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**Sequence Analysis of the Ribosomal DNA ITS2 Region
for *Phlebotomus papatasi* (Diptera: Psychodidae)**

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Abstract: We investigated the heterogeneity of the ribosomal internal transcribed spacer 2 (ITS2) within 131 individuals of *Phlebotomus papatasi* from 25 geographic localities in 10 countries by sequencing the PCR-amplified copies of this spacer. Sequence heterogeneity was observed in both sexes from all localities. There are many microsatellite sequences that show insertion-deletion polymorphisms. The length of the ITS2 ranged from 340 to 512 bp and the average GC content of the ITS2 was 29.2%. The phylogenetic relationships among the members of *P. papatasi* were inferred by neighbour-joining analysis. Although the relationships among *P. papatasi* populations could not be completely resolved, the phylogenies were in good agreement with previous enzyme electrophoresis studies.

Key words: *Phlebotomus papatasi*, ITS2, rDNA, sand fly

INTRODUCTION

Phlebotomus papatasi (Diptera: Psychodidae, Phlebotominae) is the well-known vector of *Leishmania major*, one of the causative agents of the Old World zoonotic cutaneous leishmaniasis. This sand fly species is responsible for human and animal leishmaniasis in many regions of the world (Killick-Kendrick, 1990). *P. papatasi* has a large geographical distribution; Morocco (Guernaoui *et al.*, 2005), Saudi Arabia (Killick-Kendrick *et al.*, 1985), Iran (Yaghoobi-Ershadi *et al.*, 2005), former U.S.S.R. countries (Perfilev, 1966), Palestine and the Jordan Valley (Sawalha *et al.*, 2003; Schlein *et al.*, 1982). Despite this, its population structure and the evolutionary relationships among different populations are conflicting and not well understood (Esseghir *et al.*, 1997, 2000). A recent study using mitochondrial *cytochrome b* DNA and microsatellite markers (Hamarsheh, unpublished data) provided, however evidence that *P. papatasi* does not represent a single genetically uniform population as previously reported.

The broad utility of ribosomal DNA (rDNA) is because the non-coding spacers evolve faster than the coding regions and because it is present as multiple tandemly repeated copies in the genome (Arnheim, 1983; Hillis and Dixon, 1991). In insects, rDNA consists of tandemly repeated transcriptional units with each unit containing the genes for the 18S, 5.8S and 28S ribosomal RNA (Gerbi, 1985; Hillis and Dixon, 1991). The internal transcribed spacers ITS1 and ITS2 are flanking the 5.8S rRNA gene and separate it from 18S and 28S rRNA genes. An external transcribed spacer at the 5' end of the 18S rRNA gene completes the transcriptional unit. The rDNA arrays vary in copy

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number; there are more than 1000 copies in several *Drosophila* species (Beckingham, 1982). These rDNA repeat units do not evolve independently, but at a relatively homogeneous evolutionary rate within individuals and within the species. Concerted evolution of multigene families within species has resulted in rapidly evolving spacer regions, such as the ITS2 rDNA (Dover and Coen, 1981; Arnheim, 1983; Gerbi, 1985; Tautz *et al.*, 1987), including gene conversion and unequal crossing over (Dover, 1982; Seperack *et al.*, 1988). While the coding regions of the rDNA tend to become highly conserved in evolution, the spacer regions appear relatively free to diverge, even in closely related organisms. The ITS sequences were used extensively for resolving the evolutionary affiliations at different taxonomic levels, including recently diverged taxa, such as sibling species (Collins and Paskewitz, 1996; Xu and Qu, 1997).

Substantial heterogeneity of rDNA spacers has been observed in different species of sand flies (Depaquit *et al.*, 2002, 2000; DiMuccio *et al.*, 2000) and in mosquitoes of the genera *Aedes* and *Culex* (Black *et al.*, 1989; Wesson *et al.*, 1992; Miller *et al.*, 1996).

In the present molecular study, we present the results of the analysis of the ITS2 sequences of 131 individuals of *P. papatasi* originating from 25 different geographical localities in 10 countries and evaluate the usefulness of using ITS2 as molecular marker for *P. papatasi* sand fly.

MATERIALS AND METHODS

Sand Fly Materials

P. papatasi sand flies were caught with CDC miniature light traps or sometimes on sticky papers, or came from laboratory colonies (Table 1).

