

# Journal of Entomology 

ISSN 1812-5670

Academic
Journals Inc.

# Toxicity of Ethanolic Extract of Croton heliotropiifolius in Weevil Populations of Stored Maize Grains 

${ }^{1}$ L.B. Silva, ${ }^{1}$ E.B. Torres, ${ }^{2}$ K.F. Silva, ${ }^{2}$ J.S.N. Souza, ${ }^{2}$ M.S. Lopes, ${ }^{3}$ L.H. Andrade and ${ }^{1}$ Z.F. Xavier<br>${ }^{1}$ Programa de Pos-Graduacao em Agronomia-Fitotecnia-Universidade Federal do Piaui, Campus "Profa. Cinobelina Elvas", Bom Jesus-PI, Brasil<br>${ }^{2}$ Departamento de Ciencias da Natureza-Universidade Federal do Piaui, Campus "Profa Cinobelina Elvas", Bom Jesus-PI, Brasil<br>${ }^{3}$ DCR Scholar-FAPEPI-PI, Brasil<br>Corresponding Author: L.B. Silva, Programa de Pos-Graduacao em Agronomia-Fitotecnia-Universidade Federal do Piaui, Campus "Profa. Cinobelina Elvas", Rodovia Municipal Bom Jesus-Viana, km 01-Bairro Planalto Horizonte, CEP 64900-000, Bom Jesus-PI, Brasil Tel: (89) 3562-2247


#### Abstract

The objective of this study was to evaluate, under laboratory conditions, the contact toxicity of the ethanolic extract of Croton heliotropiifolius on the main pest of stored maize, Sitophilus zeamais Motschulsky. Extract of the leaves of C. heliotropiifolius and maize weevil populations collected in four different storage units were used to determine whether the contact toxicity of ethanolic extract of C. heliotropiifolius in populations of Sitophilus zeamais incurs differentiated responses. After bioassays, we compared the mean mortality between the treatment and control groups. Insects were treated topically with the ethanolic extract of leaves of C. heliotropifolius at a concentration of $10,000 \mathrm{ppm}$ and mortality was assessed 24,48 and 72 h after application. The mortality of S. zeamais adults from the four populations was significantly higher compared with to control in all evaluation periods. The mortality of S. zeamais adults ( $<15$ days-old) topically treated with the extract increased with time of exposure to extract residues. The populations evaluated had different susceptibility in relation to contact with the ethanolic extract studied. According to the results obtained, it was concluded that the ethanolic extract of leaves of C. heliotropiifolius has contact insecticidal activity against adults of Sitophilus zeamais and that the response to this toxicity varies with the geographical origin of the population analyzed.


Key words: Botanical insecticide, Sitophilus zeamais, Croton heliotropiifolius, Euphorbiaceae, stored maize grain

## INTRODUCTION

Post-harvest grain loss is known to be high and unrecoverable because it is the final product. In the storage phase specifically, grain losses oscillate between 1.5 and $50 \%$, varying mainly by country. In more developed countries they oscillate at levels below $10 \%$ (around $5 \%$ in the U.S.), while in developing countries like Brazil losses may reach 50\% (FAO, 1994; Worley, 2002; Adam et al., 2006; Santos, 2011).

Mainly responsible for this damage are the insect pest of stored grain, especially the weevils (Santos et al., 1986; Guedes, 1990; FAO, 1994; Adam et al., 2006). Insects found in stored products

## J. Entomol., 9 (6): 413-421, 2012

show a wide variation in population levels. This is probably the result of management of diverse pests that, consistent with the region and discontinuous nature of the storage environment, accentuate seasonal population cycles leading to population bottlenecks where only a small number of individuals establish new populations (Tran and Credland, 1995; Guedes et al., 1997, 2003, 2007, 2009). This diversity has implications for the management of these species, because there are differences in the consumption (and losses) of grain caused by them. In addition, there are differences in the efficiency of the different control methods used to manage these pests (Guedes et al., 1997, 2003, 2007, 2009; Fragoso et al., 2005; Pimentel et al., 2007). Thus, the study of variation in response to the alternatives for control of these insects can be a source of important tools for integrated management.

