost, *Notemigonus chrysoleucas*. Ecotoxicol. Saf., 13: 92-96.
- Early, J.L., V.K. Nonavinakere and Z. Mallory, 1990.
   Effect of cadmium on blood glucose level in the rat. Toxicol. Lett., 54: 199-205.
- 24. Fu, H., O.M. Steinebach, C.J.A. van-den-Hamer, P.H.M. Balm and R.A.C. Lock, 1990. Involvement of cortisol and metallothionein-like proteins in the physiological responses of tilapia (*Oreochromis* mossambicus) to sublethal cadmium stress. Aquat. Toxicol., 16: 257-270.
- Partap, H.P. and S.E.W. Bonga, 1990. Effect of waterbome cadmium on plasma cortisol and glucose in the cichlid fish *Oreochromis mossambicus*. Comp. Biochem. Physiol., 95: 313-317.
- Hontela, A., C. Daniel and A.C. Ricard, 1996. Effect
  of acute and subacute exposure to cadmium on the
  interrenal and thyroid function in rainbow trout,
  Oncorhyncus mykiss. Aquat. Toxicol., 35: 171-182.
- Nath, K. and N. Kumar, 1988. Hyperglycemic response of *Heteropneustes fossilis* exposed to nickel. Acta Hydrochem. Hydrobiol., 16: 333-336.
- Radhakrishnaiah, K., Venkataramana, P. Suresh and B. Sivaramakrishna, 1992. Effects of lethal and sublethal concentrations of copper on glycolysis in liver and muscle of the freshwater teleost, *Labeo rohita* (Hamilton). J. Environ. Biol., 13: 63-68.

- Van Vuren, J.H.J., M. Van Der Merwe and H.H. Du Preez, 1994. The effect of copper on the blood chemistry of *Clarias gariepinus* (Clariidae). Ecotoxicol. Environ., Saf., 29: 187-199.
- James, R., K. Sampath and S. Alagurathinam, 1992. Effects of lead on respiratory enzyme activity, glycogen and blood sugar levels of the teleost *Oreochromis mossambicus* (Peters) during accumulation and depuration. Asian Fish Sci., 9: 87-100.
- Wedemeyer, G.A. and D.J. McLeay, 1981. Methods for Determining the Tolerance of Fishes to Environmental Stressors. In: Stress and Fish. Edited by Pickering, A.D. Academic press, London, pp: 247-275.
- Gluth, G. and W. Hanke, 1985. A comparison of physiological changes in carp, *Cyprinus carpio*, induced by several pollutants at sublethal concentrations. I. The dependency on exposure time. Ecotoxicol. Environ. Saf., 9: 179-188.
- Tort, L., B. Kargacin, P. Torres, M. Giralt and J. Hidalgo, 1996. The effect of cadmium exposure and stress on plasma cortisol, metallothionien levels and oxidative status in rainbow trout (*Oncorhynchus mykiss*) liver. Comp. Biochem. Physiol., 114: 29-34.
- Ricard, A.C., C. Daniel, P. Anderson and A. Hontela, 1998. Effects of subchronic exposure to cadmium chloride on endocrine and metabolic functions in rainbow trout, *Oncorhynchus mykiss*. Arch. Environ. Contam. Toxicol., 34: 377-381.
- Nakano, T. and N. Tomlinson, 1967. Catecholamines and carbohydrate concentration in rainbow trout Salmo gairdneri in relation to physical disturbance.
   J. Fish. Res. Bd. Can., 24: 1701-1715.
- Mazeaud, M.M. and F. Mazeaud, 1981. Adrenergic Responses to Stress in Fish. In: Stress and Fish. Edited by Pickering, A.D., Academic Press, New York, pp. 49-55.
- Witters, H.E., S. Van Puymbroeck and O.L.J. Vanderborght, 1991. Adrenergic response to physiological disturbances in rainbow trout, *Oncorhynchus mykiss*, exposed to aluminum at acid pH. Can. J. Fish Aquat. Sci., 48: 414-420.
- Larsson, A. and C. Haux, 1982. Altered carbohydrate metabolism in fish exposed to sublethal levels of cadmium. J. Environ. Biol., 3: 71-81.
- Sastry, K.V. and S. Tyagi, 1982. Toxic effects of chromium in a freshwater teleost fish, *Channa* punctatus. Toxicol. Lett., 11: 17-21.
- Sastry, K.V. and K. Subharda, 1985. In vivo effects of cadmium on some enzyme activities in tissues of the freshwater catfish, *Heteropneustes fossil*. Environ. Res., 36: 32-45.