DNA Extraction

DNA was extracted from individual flies by phenol-chloroform extraction method with slight modifications. Single male flies and the thorax and anterior abdomen of the females were homogenized

Table 1: Samples, locations and collection information (for field flies) for the 25 *P. papatasi* populations included in this study, blank fields indicate that no information was available

Country	Population	Latitude	Longitude	Altitude	Year	Capture method
Syria	Aleppo	36° 12' 10N	37° 9' 31E	422	2004	Manual aspirator
	Raqah	35° 57' 0N	39° 1' 0E	225	2004	Manual aspirator
	Dayr az Zawr	35° 19' 60N	40° 8' 60E	193	2004	Manual aspirator
Turkey	Salniurfa	37° 9' 4N	38° 47' 34E	477		Colony
	Hamdun	37° 29' 44N	39° 07' 8E	644	1999	Colony
	Nizip	37° 35' 10N	38° 56' 34E	400		Colony
Cyprus	Chisochus	40° 37' 83N	46° 86' 96E	63	2004	CDC trap
	Lapta	35° 20' 15N	33° 10' 45E	190		Colony
Jordan	Jordan Valley					Colony
Israel	Jordan Valley	30° 56' 60N	35° 22' 0E	-339	2003	CDC trap
Palestine	Jericho	31° 52' 0N	35° 27' 0E	-227	2004	CDC trap
	Jordan Valley	31° 52' 0N	35° 27' 0E	-227		Colony
	Jordan Valley				2004	CDC trap
	Qabatiyah	32° 24' 31N	35° 16' 48E	284	2004	CDC trap
Egypt	Raba	32° 23' 9N	35° 22' 55E	707	2004	CDC trap
	Dayr al Gousun	32° 21' 9N	35° 4' 28E	234	2004	CDC trap
	N. Sinai	30° 49' 60N	34° 7' 0E	111		Colony
	Sinai	30° 49' 60N	34° 7' 0E	111		Colony
	Kafr ash shaykh	31° 6' 41N	30° 56' 11E	6		Colony
	Alexandria	31° 11' 53N	29° 55' 9E	4		Colony
	Al-Qalyopia	30° 11' 53N	31° 8' 3E	26		Colony
Italy	Rom	41° 53' 60N	12° 28' 60E	14		Colony
	Rocca Priora	41° 47' 60N	12° 45' 0E	686		Colony
Tunisia	Tunis	36° 48' 10N	10° 10' 47E	0	2004	CDC trap
Morocco	Marrakesh	31° 37' 60N	8° 0' 0W	450	2005	CDC trap

with a sterile glass rod in 1.5 mL microfuge tube containing 250 μ L lysis buffer (0.1 M Tris-HCl pH 8, 50 mM EDTA pH 8), 2.5 μ L Triton X100 and 5 μ L Proteinase K. The homogenate was incubated at 60°C overnight, extracted with equilibrated phenol, phenol: chloroform: iso-amyl alcohol in 25:24:1 volume ratio (Roth, Germany), then extracted with chloroform: iso-amyl alcohol in 24:1 volume ratio. For each extraction step, tubes were mixed gently for 10 min, centrifuged using desktop centrifuge at 13000 g for 10 min, then the upper aqueous phase was transferred to a new 1.5 mL microfuge tube. DNA was precipitated with 250 μ L isopropanol and 20 μ L 3M sodium acetate at -20°C overnight. DNA was pelleted by centrifugation at 13000 g for 30 min, the supernatant removed and the pellet washed twice with 500 μ L cold 70% ethanol. DNA pellets were dried using speed-vac centrifuge (Eppendorf, Germany) for 8 min and then dissolved in 50 μ L TE buffer (10 mM Tris-HCl, 0.1 mM EDTA, pH 8). DNA was stored at 4°C.

PCR Amplification

Amplifications of the ITS2 were done using the following conditions: an initial denaturation at 95°C for five min; followed by 35 cycles at 94°C for 30 sec, 62°C for 30 sec and 72°C for 1.5 min and a final extension at 72°C for 10 min. The PCR was performed in 50 μ L reaction volume containing 50 ng genomic DNA, 10 \times PCR buffer (Applied Biosystems) 12 pM of each primer C1a: 5' CCTGGTTAGTTTCTTTTCTCCGCT-3' and JTS3: 5'-CGCAGCTAACTGTGTGAAATC-3' (Depaquit *et al.*, 2000), 2.5 mM of each dNTP, 1.5 mM MgCl₂ and 0.5 units AmpliTaq® DNA polymerase (Applied Biosystems). PCR amplifications were carried out in RoboCycler® Gradient 40 (Stratagene).

SSCP Analysis and Sequencing

Each specimen was screened for polymorphisms by single strand conformation polymorphism (SSCP) analysis. Electrophoresis for SSCP was performed on gels (0.8 mm thick) which consisted of the following: 112.5 mL ddH₂O, 37.5 mL 0.5 \times MDE®(Cambrex, Rockland), 9 mL 10 \times TBE buffer, 0.6 mL 10% ammonium persulfate and 60 μ L TEMED. Fifteen μ L of PCR product was mixed with 2 μ L of denaturing mix (1% SDS, 10 mM EDTA) and 2 μ L loading mix (95% formamide, 10 mM EDTA, 0.025% bromophenol blue and 0.025% Xylene cyanol). The tube was shaken gently, heated at 98°C for 13 min in a water bath and immediately placed on ice for at least 10 min. Sixteen μ L of the sample was loaded onto the SSCP gel. Electrophoresis was carried out on vertical slab gels (45 \times 35 cm) in a cold room (4°C) using 1 \times TBE buffer for 16 h at 1 KV, 280 mA and 10 Watts. The gel was silver stained, fixed using 1% Nitric acid for 15 min and stained with 0.2% silver nitrate for 25 min. After washing in deionized water for 15 min the gel was placed in developer (0.28 M Na₂CO₃ and 0.1% formaldehyde) until bands became visible and dried at 70°C for 2 h using a gel drier.

