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# **Short** Communication

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# α-amylase Expressions in Indian Type-2 Diabetic Patients

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α-amylases are enzymes which hydrolyze starch molecules to give diverse products including dextrins and progressively smaller polymers composed of glucose units which causes hyperglycemia and development of type 2 diabetes mellitus. Therefore, we estimated the α-amylase activity, HbA1C and blood glucose in Indian type 2 diabetic patients. Study group's follows: 20 Healthy subjects, 30 newly detected patients and 43 type 2 diabetics on treatment with metformin and pioglitazone combination and 30 pioglitazone alone treated. In all subjects blood sugar was estimated and anti diabetic activity criterion was taken for the diagnosis. In the same subjects fasting and post-prandial α-amylase activity was estimated, HbA1c levels were compared with α-amylase activity in healthy and treated subjects. The α-amylase activity in type 2 diabetic patients was significantly different from control subjects and significant relation between α-amylase activity and HbA1c was observed in diabetic subjects (p<0.01), in type 2 diabetic patients with HbA1c >12% showed significantly (p<0.01) higher αamylase activity than newly diagnosed and healthy, for three months treatment in type 2 diabetic patients showed a significant controlled the blood glucose, elevated alpha amylase levels. Post-prandial α-amylase activity was markedly raised in type 2 diabetes, this could contribute to the increased in circulating blood glucose. Fasting and post-prandial α-amylase activity and lipid profiles were significantly controlled in pioglitazone alone and pioglitazone with metformin combination treated groups. This change may effect of metformin and pioglitazone on post-prandial  $\alpha$ -amylase activity.

**Key words:** α-amylase, blood glucose, HbA1C, type 2 diabetes

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#### INTRODUCTION

Diabete mellitus (DM) is a group of disorders of carbohydrate metabolism characterized by hyperglycemia, with an estimated 170 million patients worldwide in the year 2000. Its prevalence is rapidly increasing (Wild et al., 2004). It is a clinically complex and associated with many serious complications including kidney failure, blindness and cardiovascular disease (Baliga and Weinberger, 2006). The chronic Diabetes mellitus details a substantial decrease in quality of life and life expectancy (Coffey et al., 2002; Stephens et al., 2006). In the previous studies, diabetes is increased with metabolic disorders of carbohydrates (Ben-Aryeh et al., 1988; Streckfus et al., 1994; Belazi et al., 1998). In recent studies pancreatic dysfunction may leads to diabetes mellitus (Hardt et al., 2000; Lernmark, 2000).

Clinical investigation observations indicate close relation between pancreatic exo- and endocrine parts (Okabayashi et al., 1988; Aughsteen and Kataoka, 1993; Meral et al., 2001). Insulin dysfunction and glucagon activation is due to pancreatic impairment (Adeghate, 1999; Taniyama et al., 1999; Hardt et al., 2000; Lernmark, 2000; Harding et al., 2001). The decreased production of exocrine secretion also influence on insulin deficiency (Okabayashi et al., 1988; Aughsteen and Kataoka, 1993; Kim et al., 2000).

Interestingly, the relation between insulin and glucagon concentrations in extracellular fluid and their antagonistic actions also seem to play a key role in pancreatic exocrine function disorders. Insulin secretion is enhancing the exocrine activity like alpha amylase functions but glucagon inhibits it (Taniyama *et al.*, 1999; Hardt *et al.*, 2000). Amylases are enzymes that hydrolyze the carbohydrates into small glucose particles (Windish and Mhatre, 1965).

All the alpha amylase enzyme molecules acts on alpha 1-4, 1-6 links of glucose residues (Van der Maarel *et al.*, 2002). Amylases were categorized into two, that one is endoamylase and another one is exomylase the endoamylases are involved in hydrolysis of carbohydrates in to different chain lengths of linear and branched molecules and exoamylases reduced in to short molecules (Gupta *et al.*, 2003). The enzyme alpha amylase is one of the most important in human body responsible of hydrolysis of starch in to small sugar molecules (Alexander, 1992).

Amylase is one of the main enzymes produced in exocrine pancreatic cells, may be recognized as an adequate indicator of organ's activity both in physiological and pathological states. Moreover, these studies might help in the development of effective alpha amylase inhibitors.

### MATERIALS AND METHODS

Amylase kit purchased from Becan Pvt. Ltd, Mumbai, India. Glucose kit was from Excel Diagnostics, Hyderabad, India. Metformin and pioglitazone combination tablets were from Sun Pharma Pvt Ltd, Sikkim, India. Pioglitazone tablets were from Sun Pharma Pvt Ltd, Sikkim.