- 41. Tewari, H., T.S. Gill and J. Pant, 1987. Impact of chronic lead poisoning on the hematological and biochemical profiles of a fish, *Barbus conchonius*. Bull. Environ. Contam. Toxicol., 38: 748-752.
- 42. Vijayram, K., P. Geraldine, T.S. Varadarajan, G. John and P. Loganathan, 1989. Cadmium induced changes in the biochemistry of an air breathing fish *Anabs testudineus*. J. Ecobiol., 1: 245-251.
- Förlin, L., C. Haux, L. Karlsson-Norrgern, P. Runn and A. Larsson, 1986. Biotransformation enzymes activities and histopathology in rainbow trout, *Salmo gairdnenri*, treated with cadmium. Aquat. Toxicol., 8: 51-64.
- 44. Kozaric, Z., R. Novak and E. Srebocan, 1992. The effect of sublethal concentrations of cadmium on the glycogen content and activities of some enzymes of glucose metabolism in the liver of carp (*Cyprinus* carpio). Verterinarski Archiv., 62: 335-340.
- 45. Sastry, K.V. and V. Shukla, 1994. Acute and chronic toxic effects of cadmium on some haematological, biochemical and enzymological parameters in the freshwater teleost fish *Channa punctatus*. Acta Hydrochim. Hydrobiol., 22: 171-176.
- 46. Singh, H.S. and T.V. Reddy, 1990. Effect of copper sulfate on hematology, blood chemistry and hepatosomatic index of an Indian catfish, *Heteropneustes fossilis* (Bloch) and its recovery. Ecotoxicol. Environ. Saf., 20: 30-35.
- 47. Kumar, A. and R. Barthwal, 1991. Hexavalent chromium effects on hematological indices in rats. Bull. Environ. Contam. Toxicol., 46: 761-768.
- 48. Kawada, J., M. Nishida, Y. Yoshimura and K. Mitami, 1980. Effects of organic and inorganic mercurials on thyroid function. J. Pharm. Biodyn., 3: 149-159.
- Bhattacharya, T., S. Bhattacharya, A.K. Ray and S. Dey, 1989. Influence of industrial pollutants on thyroid function in *Channa punctatus* (Bloch). Ind. J. Exp. Biol., 27: 65-68.
- Kirubagaran, R. and K.P. Joy, 1994. Effects of short-term exposure to methylmercury chloride and its withdrawal on serum levels of thyroid hormones in the catfish *Clarias batrachus*. Bull. Environ. Contam. Toxicol., 53: 166-170.
- 51. Tietz, N.W.,1987. Fundamentals of Clinical Chemistry. Saunders WB Co.
- 52. Hilmy, A.M., N. El-Domaity and M.Y. Daabees, 1986. The use of the chelating agent EDTA in the treatment of acute cadmium toxicity, tissue distribution and some blood parameters in the Egyptian toad *Bufo regularis*. Comp. Biochem. Physiol., 85: 67-74.

- Ruparelia, S.G., Y. Verma, N.S. Mehta and S.R. Saiyed, 1989. Lead-induced biochemical changes in freshwater fish, *Oreochromis* mossambicus. Bull. Environ. Contam. Toxicol., 43:310-314.
- 54. Gopal, V., S. Paravathy and P.R. Balasubramanian, 1997. Effect of heavy metals on the blood protein biochemistry of the fish *Cyprinus carpio* and its use as a bio-indicator of pollution stress. Environ. Monitor. and Assess., 48: 117-124.
- Hilmy, A.M., N. El-Domaity, A.Y. Daabees and H.A. Abdel Latif, 1987. Some physiological and biochemical indices of zinc toxicity in two freshwater fishes, *Clarias lazera* and *Tilapia zillii*. Comp. Biochem. Physiol., 87: 297-301.
- Karan, V., S. Vitorovic, V. Tutundzic and V. Poleksic, 1998. Functional enzyme activity and gill histology of carp after copper sulfate exposure and recovery. Ecotoxicol. Environ. Saf., 40: 49-55.
- Shakoori, A.R., J. Alam, F. Aziz and S.S. Ali, 1990.
   Biochemical effects of bifenthrin (talstar) administered orally for one month on the blood and liver of rabbit. Proc. Pak. Congr. Zool., 10: 61-81.

- Campbell, E.J., C.J. Dickinson, J.D. Spater, C.W., Edwards and K. Sikora, 1984. Clinical Physiology. Bulter and Tanner Ltd. London.
- Thophon, S., M. Kruatrachue, E.S. Upatham,
   P. Pokethitiyook, S. Sahaphong and S. Jaritkhuan,
   2003. Histopathological alterations of white seabass,
   Lates calcarifer, in acute and subchronic cadmium exposure. Environ. Pollut., 121: 307-320.
- Shimizu, M. and S. Morita, 1990. Effects of fasting on cadmium toxicity, glutathione metabolism and metallothionein synthesis in rats. Toxicol. Applied Pharmacol., 103:28-39.
- Habeebu-Sultan, S.M., Liu-Jie and D. Klaassen-Curtis, 1998. Cadmium-induced apoptosis in mouse liver. Toxicol. Applied Pharmacol., 149: 203-209.
- De Smet, H. and R. Blust, 2000. Stress responses and changes in protein metabolism in carp, *Cyprinus carpio* during cadmium exposure. Ecotoxicol. Environ. Saf., 48: 255-26.