Samples presented unique SSCP banding profiles were sequenced in both directions using automated sequencing techniques at sequencing facility, Department of Genetics, Humboldt University, Berlin.

Data Analysis

Sequences (481-515 bp) were manually edited and aligned using CLUSTAL W (Higgins *et al.*, 1994). Gaps were treated as missing data. MEGA version 3.0 (Kumar *et al.*, 2004) and DnaSP (Rozas *et al.*, 2003) was used for the statistical analysis of sequences. The sequences were analyzed using neighbour-joining (NJ) (Saitou and Nei, 1987) with Kimura's two-parameter correction (Kimura, 1980), as implemented in the TREECON for Windows v. 1.3b (Van de Peer and De Wachter, 1994); gaps were excluded from the analysis. Bootstrapping (Felsenstein, 1985) was performed using NJ with 1000 replicates. The trees were rooted using the ITS2 sequence of *Phlebotomus sergenti* (GenBank: AF218323) as an out group.

RESULTS

Forty three sequences comprising 5.8S gene, the ITS2 and the 5' end of the 28S ribosomal gene, were generated for four individuals from Cyprus, one from Jordan, one from Morocco, 11 from Egypt, six from Italy, six from the Middle East, eight from Syria and six from Turkey. All sequences were determined and positively confirmed as ITS2 with a BLAST search of GenBank. Sequences are available in GenBank under accession numbers (DQ887632-DQ887674).

The size of the amplified fragments varied considerably among studied individuals; from 340 bp in Syr3T157 to 512 bp in Tuk3T14 with an average of 482.6 bp (Table 1). The sequences were aligned along with the ITS2 sequence for *P. papatasi* from Rhuhaibe, Syria which is available in the GenBank under accession number: AF218321. Much of the variation in length was due to insertion /deletion mutations and differences in the numbers of simple sequence repeats which comprised principally simple di-nucleotide repeat insertions. The overall nucleotide composition (A, T, C and G) of the sand fly populations considered here was 36.1, 34.7, 13.5 and 15.7%, respectively. Motifs containing A and T residues outnumbered most of the permutations and constituted much of the elevated AT content.

Sequence Heterogeneity

The alignment of the ITS2 sequences resulted in a total of 563 characters including gaps (Fig. 1). Of the 563 characters, 137 were variable (polymorphic), 179 were invariable (monomorphic) and 75 were parsimony informative sites. No loci diagnostic for populations were detected for ITS2 and sequences were not identical for individuals originated from the same population.

In sequencing PCR products, sequence heterogeneity was noted in most of individuals. Sequencing was repeated in certain individuals 3-4 times, but without resolving the heterogeneity. Almost all sequenced samples show different levels of sequence heterogeneity even if they are originated from the same population. It seems that there is a difficulty in reading some nucleotides in the same position which is probably due to different ITS2 copies. Observations on the sequence alignment reflected an elevated sequence variation in the 3' region compared to the 5' region.

Phylogenetic Analysis

Aligned complete sequences from each individual were analyzed phylogenetically using NJ analysis; gaps between paired alignments were treated as missing data.

A bootstrap consensus neighbour-joining (NJ) tree produced from Kimura's two-parameter is shown in Fig. 2. Most of the phylogenetically critical nodes were not well supported as can be seen by low bootstrap values (only bootstrap values above 50% were shown). Although the phylogenetical relationships were completely unresolved and did not show significant structuring, a strongly supported microclade (100%) grouped two individuals; E1T53 and E1T11, both derived from Egypt.

PCR-RFLP Assays

To determine whether multiple copies of ITS2 may vary within individuals, we subjected the PCR-amplified copies of this spacer to restriction analysis. The RFLP patterns of the amplified ITS2 obtained with the restriction enzymes *Sau961*, *AvaII* and *RsaI* did not exhibit any variations among the different populations, however the sum of all restricted fragments produced from all restricted enzymes exceed the maximum size of the non-restricted PCR product which probably indicates the presence of many different ITS2 copies (Fig. 3).