### Study design follows:

- **Group 1:** Twenty non-diabetic subjects (control)
- **Group 2:** Thirty newly diagnosed type 2 diabetes without treatment
- **Group 3:** Thirty newly diagnosed type 2 diabetic patients with pioglitazone treatment for three months
- **Group 4:** Forty three diabetic patients on oral antidiabetic combination treatment for three months (pioglitazone with metformin)

All patients were recruited at the department of general medicine, Mahatma Gandhi memorial Hospital, Warangal andhra Pradesh, India, from March 2008 to April 2010. All subjects were attending general health check up at our outpatient department in MGM Hospital. Subjects were excluded if they had chronic gastrointestinal diseases associated with chronic pancreatitis, history of any malignant disease, history of alcohol abuse, kidney or liver failure and other diseases affecting carbohydrate metabolism. Fasting as well as post-prandial blood samples were collected from all subjects and fasting and post-prandial serum glucose (Trinder, 1969), alpha amylase levels and fasting lipid profiles (Bucolo and David, 1973) were estimated by a colorimetric assay. The study was approved by institutional Human ethics committee (University College of pharmaceutical sciences, Kakatiya University, Warangal) and informed consent was obtained from each subject according to the principles of the declaration of Helsinki.

**Statistical analysis:** All variables are expressed as Mean±SD. Group differences of continuous variables were compared using ANOVA followed by Student-Newman Keuls post hoc test. For all analyses, a p<0.05 was considered to be statistically significant. All analyses were performed using INSTAT 1.12 (Graph-Pad Software, Inc., San Diego, CA).

#### RESULTS

Alpha amylase levels in fasting and postprandial statuses in different groups were compared (nondiabetic,

Table 1: Clinical characteristics and alpha amylase expression in participating subjects in this study

Parameter	Non diabetic	Newly-detected	Pioglitazone treated	Oral antidiabetic (combination)
Age (years)	25.85±2.4	44.12±12.1	44.12±12.1	49.9±10.6
Sex (M/F)	20 (12/08)	30 (17/13)	30 (17/13)	43 (25/18)
BMI (Kg m <sup>-2</sup> )	$21.5\pm2.3$	26.3±3.3	24.3±3.3	23.5±2.3
Fasting glucose mg (%)	$76.8 \pm 11.4$	169.3±23.4	107.8±18.5**	95.1±25.4***
Post-prandial glucose mg (%)	104.5±11.1	243.6±29.5	131.3±22.7***	124.6±20.5***
Fasting α-amylase activity (U L <sup>-1</sup> )	14.2±1.1	55.32±3.9	33.1±2.9*	35.6±1.7*
Post-prandial $\alpha$ -amylase activity (U L <sup>-1</sup> )	$33.1\pm1.4$	81.2±4.2	40.4±1.9***	37.2±1.5***
HbA1c (%)	4.7±0.31%	14.3±1.50%	7.3±1.11% ***	5.8±0.56%****

Data values were expressed as Mean±SD. p-value less than 0.05 are considered as statistically significant. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, \*\*\*\*p<0.0001

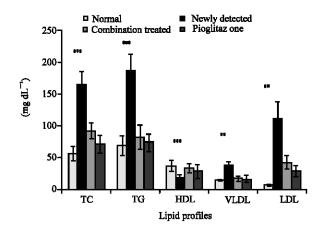


Fig. 1: Serum lipid profiles in healthy, newly detected and treated groups, Data values are expressed as Mean±SD. \*\*\* p<0.01, \*\*\*\* p<0.001

newly detected, pioglitazone alone treated and combination drug treated). Table 1 represents the clinical characteristics of all group subjects alpha amylase levels fasting and postprandial in oral antidiabetic (combination) drugs (35.6±1.7 and 37.2±1.5 U L<sup>-1</sup>) and pioglitazone alone treated groups showed significantly lowered (33.1±2.9 and 40.4±1.9 U L<sup>-1</sup>) than newly diagnosed type-2 diabetic patients (55.32±3.9 and 81.2±4.2 U L<sup>-1</sup>). Statistical significant levels of alpha amylase in different groups represented in Table 1.

The results of present study indicates that fasting alpha amylase levels in different groups were statistically significant (p<0.05). The significant relation between alpha amylase and HbA1c was observed in type 2 diabetic patients (p<0.001). Type 2 diabetes with HbA1c >12% showed significantly (p<0.031) higher alpha amylase activity (Mean±SD; 74.3±4.6 UL<sup>-1</sup>) and lipid profiles LDL, Triglycerides and total cholesterol levels (Fig. 1) than healthy ((HbA1c<7% and alpha amylase activity (23.4±1.2 UL<sup>-1</sup>, respectively)). Variations in alpha amylase activity for 3 months treatment in type 2 diabetic patients showed a significant reduced the HbA1c (p<0.001), similarly fasting and post-prandial, alpha amylase levels and LDL, Triglycerides and total cholesterol levels in

pioglitazone and combination treated groups but increase the HDL levels (p<0.001), except between the non-diabetic Vs anti-diabetic drug treated groups.