The main method of control of weevils in stored grain is the use of insecticides, because it is the simplest method of control. Insecticides offer a fast and economical way to contain infestations of pests of stored products, mainly in tropical areas, due to the lack of alternative methods for control of populations of Sitophilus zeamais Motschulsky (Coleoptera: Curculionidae). The frequent use of insecticides to protect stored grain from insects has resulted in the development of resistance to these compounds in these pests and control failures have been observed in several countries (Subramanyam and Hagstrum, 1995), including Brazil (Ribeiro et al., 2003).

Management alternatives are necessary for insect pests of stored products, particularly considering the scope of the problems of insecticide resistance. Currently few conventional insecticides are permitted to be used against these pests. The limited number of control agents available drives the exploration of alternative compounds to conventional insecticides, which are particularly important for family farms, but could also present viable solutions for large storage units. In this context, reports of insecticidal activity of different plant species (Vieira et al., 2001; Moreira et al., 2005, 2007; Rosell et al., 2008; Rajendran and Sriranjini, 2008) should be considered in the search for biopesticides to replace not only conventional insecticides but also fumigants.

Studies on the toxicity of plant extracts to populations of S. zeamais contribute to guide actions that can be used in phytochemical studies and management of insect pests. For this reason, we sought to assess in this study whether the contact toxicity of ethanolic extract of C. heliotropiifolius Kunth (Euphorbiaceae) in populations of S. zeamais incurs differentiated responses.

## MATERIALS AND METHODS

Populations of $S$. zeamais were subjected to exposure by contact with ethanolic extract of C. heliotropiifolius to identify the insecticidal potential of this plant species.

Insects: We used four populations of S. zeamais collected in the cities of Bom Jesus (PI) Brazil, Redencao (PI), Volta Redonda (RJ) and São Jose do Rio Pardo (SP). The populations were multiplied and maintained in 1.5 L glass containers at a temperature of $27 \pm 2^{\circ} \mathrm{C}$ and relative humidity of $70 \pm 10 \%$.

The populations were fed grains of maize stored under refrigeration to avoid infestation. The maize was removed from the freezer and allowed to equilibrate to room temperature, approximately $27^{\circ} \mathrm{C}$, before feeding.

Plant extract: Leaves of C. heliotropilfolius were collected in the first half of May 2010 in the city of Bom Jesus in southern Piaui state, Brazil. The species was identified by Dra. Inês Cordeiro, Institute of Botany.

```
J. Entomol., 9 (6): 413-421, 2012
```

The dried leaves were triturated at room temperature in a mill and subjected to extraction (3x) with hydrated ethanol (96\%) to obtain the active chemical constituents. After evaporation of ethanol in a rotary evaporator, there remained a green solid, designated "ethanolic extract".

Bioassay topical application: The volume of $50.0 \mu \mathrm{~L}$ of ethanolic extract of C. heliotropiifolius, at a concentration of $10,000 \mathrm{ppm}$, was applied to the dorsal region of S. zeamais using an automatic pipette. The experimental unit consisted of groups of 20 adult insects ( $<15$ days-old) treated individually with $50 \mu \mathrm{~L}$ of extract and then confined in petri dishes. The controls received only the ethanol solvent. The experiment was repeated 10 times for each population. Mortality was assessed after 24,48 and 72 h .

Statistical analysis: We applied analysis of variance (PROC GLM, MANOVA; SAS, 2002), followed by univariate analysis of variance and Tukey's HSD test, when appropriate (SAS, 2002) to comp are the mean proportion of mortality among the populations studied in bioassay the topical toxicity. We used the Student's t-test to compare the effect of ethanolic extract versus control for each assessment time. Regression equations of mortality as a function of time of application were used to determine the lethal time for 50 and $90 \%$ of the population. All results were analyzed considering a significance level of $5 \%$.

## RESULTS

The mortality of $S$. zeamais adults from four populations topically treated with $10,000 \mathrm{ppm}$ of ethanolic extract from C. heliotropiifolius was significantly higher compared to control in all periods of observation, by Student's t-test at 5\% probability (Table 1).