AF218321	2222222222	2222222222	2222222222	2222222223	3333333333	3333333333	3333333333	3333333333	3333333333	3333333333	3333333333	3333333333	3333333333	3333333333	3333333333	3333333333	3333333333	3333333333	3333333333	
CypT20	6666666667	7777777778	8888888889	9999999990	0000000001	1111111112	2222222223	3333333334	4444444445	5555555556	6666666667	7777777778	8888888889	9999999990	0000000001	1111111112	2222222223	3333333334	4444444445	
CypT34	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	1234567890	
CypT48	GAATGTACCC	AA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCA	TTGG--ATAAT	TTATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Egt5T141	GAATGTACCC	AA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCA	TTGG--ATAAT	TTATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Egy1T11	TAATGGTCCC	CA--AGGATA	AAATA--ATA	AAAATTGAAA	TAAATATTTAA	--AAGGGGAA	AATAAAATTA	TTGG--ATAAA	TTAATTTAAA	AACCAATTGG	TTTTCCAAAA	AACCTT--TTTT	TTTTAAAAAA							
Egy1T39	GAATGTACCC	AA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCA	TTGG--ATAAT	TTATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Egy1T53	TAATGGTCCC	CA--AGGTTA	AAAT----TA	AAAATTGAAA	TATTTTTTTT	--AAGGGGAT	AATAAAATTA	TTGG--ATAAA	TTAATTTAAA	AAACAATTGG	TTTTCCAAAA	AACCTT--ATTA	TTTTAAAAAA							
Egy2T26	GAATGTACCC	AA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCA	TTGG--ATAAT	TTATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Egy2T40	GAATGTGCCC	AA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCA	TTGG--GTAAT	TTATCTTAAA	GACATATTGT	TACACAGAAA	AACCTT--ATTA	TAATAAAAAA							
Egy2T54	TAATGTGCCC	CATGTG--TA	AAA----TTA	AAAGTTGAAA	AAATATATATA	--AGGGGGGT	ATCATATTCCT	TTGG--GTAAT	TTATCTTAAA	AACATATTGT	TTTACACAAA	AACCTT--TTTT	TATTTAAAAA							
Egy3T41	GAATGTACCC	AAG--TGG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	TAGGGGGTAT	ATCATAGTCT	GTGT--AAAAT	ATATCATAAA	CACATATTGT	TATACAGAAA	AACCTT--TTTT	ATTTTAAAAA							
Egy3T69	GAATGTACCC	AA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCC	TTGG--ATAAT	TTATCTTAAA	GACCTATTGT	TATAAAAAAA	AACCTT--ATTT	TTTTTAAAAA							
Egy4T28	GATGCCCC	CCC--TGATA	TATAAAAGA	AAAGTAAAAA	TATATGGGGG	--GGGGTATT	ATCATAGTTA	TTGG--ATAAT	TTATCTTAAA	GACATGTTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Egy4T70	ATAAGAGCCG	GG--AAACTT	TAAT--TGG	GTTTTTAAAA	AAATTTTTTT	AGAGGGGCGA	AAATTTTTTT	CCACCCCTCC	AAAGTGTAAA	AAAA--TGT	TAAAAAAA	AAATTTTTTT	TTTTTAAAAA							
Ity1T22	GAATGTACCC	CAATG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AAGGGGGT	ATCATATTTCA	TTCC--AAAAT	TCCAAGTTTT	TATACAAAGG	TATAAAAAAA	AAATTT--TATT	TGTTTTAAAA							
Ity1T36	GAATGTACCC	AA--TGG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCA	TTGG--ATAAT	TTATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Ity2T37	GATGGACCCC	CC--GG--AT	TAAT--ATTA	AAAGTTGAAA	TATTTTTTTA	--AGGGGGAT	ATAATAATAA	TGGG--TTAAT	TAAATTTAAA	AACAAATTGT	TATAAAAAAA	AAATA--ATTA	TTTTTAAAAA							
Ity2T38	GAATGTGCCC	GA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCA	TTGG--ATAAT	TTATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Ity3T52	GAATGTACCC	CA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGGAT	ATCATATTTCA	TTGG--ATAAT	TTATCTTAAA	AACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Ity3T66	GAATGTACCA	CA--TG--TA	GAAT--ATTA	AAAGTTGAAA	TATATTTTTT	--AGGGGGGT	ATTAATAATA	TTGG--AGAAAT	TTATCTTAAA	AACATATTGT	TATACAGAAA	AAC--AT--A	TTTAAAAA							
ILTjv	GAATGTACCC	AA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCA	TTGG--ATAAT	TTATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