#### DISCUSSION

The results shows that in the case of type 2 diabetes their FBS and HbA1c levels were far above the normal range but their serum alpha amylase enzyme levels were significantly elevated. By Pioglitazone monotherapy their FBS, HbA1c and alpha amylase levels were well controlled to the near normal values. The pioglitazone therapy is very effective for obese type 2 diabetic patients. The group 4 patients showed a marked amelioration of hyperglycemia and alpha amylase and HbA1c with combination therapy of pioglitazone and metformin. The highest fall of alpha amylase observed in combined therapy may be due to two factors. i.e., due to very high pretreatment level of FBS and also due to insulin sensitizer action of pioglitazone. Similarly the comparatively higher FBS fall for pioglitazone group is the drug of choice that worked here also as it enhances the peripheral uptake of glucose. We can observe a parallel decrease of HbA1c with alpha amylase in the pioglitazone and metformin combined therapy group. Therefore, combination therapy is more effective than monotherapy for controlling.

Metformin Inhibits Proinflammatory Responses and Nuclear Factor-B in Human Vascular Wall Cells and antioxidant activity (Winder and Hardie, 1999; Isoda *et al.*, 2006). Several reports have demonstrated an anti-inflammatory action of the specific ligands of PPAR $\gamma$  activators of PPAR $\gamma$  inhibit the generation of proinflammatory cytokines such as interleukin-1 $\beta$  (IL- 1 $\beta$ ), tumor necrosis factor-a (TNF-a) and interleukin-6 (Suganami *et al.*, 2005). PPAR $\gamma$  agonists increases HDL particles to inhibit dylipidemea and antioxidant activity (Barter *et al.*, 2004). Pioglitazone reduced serum triglycerides and increased HDL-C to a greater extent than either rioglitazone or rosiglitazone. In addition, trioglitazone and rosiglitazone increased LDL-C whereas, pioglitazone did not (Van der Maarel *et al.*, 2002;

Gupta et al., 2003). All these studies and our present findings stress the importance of pioglitazone in combination therapy but with all the required precautions.

## **CONCLUSIONS**

Indian ethnic group's shows increased Post-prandial Alpha Amylase levels were significantly higher in type 2 diabetic group when compared to normal controls. This is a contradictory finding that serum Alpha Amylase activity in patients with type-2 diabetes mellitus relatively positive with HbA1c, blood glucose and lipid profile levels were completely controlled to normal by pioglitazon alone and combination with metformin treatment but increased the HDL levels. The results may provide important information concerning the activity of alpha amylase in type 2 diabetic patients and its association with alphaamylase and glucose.

# REFERENCES

- Adeghate, E., 1999. Distribution of calcitonin-gene-related peptide, neuropeptide-Y, vasoactive intestinal polypeptide, cholecystokinin-8, substance P and islet peptides in the pancreas of normal and diabetic rats. Neuropeptides, 33: 227-235.
- Alexander, R., 1992. Maltodextrins: Production, Properties and Applications. In: Starch Hydrolysis Products: Worldwide Technology, Production and Applications, Schenk, F. and R. Hebeda (Eds.). John Wiley and Sons Inc., New York, USA., ISBN-13: 9780471187967, pp: 62-122.
- Aughsteen, A.A. and K. Kataoka, 1993. Morphometric studies on the juxta-insular and tele-insular acinar cells of the pancreas in normal and streptozotocin-induced diabetic rats. J. Electron Microscopy, 42: 79-87.
- Baliga, B.S. and J. Weinberger, 2006. Diabetes and stroke: Part one-risk factors and pathophysiology. Curr. Cardiol. Rep., 8: 23-28.
- Barter, P.J., S. Nicholls, K.A. Rye, G.M. Anantharamaiah, M. Navab and A.M. Fogelman, 2004. Antiinflammatory properties of HDL. Circ. Res., 95: 764-772.
- Belazi, M.A., A. Galli-Tsinopoulou, D. Drakoulakos, A. Fleva and P.H. Papanayiotou, 1998. Salivary alterations in insulindependent diabetes mellitus. Int. J. Paediatric Dentistry, 8: 29-33.
- Ben-Aryeh, H., M. Cohen, Y. Kanter, R. Szargel and D. Laufer, 1988. Salivary composition in diabetic patients. J. Diabetes Complications, 2: 96-99.