The populations evaluated had different susceptibility in relation to contact with the ethanolic extract of $C$. heliotropilifolius at a concentration of $10,000 \mathrm{ppm}$ (Table 1). The highest observed

Table 1: Average mortality of adults of Sitophilus zeamais from four populations, toxicity caused by contact with ethanolic extract of Croton heliotropiifolius

| Population | Hours after application | Treatment |  | t-value | p-value | Comparison |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Control (a) | 10.000 ppm (b) |  |  |  |
| Redencao | 24 | $1.00 \pm 1.150^{\text {a }}$ | $4.00 \pm 2.588^{\mathrm{ab}}$ | -3.35 | 0.0040 | $a<b$ |
|  | 48 | $1.70 \pm 1.700^{\mathrm{a}}$ | $8.00 \pm 2.91^{\text {a }}$ | -5.92 | $<0.0010$ | $a<b$ |
|  | 72 | $2.90 \pm 2.690^{\text {a }}$ | $18.80 \pm 1.62^{\text {a }}$ | -16.03 | $<0.0010$ | $a<b$ |
| Bom Jesus | 24 | $0.80 \pm 0.920^{\text {a }}$ | $4.70 \pm 1.49^{\text {a }}$ | -7.03 | <0.0010 | $a<b$ |
|  | 48 | $1.60 \pm 1.580^{\text {a }}$ | $13.20 \pm 2.30^{\text {b }}$ | -13.15 | $<0.0010$ | $a<b$ |
|  | 72 | $2.80 \pm 2.970^{\mathrm{a}}$ | $17.20 \pm 1.81^{\text {a }}$ | -13.07 | $<0.0010$ | $a<b$ |
| Volta Redonda | 24 | $0.50 \pm 0.850^{\text {a }}$ | $4.18 \pm 1.60^{\mathrm{ab}}$ | -6.48 | $<0.0010$ | $a<b$ |
|  | 48 | $2.60 \pm 1.900^{\text {a }}$ | $4.64 \pm 1.57^{\text {c }}$ | -2.69 | 0.0140 | $a<b$ |
|  | 72 | $3.20 \pm 2.490^{\text {a }}$ | $5.64 \pm 1.96^{\text {b }}$ | -2.51 | 0.0220 | $a<b$ |
| Rio Pardo | 24 | $0.70 \pm 0.820^{\text {a }}$ | $2.10 \pm 1.52^{\text {b }}$ | -2.56 | 0.0200 | $a<b$ |
|  | 48 | $1.50 \pm 1.270^{\mathrm{a}}$ | $5.00 \pm 1.56^{\text {c }}$ | -5.50 | <0.0010 | $a<b$ |
|  | 72 | $2.00 \pm 1.760^{\text {a }}$ | $8.20 \pm 2.10^{\circ}$ | -7.15 | $<0.0010$ | $\mathrm{a}<\mathrm{b}$ |

Values are Mean $\pm$ SD, $\mathrm{a}<\mathrm{b}$ : Significant difference by Student's t-test at $\mathrm{p}<0.05$. Mean mortality individuals followed by the same letter in the column indicates no difference between the populations as a function of exposure time, by Tukey's test ( $\mathrm{p}>0.05$ ), $\mathrm{N}=200$
J. Entomol., 9 (6): 413-421, 2012


Fig. 1: Cumulative average percentage mortality as a function of exposure time to extract of Croton heliotropiifolius in four populations of Sitophilus zeamais
mortality was in populations from Bom Jesus and Redencao-more than $80 \%$ throughout the period of exposure to residues of the extract. All populations topically treated with $10,000 \mathrm{ppm}$ of extract of C. heliotropiifolius showed a mortality rate above 20\% (Fig. 1), indicating that the extract contains compounds that decrease the survival of the insects.

The mortality of S. zeamais adults topically treated with extract of C. heliotropiifolius at a concentration of $10,000 \mathrm{ppm}$ increased as a function of time of exposure to chemical constituents of the extract (Fig. 1, Table 1).

Through the data presented in Table 1, it appears that in the case of controls, there was no significant difference in the average mortality of S. zeamais among the populations studied at any observation time after the start of the experiment. After 24 h lower mortality was observed in the population from Rio Pardo relative to Bom Jesus. After 48 h , the average mortality for the populations from Volta Redonda and Rio Pardo was significantly lower than for the Redencao and Bom Jesus and there was no significant difference between them. The average mortality of Redencao was significantly lower than that of Bom Jesus. After 72 h , the mean mortality in S. zeamais populations from Redencao and Bom Jesus was higher and their values were similar, while mortality in the population from Volta Redonda was lower and significantly less than that of Rio Pardo. Considering the extent of the toxicity of the C. heliotropiifolius extract, it can be concluded that individuals in the population from Volta Redonda are more resistant to the toxic effect of the extract at a concentration of $10,000 \mathrm{ppm}$.