JorT138	GAATGTACCC	CA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGGAT	ATCATATTTCA	TTGG--ATAAT	TTATCTTAAA	AACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Pal1T21	GAATGTACCC	AA--TTG--GA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGGAT	ATCATAGTCA	TTGG--ATAAT	TTATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Pal1T49	GAATGTGCCC	AA--TG--TA	TAAA--ATAA	AAAGTTGAAA	AAATATATATA	--AGGGGTAT	ATCATATTCCT	TTGG--ATAAT	TTATCTTAAA	AACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Pal1T63	AATGGTCCCC	A--TTG--TT	TATT--TTTA	AAGTTTAAAT	TTTTTTTTTT	--AGGGGTAA	ATAA--AAITA	TTGGAATAAT	TAAATTTAAA	AACAAATGGT	TTTTCCAAAA	ACTTT--TTTT	TTTTTAAAAA							
Pal3QT160	AATTTGCCCC	A--TTG--TT	TAAA--ATTA	AAATTTGAAAT	TATTTTTTATA	--AGGGGTAT	ATCA--AAITC	TTGGGATAAT	TTATTTTAAA	AACAAATTGT	TATCCAAAAA	ACCCTT--TTTT	TTTTTAAAAA							
Pal4RT162	TAATGGTCCC	CA--TGG--TA	AAAA--ATAA	AAATTTAAAA	AAATTTTTAAA	--AGGGGAAA	ATAA--AAITC	TTGGGAAATT	TAAATTTAAA	AACATATGGT	TA--ACCAAAA	AACCTT--TTTT	TATTTAAAAA							
syr1T15	TATTTGTCAC	GA--TG--AA	TA--AT--ATTA	AAAGTTAAAA	TATTTTTTATA	--AGCAGTAT	ATAATAGTCC	TTGA--ATAAT	TTATATAAAA	GACTTATTGT	TATAAAAAAA	AACCTT--ATTA	TAAAAAAAT							
syr1T29	GAATGTACCC	AA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCA	TTGG--ATAAT	TTATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
syr2T58	AATGGTCCCC	A--AGG--TT	AAAA--ATAA	AAATTTAAAT	ATTTTTTTTT	--AGGGGATT	ATAA--AAITA	TTTGAATAAT	TAAATTTAAA	AACATTTTGT	TATCCAAAAA	ACCCTT--ATAA	TTTTTAAAAA							
syr3T157	GAAAGTGCCC	AC--TG--TG	TAAT--ATTA	AAAATTGAAA	AAATATATTTA	--AGGGGACC	ACCCCACTCC	CCCGCACAAAT	CTATCTTATA	GACATATTGT	GATATACAAA	AGCTT-----								
syr3T37	GAATGTACCC	AA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATATAATA	TTGG--ATAAT	TTATCTTAAA	AACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
syr3T31	GAATGTACCC	AA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCA	TTGG--ATAAT	TTATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
syr3T45	GAATGTACTC	AA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCA	TTGG--ATAAT	TTATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
syr1T17	GAGGTGCCC	CA--TGG--TA	TAAT--ATAA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCT	TTGT--ATAAT	ATATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	T-----							
Tuk1T151	GAATGTCCCC	AA--TG--TA	TATT--ATTA	AAAGTTGAAA	TATATATATG	--GGGGGTAT	ATCATATTTCC	TTGA--ATAAT	TTATCTTAAA	GACTTATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Tuk1T18	GAATGTCCCC	AAT--TG--GT	TAAT--ATTA	AAAGTTGAAA	ATTTTTTTTT	--AGCGGGAT	ATCAAAAATCA	TTGG--ATAAT	TTATCTTAAA	AACATTTTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Tuk1T32	GAGTG--CCCC	CTG--TG--TA	TAAT--ATAA	AAAGTTGAAA	TATATATATG	--GGGGGTAT	ATCATAGTCT	GTGG--ATAAT	TTATCATAAA	CACATATTGT	TATACAGAAA	AACCTT--ATT-	ATATAAAAAA							
Tuk1T46	GAATGTACCC	AA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCA	TTGG--ATAAT	TTATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Tuk2T33	GAAAGTCCCC	AA--GG--TT	TAAT--TTTT	AAAGTTAAAA	TATTTTTTTA	--AGGGGTAT	ATCAAAATTCCT	TTGG--TAAAT	TTATTTTAAA	AACAAATGGT	TTTTCCAAAA	AACCTT--TTTT	TTTTTAAAAA							
Tuk3T14	GAATGTACCC	AA--TG--TA	TAAT--ATTA	AAAGTTGAAA	TATATATATA	--AGGGGTAT	ATCATAGTCA	TTGG--ATAAT	TTATCTTAAA	GACATATTGT	TATACAGAAA	AACCTT--ATTA	TATTTAAAAA							
Moc1T144	TTTTGTCCCC	CC--CG--TT	TAAT--ATTA	AAAGTTGAAA	AAATTTTTTTA	--AGGGGGGG	ATCATATCT	TTGG--TTAAT	TATTTTTCTT	AACGACTTTT	TTTTACACAA	AACCTA--CTTA	TTTTTAAAAA							