- Bucolo, G. and H. David, 1973. Quantitative determination of serum triglycerides by the use of enzymes. Clin. Chem., 19: 476-482.
- Coffey, J.T., M. Brandle, H. Zhou, D. Marriott and R. Burke *et al.*, 2002. Valuing health-related quality of life in diabetes. Diabetes Care, 25: 2238-2243.
- Gupta, R., P. Gigras, H. Mohapatra, V.K. Goswami and B. Chauhan, 2003. Microbial α-amylases: A biotechnological perspective. Process Biochem., 38: 1599-1616.
- Harding, H.P., H. Zeng, Y. Zhang, R. Jungries and P. Chung et al., 2001. Diabetes mellitus and exocrine pancreatic dysfunction in perk-/- mice reveals a role for translational control insecretory cell survival. Mol. Cell, 7: 1153-1163.
- Hardt, P.D., A. Krauss, L. Bretz, M. Porsch-Ozcurumez and H. Schnell-Kretschmer et al., 2000. Pancreatic exocrine function in patients with type 1 and type 2 diabetes mellitus. Acta Diabetol., 37: 105-110.
- Isoda, K., J.L. Young, A. Zirlik, L.A. MacFarlane and N. Tsuboi *et al.*, 2006. Metformin inhibits proinflammatory responses and nuclear factor-kB in human vascular wall cells. Arteriosclerosis Thrombosis Vasc. Biol., 26: 611-617.
- Kim, K.H., H.S. Lee, C.D. Kim, H.J. Chun and C.W. Song et al., 2000. Evaluation of pancreatic exocrine function using pure pancreatic juice in noninsulin-dependent diabetes mellitus. J. Clin. Gastroenterol., 31: 51-54.
- Lernmark, A., 2000. Rapid-onset type 1 diabetes with pancreatic exocrine dysfunction. N. Engl. J. Med., 342: 344-345.
- Meral, I., Z. Yener, T. Kahraman and N. Mert, 2001. Effect of *Nigella sativa* on glucose concentration, lipid peroxidation, antioxidant defence system and liver damage in experimentally induced diabetic rabbits. J. Vet. Med., 48: 593-599.
- Okabayashi, Y., M. Otsuki, A. Ohki, I. Suehiro and S. Baba, 1988. Effect of diabetes mellitus on pancreatic exocrine secretion from isolated perfused pancreas in rats. Dig. Dis. Sci., 33: 711-717.
- Stephens, J.M., M.F. Botteman and J.W. Hay, 2006. Economic impact of antidiabetic medications and glycemic control on managed care organizations: A review of the literature. J. Manag. Care Pharm., 12: 130-142.
- Streckfus, C.F., S. Marcus, S. Welsh, R.S. Brown, G. Cherry-Peppers and R.H. Brown, 1994. Parotid function and composition of parotid saliva among elderly edentulous African-American diabetics. J. Oral Pathol. Med., 23: 277-279.

- Suganami, T., J. Nishida and Y. Ogawa, 2005. A paracrine loop between adipocytes and macrophages aggravates inflammatory changes: Role of free fatty acids and tumor necrosis factor α. Arterioscler. Thromb. Vasc. Biol., 25: 2062-2068.
- Taniyama, H., K. Hirayama, Y. Kagawa, T. Kurosawa, M. Tajima, T. Yoshino and H. Furuoka, 1999. Histopathological and immunohistochemical analysis of the endocrine and exocrine pancreas in twelve cattle with Insulin-Dependent Diabetes Mellitus (IDDM). J. Vet. Med. Sci., 61: 803-810.
- Trinder, P., 1969. Determination of blood glucose using an oxidaseperoxidase system with a non-carcinogenic chemogen. J. Clin. Pathol., 22: 158-161.
- Van der Maarel, M.J.E.C., B. van der Veen, J.C.M. Uitdehaag, H. Leemhuis and L. Dijkhuizen, 2002. Properties and applications of starchconverting enzymes of the α-amylase family. J. Biotechnol., 94: 137-155.

- Wild, S., G. Roglic, A. Green, R. Sicree and H. King, 2004. Global prevalence of diabetes: Estimates for the year 2000 and projections for 2030. Diabetes Care, 27: 1047-1053.
- Winder, W.W. and D.G. Hardie, 1999. AMP-activated protein kinase, a metabolic master switch: Possible roles in type 2 diabetes. Am. J. Physiol., 277: 1-10.
- Windish, W.W. and N.S. Mhatre, 1965. Microbial Amylases. In: Advances in Applied Microbiology, Wayne, W.U. (Ed.). Vol. 7. Academic Press, New York., USA., pp. 273-304.