The population with lethal time before $\mathrm{TL}_{50}$ value was Bom Jesus ( 41 h ), while the after $\mathrm{TL}_{50}$ was observed in the population from Volta Redonda, with a lethal time of 441 h to kill $50 \%$ of individuals (Table 2).

The order of susceptibility of populations based on $\mathrm{TL}_{50}$ shows there is a difference in the response with regard to toxicity, when comparing the four populations of Sitophilus zeamais topically treated with the extract $C$. heliotropilfolius at a concentration of $10,000 \mathrm{ppm}$. It is important to investigate the mechanism of action of compounds from plants with insecticidal activity and correlate the mechanism of action of these compounds with commercial insecticides used in

Table 2: Regression equation for percentage of mortality, using as variable hours after application

| Populations | Independent variable | Regression coefficient | SD | Significance | $\mathrm{TL}_{50}$ (h) | $\mathrm{TL}_{90}(\mathrm{~h})$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Redencao | Intercept | -22.600 | 7.02 | 0.003 | 47.0 | 73 |
|  | Hours | 1.540 | 0.13 | $<0.001$ |  |  |
|  | R ${ }^{2}$ | 0.820 |  |  |  |  |
| Bom Jesus | Intercept | -4.000 | 5.23 | 0.450 | 41.5 | 72 |
|  | Hours | 1.300 | 0.10 | $<0.001$ |  |  |
|  | $\mathrm{R}^{2}$ | 0.850 |  |  |  |  |
| Volta Redonda | Intercept | 15.500 | 4.17 | 0.001 | 441.5 | 704 |
|  | Hours | 0.170 | 0.08 | 0.048 |  |  |
|  | $\mathrm{R}^{2}$ | 0.133 |  |  |  |  |
| Rio Pardo | Intercept | -5.000 | 4.15 | 0.238 | 71.0 | 134 |
|  | Hours | 0.635 | 0.08 | $<0.001$ |  |  |
|  | $\mathrm{R}^{2}$ | 0.692 |  |  |  |  |

LT: Lethal time
control of the maize weevil. Different responses of maize weevil populations have been well documented, as regards resistance to pyrethroids and organophosphates insecticides (Pereira et al., 2009; Correa et al., 2010).

## DISCUSSION

A high percentage of mortality in S. zeamais adults was observed in populations from Bom Jesus and Redencao, when treated topically with the ethanolic extract of C. heliotropiifolius. This toxicity can be attributed to the chemical composition of leaves of $C$. heliotropiifolius.

Species of this genus are known to contain insecticidal compounds such as catechin, gallocatechin, diterpenes and sesquiterpenes (Alexander et al., 1991; Peres et al., 1997, 1998; Doria et al., 2010; Neves and da Camara, 2012). Similarly, the ethyl acetate fraction of the species Croton urucurana caused $100 \%$ mortality in third instar larvae of Anagasta kuehniella and when incorporated into artificial diet and fed to third instar nymphs of Dysdercus maurus before the third day (Silva et al., 2009, 2012). The major component of C. heliotropiifolius essential oil was identified as $\beta$-caryophyllene, which was effective against $A$. aegypti $\left(\mathrm{LC}_{50} 544 \mathrm{ppm}\right)$. The authors suggested that the larvicidal potential is due to the existence of a synergistic effect of minor components in the essential oils (Doria et al., 2010). Croton species have high insecticidal power and in some cases are more toxic to insects than pyrethrum, a substance already commercialized in several parts of the world as an insecticide and found in chrysanthemum flowers (Guerra, 1985). Moreover, Couto and Sigrist (1995) commented that pyrethrum has a broad spectrum of insecticidal action, an observation consistent with the results obtained by Alexander et al. (1991), Kubo et al. (1991) and Morais et al. (2006).

Ntonifor et al. (2011) and Abdullahi et al. (2011) reported a high percentage of mortality of Sitophilus zeamais after treatment with plant extracts of Chenopodium ambrosioides and oil of lemon peel, respectively, at different concentrations. They observed that mortality increases with time of exposure to compounds present in the extracts, consistent with the results reported in this study, in which the highest percentage of mortality was observed 72 h after exposure via topical application in the four populations studied.