Fig. 1: Continued


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5555555555 5555555555 5555555555 5555555555 555
2222222223 3333333334 4444444445 5555555556 666
1234567890 1234567890 1234567890 1234567890 123
AF218321 *TTTAAGCATA TTAATAAGC- GGAGGAAAAG AA- ACTAACC AGG
CypT20 *TTTAAGCATA ATAATAAGC- GGAGGAAAAA AA- ACTAACC CGG
CypT34 *ATTAACACA TTAATAAAC- GGGGGAAAAA AAAACTACCC CGG
CypT48 *TTTAAGCATA TTAATAAGC- GGAGGAAAAG AA- ACTAACC AGG
CypT6 *TTTAAGCATA TTAATAAGC- GGAGGAAAAG AA- ACTAACC CGG
Egt5T141 *TTTAACCATA TAAAAAACCC GGAGGAAAAA AA- ACTACCC AGG
Egy1T11 *TTTAAGCATA TTAATAAGC- GGAGGAAAAG AA- ACTAACC CGG
Egy1T39 *TTTAAGCCTA TTAATAAGC- GGAGGAAAAG AA- ACTAACC CGG
Egy1T53 *TTTAAGCATA TTAATAAGC- GGAGGAAAAG AA- ACTAACC AGG
Egy2T26 *TTTAAGCATA TTAATAAGC- GGAGGAAAAG AA- ACTAACC GGG
Egy2T40 *TTTTGCCTC CCAATAACCC GAGGGAAAAA AACAAATGCC AGG
Egy2T54 *TTAAGCATT TTTATACGC- GGAGGAAAAG AA- ACTAACC AGG
Egy3T41 *TTTACACATA TAAATCCGC- GGAGGAAAAA AA- ACTAACC AGG
Egy3T69 *AAAATGAATG CTAATTC-CA CATATTTAAA AAAAATACCC AGG
Egy4T28 *TTTAATCATA ATAAGAAGC- GGAGGAAAAA AA- CCCCCCC AGG
Egy4T70 -----
Ity1T22 *TTTAAGCATA TTAATAAGC- GGAGGAAAAG AA- ACTAACC AGG
Ity1T36 *TTTAAGCATA TTAATAAGC- GGGGGAAAAG AA- ACTAACC AGG
Ity2T37 *TTTAAGCATA TTAATAAGC- GGAGGAAAAG AAAAAAACCC GGG
Ity3T38 *TTTAAGCATA TTAATAAGC- GGAGGAAAAA AA- ACTCACC AGG
Ity3T52 *TTTAAGCATA TTAATAAGC- GGAGGAAAAG AA- AAAAACCC AGG
Ity3T66 *TTTAAGCATA ATAATAATC- AGAGGAGAAG AAAAAAACCC AGA
ILTjv *TTTAAGCATA TTAATAAGC- GGAGGAAAAG AA- ACTAACC AGG
JorT138 *GTTTTTCATC TTATTAAGC- GGAGGAGGAG AA- AATACTC AGG
Pal1T21 *TTTAAGCATA TTAATAAGC- GGGGGAAAAA AA- ACTAACC AGG
Pal1T49 *TTTAAACATA TTAATAAGC- GGAGGAAAAA AA- ACTCCCC GGG
Pal1T63 *TTTAAACCTT TTAATTAA-C CGAGGAAAAA AAACCT-ACC CGG
Pal3QT160 *TTTTAACCTT TTTATAAAAC CGCGGAAAAA AAAACACACC CGG
Pal4RT162 *TTC AACCTTA TTAACCCGC- -GGAGAAAAA AAAACTACCC GGG
syr1T15 *TTTAATCATA TTAATAAGC- GGAGGAAAAA AA- ACTAACG NGG
Syr1T29 *TTTAAGCATA TTAATAAGC- GGAGGAAAAG AA- ACTAACC GGG
Syr2T58 *TTTAACCCAT TTAATAAAC- GAGGGAAAAA AAATCT-ACC AGG
Syr3T157 -----
Syr3T3 *TTTAAGCATA TTAATAAGC- GGAGGAAAAA AA- ACTAACC AGG
syr3T31 *TTTAAGCATA TTAATAAGC- GGAAGGAAAA AA- ACTAACC AGG
Syr3T45 *TTTAAGCATA TTAATAAGC- GGAGGAAAAG AA- ACTAACC AGG
SyrT17 *TTTAAGCATA TTAATAAGC- GGAGGAAAAA AA- ACTAACC AGG
Tuk1T151 *TTTAACCTTA TTAATCGGC- GGAGAAAAAA AA- ACTAACC AGG
Tuk1T18 *TTT TAG---T TTTATAAGC- GGGAGAAAAG A---TAACC AGG
Tuk1T32 *TTTAAGCATA TTAATAAGC- GGGAGAAAAA AA- ACTAACC GGG
Tuk1T46 *TTTAAGCATA TTAATAAGC- GGAGGAAAAA AA- ACTACCC GGG
Tuk2T33 *TTTTGGCCCG TTAATAACC- GGAGGAAAAA AA- ATTCACC AGG
Tuk3T14 *TTTAAGCATA TTAATAAGC- GGAGGAAAAG AA- ACTAACC AGG
MocT144 *GTTTAGCATA ATCCTAAGC- GGAGGAAAAG AA- ACTAACC AGG

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Fig. 1: Sequence alignment of the ITS2 and flanking 5.8S and 28S coding regions of the rDNA of 43 *P. papatasi* individuals. Start and end of the presumptive ITS2 region are marked by asterisks. Insertions or deletions (indels) in alignment are denoted by a dash (-). Abbreviations are the same as in Table 1