Research on responses of populations of insect pests of stored grains has been documented frequently in the last two decades (Subramanyam and Hagstrum, 1995; Guedes et al., 1997, 2003, 2007, 2009; Guedes and Zhu, 1998; Lorini and Galley, 1999; Perez-Mendoza, 1999;

Ribeiro et al., 2003). In the present study, the results obtained in topical application bioassays show there is a difference in the response of populations to the same compound. Comparing the four populations, two were collected in the South of Piaui, where use of insecticides for pests of stored grain is low and the other two were collected in the Southeast and Midwest, where there is constant use of insecticides to control pests of stored grains. The latter populations were less susceptible to the effect of C. heliotropiifolius and variation between the two was low. The difference in the response of populations from Rio Pardo and Volta Redonda relative to Bom Jesus and Redencao may result from the genetic origin of resistance in these populations to pyrethroid insecticides and phosphorous (Pereira et al., 2009; Correa et al., 2010).

Correa et al. (2010) observed that there is a difference in the response of male and female insects from the same population. The authors observed that the proportion of time spent in the untreated half is significantly higher in females compared to males. They also observed variations in responses between the populations. Females obtained from the cities of Vicosa and Canarana presented a lower proportion of time spent in relation to males.

Mortality bioassays to verify the toxicity of plant extracts are important in guiding future research for the development of new and effective products to control pests of stored grains. Data on the response of different populations are important to guide the development of new products as well as assist in integrated pest management.

The large differences observed among the maize weevil populations result from management of these insect pests and the discontinuous nature of the grain storage process. Storage units probably accentuate the seasonal cycles of these populations, enabling the rapid establishment of new populations from a small number of individuals (Tran and Credland, 1995). Some researchers report that this interpopulational variation could be caused by differences in sensory processes (Desneux et al., 2007; Hoy et al., 1998).

According to the results presented here, it is concluded that the ethanolic extract of leaves of C. heliotropiifolius has contact insecticidal activity against Sitophilus zeamais adults and the response to this toxicity varies with the geographic origin of the population analyzed. At a concentration of $10,000 \mathrm{ppm}$, all populations topically treated with the extract of $C$. heliotropiifolius had a higher mortality rate than the control.

The ethanolic extract of leaves of C. urucurana has insecticidal activity against Anagasta kuehniella and Dysdercus maurus (Silva et al., 2009, 2012). Species of this genus are known to contain insecticidal compounds such as catechin, gallocatechin, diterpenes and sesquiterpenes (Alexander et al., 1991; Peres et al., 1997, 1998; Neves and da Camara, 2012).

The ethanolic extract of leaves of C. heliotropiifolius can be incorporated into alternative methods of pest control of stored grains, assisting small farmers, so that they can develop their own methods of control based on cost-benefit, sustainability and ecological balance and thus develop alternative formulations of pesticides to control or supplement conventional insecticides. Obviously, efficacy and safety testing for new post-harvest product formulations is required before large-scale use. However, since C. heliotropiifolius is often used as a medicinal plant, it is unlikely to cause damage to the environment or safety issues.

## ACKNOWLEDGMENTS

We gratefully acknowledge Prof. Dr. Raul N.C. Guedes for providing the insect populations used in this study and the National Counsel of Scientific and Technological Development (CNPq) for granting the scientific initiation scholarship to Banco do Nordeste and financial support this research.

```
J. Entomol., 9 (6): 413-421, 2012
```


## REFERENCES

Abdullahi, N., A. Kabir and M. Yushau, 2011. Studies on the efficacy of lime peel oil in protecting stored maize against adult maize weevils (Sitophilus zeamais: Motschulsky). J. Entomol., 8: 398-403.
Adam, B.D., T.W. Phillips and P.W. Flinn, 2006. The economics of IPM in stored grain: Why don't more grain handlers use IPM?. Proceedings of the 9th International Working Conference on Stored Product Protection, October 15-18, 2006, ABRAPOS, Campinas, Brazil, pp: 3-12.
Alexander, I.C., O.K. Pascoe, P. Marchand and L.A.D. Williams, 1991. An insecticidal diterpene from Croton linearis. Phytochemistry, 30: 1801-1803.
Correa, A.S., E.M.G. Cordeiro, L.S. Braga, E.J.G. Pereira and R.N.C. Guedes, 2010. Physiological and behavioral resistance to esfenvalerate+fenitrothion in populations of the maize weevil, Sitophilus zeamais. Proceedings of the 10th International Working Conference on Stored Product Protection, June 27-July 2, 2010, Estoril, Portugal, pp: 857-851.
Couto, H.T.Z. and P.O. Sigrist, 1995. The power insecticidal chrysanthemum. Rev. Univ. Agron. Zoot., 1: 46-47.
Desneux, N., A. Decourttye and J.M. Delpuech, 2007. The sublethal effects of pesticides on beneficial arthropods. Annu. Rev. Entomol., 52: 81-206.
Doria, G.A.A., W.J. Silva, G.A. Carvalho, P.B. Alves and S.C.H. Cavalcanti, 2010. A study of the larvicidal activity of two Croton species from northeastern Brazil against Aedes aegypti. Pharm. Biol., 48: 615-620.
FAO, 1994. Grain Storage Techniques: Evolution and Trends in Developing Countries. In: Group for Assistance on Systems Relating to Grain after Harvest (GASGA), Proctor, D.L. (Ed.). FAO, USA.
Fragoso, D.B., R.N.C. Guedes and L.A. Peternelli, 2005. Developmental rates and population growth of insecticide-resistant and susceptible populations of Sitophilus zeamais. J. Stored Prod. Res., 41: 271-281.
Guedes, R.N.C., 1990. Integrated management for the protection of stored grain against insects. Rer. Bras. Armaz., 15: 3-48.
Guedes, R.N.C., S. Kambhampati, B.A. Dover and K.Y. Zhu, 1997. Biochemical mechanisms of organophosphate resistance in Rhyzopertha dominica (Coleoptera: Bostrichidae) populations from the United States and Brazil. Bull. Entomol. Res., 87: 581-586.
Guedes, R.N.C. and K.Y. Zhu, 1998. Characterization of malathion resistance in a Mexican population of Rhyzopertha dominica. Pestic. Sci., 53: 15-20.
Guedes, R.N.C., R.H. Smith and N.M.P. Guedes, 2003. Host suitability, respiration rate and the outcome of larval competition in strains of the cowpea weevil, Callosobruchus maculatus. Physiol. Entomol., 28: 298-305.
Guedes, R.N.C., N.M.P. Guedes and R.H. Smith, 2007. Larval competition within seeds: From the behavior process to the ecological outcome in the seed beetle Callosobruchus maculates. Aust. Ecol., 32: 697-707.
Guedes, R.N., L.C. Magalhaes and L.V. Cosme, 2009. Stimulatory sublethal response of a generalist predator to permethrin: hormesis, hormoligosis, or homeostatic regulation? J. Econ. Entomol., 102: 170-176.
Guerra, M.S., 1985. Alternative for the Control of Pests and Diseases of Cultivated Plants and their Products. Embrapa Publ., Braszil, Pages: 165.
Hoy, C.W., G.P. Head and F.R. Hall, 1998. Spatial heterogeneity and insect adaptation to toxins. Annu. Rev. Entomol., 43: 571-594.

Kubo, I., Y. Asaka and K. Shibata, 1991. Insect growth inhibitory nor-diterpenes, cis-dehydrocrotonin and trans-dehydrocrotonin, from Croton cajucara. Phytochemistry, 30: 2545-2546.
Lorini, I. and D.J. Galley, 1999. Deltamethrin resistance in Rhyzopertha dominica (F.) (Coleoptera: Bostrichidae), a pest of stored grain in Brazil. J. Stored Prod. Res., 35: 37-45.
Morais, S.M., E.S. Cavalcanti, L.M. Bertini, C.L. Oliveira, J.R. Rodrigues and J.H. Cardoso, 2006. Larvicidal activity of essential oils from Brazilian Croton species against Aedes aegypti L. J. Am. Mosq. Control Assoc., 22: 161-164.
Moreira, M.D., M.C. Picanco, E.M. Silva, S.C. Moreno and J.C. Martins, 2005. Use of Botanicals in Pest Control. In: Alternative Control of Pests and Diseases, Venzon, M., T.J. Paula and A. Pallini (Eds.). EPAMIG/CTZM, Vicosa do Ceara, Ceara, Brazil, pp: 89-120.

Moreira, M.D., M.C. Picanco, L.C. Barbosa, R.N. Guedes, E.C. Barros and M.R. Campos, 2007. Compounds from Ageratum conyzoides: Isolation, structural elucidation and insecticidal activity. Pest Manage. Sci., 63: 615-621.
Neves, I.A. and C.A.G. da Camara, 2012. Volatile constituents of two Croton species from caatinga biome of Pernambuco-Brazil. Rec. Nat. Prod., 6: 161-165.
Ntonifor, N.N., D.N. Forbanka and J.V. Mbuh, 2011. Potency of Chenopodium ambrosioides powders and its combinations with wood ash on Sitophilus zeamais in stored maize. J. Entomol., 8: 375-383.