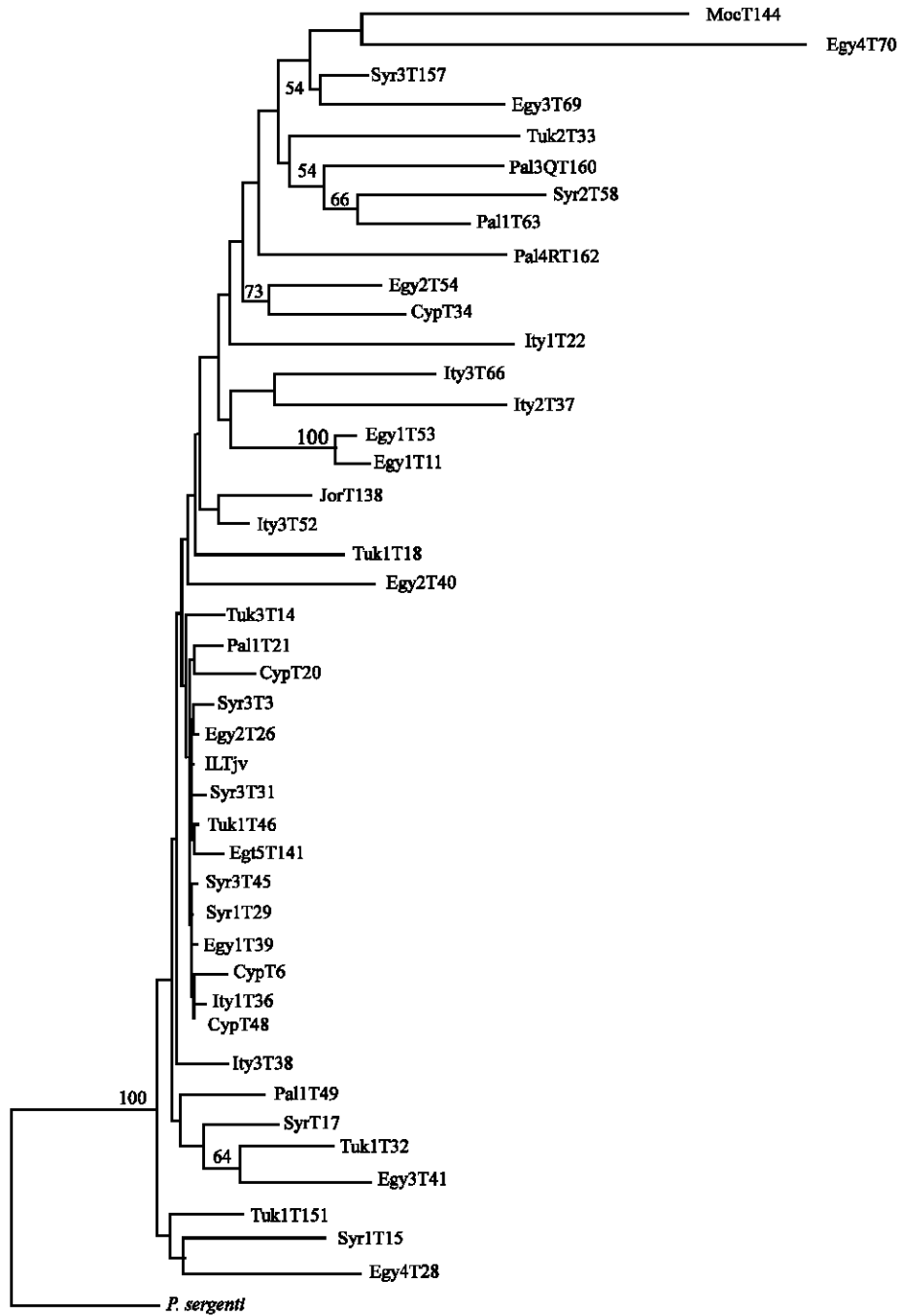


Fig. 2: A neighbour-joining bootstrap tree based on rDNA ITS2 sequence data for 43 samples of *P. papatasi* from 25 different populations originating from 10 countries. The tree was rooted with *P. sergenti* ITS2 sequence (GenBank: AF462332.1). Numbers show bootstrap confidence levels above 50% (from 1000 replications) above the branches tested. Sample codes are the same as in Table 2

Table 2: Length and GC content of ITS2 for the 43 samples sequenced in this study