Pereira, C.J., E.J.G. Pereira, E.M.G. Cordeiro, T.M.C. Della Lucia, M.R. Totola and R.N.C. Guedes, 2009. Organophosphate resistance in the maize weevil Sitophilus zeamais: magnitude and behavior. Crop Prot., 28: 168-173.
Peres, M.T., F.D. Monache, A.B. Cruz, M.G. Pizzolatti and R.A. Yunes, 1997. Chemical composition and antimicrobial activity of Croton urucurana Baillon (Euphorbiaceae). J. Ethnopharmacol., 56: 223-226.
Peres, M.T.L.P., M. Pizzolatti, R. Yunes and F.D. Monache, 1998. Clerodane diterpenes of Croton urucurana. Phytochemistry, 49: 171-174.
Perez-Mendoza, J., 1999. Survey of insecticide resistance in Mexican populations of maize weevil, Sitophilus zeamais Motschulsky (Coleoptera: Curculionidae). J. Stored Prod. Res., 35: 107-115.
Pimentel, M.A., L.R. Faroni, M.R. Totola and R.N. Guedes, 2007. Phosphine resistance, respiration rate and fitness consequences in stored-product insects. Pest Manage. Sci., 63: 876-881.
Rajendran, S. and V. Sriranjini, 2008. Plant products as fumigants for stored-product insect control. J. Stored Prod. Res., 44: 126-135.

Ribeiro, B.M., R.N.C. Guedes, E.E. Oliveira and J.P. Santos, 2003. Insecticide resistance and synergism in Brasilian populations of Sitophilus zeamais (Coleoptera: Curculionidae). J. Stored Prod. 39: 21-31.
Rosell, G., C. Quero, J. Coll and A. Guerrero, 2008. Biorational insecticides in pest management. J. Pestic. Sci., 33: 103-121.

SAS, 2002. SAS/STAT User's Guide. SAS Institute Inc., Cary, NC, USA.
Santos, J.P., 2011. Stored grain pests. http://www.agencia.cnptia.embrapa.br/ gestor/milho/arvore/CONTAG01_38_168200511158.html
Santos, J.P., I.V.M. Cajueiro and R.A. Fontes, 1986. Evaluation of Losses Caused by Insects in Stored Maize at the Farm Level, in three States. In: Annual Technical Report of the National Research Center for Maize and Sorghum, Paiva, E. (Ed.). CNMPM/EMBRAPA, Sete Lagoas, Brazil, pp: 65-66.

## J. Entomol., 9 (6): 413-421, 2012

Silva, L.B., M.T.L.P. Peres, W. Silva and M.L.R. Macedo, 2009. Effects of Croton urucurana extracts and crude on Anagasta kuehniella (Lepidoptera: Pyralidae). Braz. Arch. Biol. Technol., 3: 653-664.
Silva, L.B., Z.F. Xavier, C.B. Silva, O. Faccenda, A.C.S. Candido and M.T.L.P. Peres, 2012. Insecticidal effects of Croton urucurana extracts and crude resin on Dysdercus maurus (Hemiptera: Pyrrocoridae). J. Entomol., 9: 98-106.
Subramanyam, B. and D.W. Hagstrum, 1995. Resistance Measurement and Management. In: Integrated Management of Insects in Stored Products, Subramanyam, B. and D.W. Hagstrum (Eds.). Marcel Dekker, New York, pp: 331-397.
Tran, B.M.D. and P.F. Credland, 1995. Consequences of inbreeding for the cowpea seed beetle, Callosobruchus maculatus. Biol. J. Linn. Soc., 56: 483-503.
Vieira, P.C., J. Mafezoli and M.W. Biavatti, 2001. Insecticides of Plant Origin. In: Natural Products to Control Insects, Ferreira, J.T.B., A.G. Correa and P.C. Vieira (Eds.). University of Sao, Sao Carlos, Brazil, pp: 23-45.
Worley, J., 2002. Aerating Grain in Storage. Service Bulletin No. 712, Georgia Cooperative Extension, Athens, GA, EUA.