Sample code	Length (bp)	GC (%)	Accession No.
AF218321	488	28.5	
CypT20	490	28.6	DQ887632
CypT34	491	28.7	DQ887633
CypT48	490	28.4	DQ887634
CypT6	490	28.6	DQ887635
Egt5T141	486	28.8	DQ887636
Egy1T11	493	28.0	DQ887637
Egy1T39	483	29.2	DQ887638
Egy1T53	484	27.9	DQ887639
Egy2T26	487	28.5	DQ887640
Egy2T40	490	30.8	DQ887641
Egy2T54	484	31.0	DQ887642
Egy3T41	495	30.3	DQ887643
Egy3T69	484	27.7	DQ887644
Egy4T28	494	32.0	DQ887645
Egy4T70	419	32.9	DQ887646
ILTjv	485	28.7	DQ887653
Ity1T22	492	29.7	DQ887647
Ity1T36	491	28.9	DQ887648
Ity2T37	483	30.2	DQ887649
Ity3T38	490	29.8	DQ887650
Ity3T52	484	27.9	DQ887651
Ity3T66	485	28.1	DQ887652
JorT138	480	29.6	DQ887654
MocT144	481	33.5	DQ887674
Pal1T21	488	28.9	DQ887655
Pal1T49	487	29.0	DQ887656
Pal1T63	484	27.7	DQ887657
Pal3QT160	486	29.6	DQ887658
Pal4RT162	486	29.2	DQ887659
Syr1T15	486	27.3	DQ887660
Syr1T29	484	28.9	DQ887661
Syr2T58	484	27.9	DQ887662
Syr3T157	340	33.6	DQ887663
Syr3T3	485	27.6	DQ887664
Syr3T31	487	28.3	DQ887665
Syr3T45	483	28.5	DQ887666
SyrT17	485	30.1	DQ887667
Tuk1T151	490	29.4	DQ887668
Tuk1T18	483	28.7	DQ887669
Tuk1T32	490	31.0	DQ887670
Tuk1T46	487	28.7	DQ887671
Tuk2T33	487	29.1	DQ887672
Tuk3T14	512	28.3	DQ887673
Average	482.6	29.2	

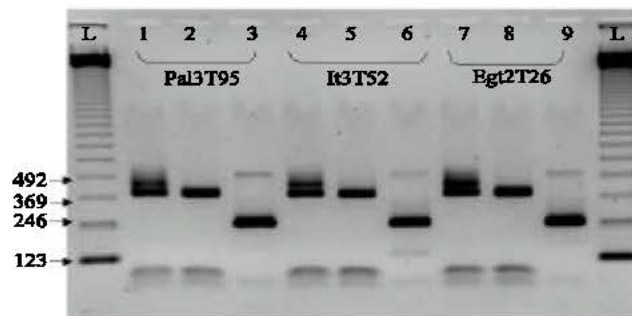


Fig. 3: Polymerase chain reaction restriction fragment length polymorphism patterns for the internal transcribed spacer 2 (ITS2) of three *P. papatasi* individuals; Pal3T95, It3T52 and Egt2T26 after digestion with *Sau961*(lane1-3), *AvaII* (lane 4-6)and *RsaI* (lane 7-9) restriction enzymes

DISCUSSION

The high level of genetic diversity in the ITS2 region of the *P. papatasi* individuals studied here denotes intraspecific variation and even intra-individual variation in the multiple copies of the rDNA subunits. The GC content (29.2%) of the ITS2 sequences obtained fell below the range of the anopheline species (Beebe and Cooper, 2000). The ITS2 region in *P. papatasi* appears to display several AT dinucleotide microsatellite repeats which in part contributes to the polymorphic nature of the ITS2. This could explain the bias toward A and T which was consistent with the base composition of ITS2 sequences of other sand fly species such as *Phlebotomus sergenti*, *Phlebotomus similis* (Depaquite, 2002) and *Lutzomyia shannoni* (Miller *et al.*, 1997).

It seems that variants within individual sand flies often differed as much as those between flies from different localities. This is actually the reason for poor geographical structuring in the NJ tree and also contradicts with our previous studies on *P. papatasi* based on *cytochrome b* gene sequences (data not shown). This is not surprising however, since *cytochrome b* gene is maternally inherited and not recombinant while ITS2 is free to diverge.

At the genomic level, the effectiveness of molecular drive at homogenizing a multigene family depends on the rates of unequal crossing over and gene conversion and is negatively correlated with both mutation rate and number of unlinked loci of the multigene family (Perelson and Bell, 1977). Intra-individual sequence heterogeneity indicated that the process of concerted evolution had not homogenized the ITS2 sequence in some individuals. Perelson and Bell (1977) noted that unequal crossing over increases heterogeneity among copies of a multigene family if mutation rates are high relative to homogenization rates.

Extensive intragenomic variation can confound phylogenetic and population studies based on ribosomal spacers, however it is necessary to first determine the extent of homogenization of a multigene family at several hierarchical levels (within individual, population and species) before employing them in phylogenetic studies (Williams *et al.*, 1988; Sanderson and Doyle, 1992).

The high polymorphism of ribosomal ITS2 in *P. papatasi* is mainly due to insertion/deletion mutations and changes in short tandem repeat sequences. Increases or decreases in the size of ITS2 may have been caused by replication slippage. In this study ITS2 sequences were shown to be too variable for phylogenetic analysis so that genetic relationships between different populations could not completely resolved, this actually makes the use of the current ITS2 sequences of little value for phylogenetic interpretation.

As a conclusion, the ITS2 does not represent a suitable marker for inferring phylogenetic and population genetic relationships across *P. papatasi* sand flies and that other genetic markers are needed for this purpose.

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