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## The Bioenergetics of the House Fly *Musca domestica vicina* and the Blowfly *Lucilia sericata*

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### Abstract

Temperature caused a significant change in the bioenergetics of the two forensically important flies namely, the housefly *Musca domestica vicina* Macquart and the blow fly *Lucilia sericata* Meigen in the laboratory under different constant temperatures. The oxygen consumption of both species increased with an increase in the larval weight at all tested temperatures. Conversely, as the weights of both species increased, the oxygen consumed per mg fresh or dry weight decreased. The pupal respiration followed the U-shaped curve. The calculated temperature coefficient ( $Q_{10}$ ) values were not very high for all stages of both species at lower temperature. The instantaneous rate of assimilation of the feeding larvae of both species increased with increasing weight and age. The instantaneous growth efficiency and the cumulative growth efficiency values at all tested temperatures were higher for *L. sericata* than *M. domestica*. The oxygen consumption and food assimilation are age markers of the immature stages of these two forensically important flies and can be used easily in the accurate estimation of the postmortem interval.

### Introduction

Tropic-dynamic aspect of ecology has resulted in a number of studies on the bioenergetics of consumer species, usually in the laboratory where metabolic parameters may be precisely measured. Such information can then be extrapolated to field situations, not only for the species in question, but also for ecologically similar groups (Welch, 1968). In bioenergetics studies, the metabolism can be measured in considerable detail as a function of the most significant factors, both internal; such as age, size, physiological state etc. and external; such as temperature, relative humidity, the nature and availability of food etc. The relationship between oxygen consumption and weight of larvae or unit weight were previously studied by Drummond and Chamberlain (1961), Healey (1967), Guerra and Cochran (1970), Tilbrook and Block (1972), Otto (1974), Block and Tilbrook (1975), Swift (1976), Putman (1977) Ziser and Nettles (1979) and Martin (1991). The oxygen consumption of the pupal stage always forms a U-shaped curve like that of most other metamorphosing insects (Keister, 1953; Park and Buck, 1960; Guerra and Cochran, 1970; Shishin'ova, 1986; and Shaurub, 1993). Waldbauer (1968) stated that the highest value of the instantaneous growth efficiency ( $K_2$ ) was coincide with the most intense production phase. Whereas, the cumulative growth efficiency was varied with temperature, developmental stage, or the weight of the larvae (Hanski, 1976). Therefore, the present study deals with the energetics of two forensically important species, *Musca domestica* and *Lucilia sericata*. From such studies, information about the species' potentialities under different conditions can be provided, and the possibilities of using these factors to determine the age of the larvae can be established.

### Materials and Methods

Cultures of *M. domestica* and *L. sericata* were established in the laboratory of the Zoology Department, Faculty of Science, Alexandria University, and were maintained at 27°C and relative humidity of 70%. Larvae of *M. domestica* were fed on chicken's mash whereas those of *L. sericata* were fed on fresh liver. Fifty eggs for each species were placed in 20 gm of larval medium. Ten replicates were reared under five constant temperatures (15, 20, 25, 30 and 35°C). Oxygen consumption was measured with conventional Warburg respirometer (Dixon, 1951), by direct manometric method outlined by Umbreit et al. (1957). All the experiments were carried out during same hour each day and the tested specimens were used only for once. The data were presented in the forms:

1.  $\mu\text{O}_2$  consumed/individual/hr.
2.  $\mu\text{O}_2$  consumed/mg fresh weight/hr.
3.  $\mu\text{O}_2$  consumed/mg dry weight/hr.

The temperature coefficient ( $Q_{10}$ ) was calculated for each 5°C increment in temperature (Cairns, 1978) using formula:

$$\text{Log } Q_{10} = \frac{10(\log K_1 - \log K_2)}{t_1 - t_2}$$

where  $K_1$  and  $K_2$  are the oxygen consumption rates at  $t_1$  and  $t_2$  °C respectively (Hoar, 1966).

For the estimation of larval assimilation; ten replicates of each species, at each of the experimental temperatures were prepared as previously mentioned. Replicates were undisturbed till the end of the feeding period. According to Hanski (1976); assimilation was measured according to equation  $A_i = P_i + R_i$  where  $A_i$  is the instantaneous rate of assimilation,  $P_i$  is the instantaneous rate of production

$R_i$  is the instantaneous rate of respiration, or  $A_c = 0.98$  ( $B_{t1}, B_{t2}$ ) where  $A_c$  is the cumulative food assimilation,  $B_{t1}$  and  $B_{t2}$  were the weights of the larval medium at the beginning and the end of the experiment, respectively. The growth efficiency was calculated as a cumulative index  $K_{2c}$  (Klekowski *et al.*, 1967), where  $K_{2c} = P_c / A_c \cdot 100$  and as an instantaneous index  $K_{2i} = P_i / A_i \cdot 100$ . Both indices were calculated for each of the cited temperature.

**Results**

The daily oxygen consumption of the developing embryos inside eggs, feeding and post feeding larvae, and pupal stage of both flies raised at five different constant temperatures, are illustrated graphically in Fig. 1-6. The results showed that oxygen consumption of the various immature stages of both species increased with increasing temperature. Person correlation coefficient displayed a significant correlation ( $r = 0.97, 0.97, 0.95$  and  $0.87$  for the developing embryos inside eggs, feeding larvae, post feeding larvae and pupae of *M. domestica* and  $r = 0.99, 0.97, 0.94$  and  $0.91$  for similar stages of *L. sericata*).

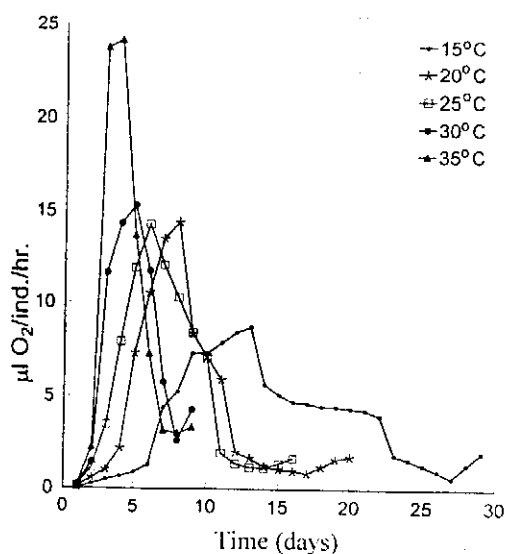


Fig. 1: The daily oxygen consumption ( $\mu\text{O}_2/\text{ind.}/\text{hr.}$ ) of the different immature stages of *M. domestica*, raised at five different constant temperatures.

At all tested temperatures, for the two species, the developing embryos inside eggs had the lowest consumption value. The oxygen consumption increased at the onset of the feeding larval stage and reached its maximum value at the end of this stage then it started to decrease at the beginning and during the post feeding period. This is followed by a decrease during the pupal stage. The daily oxygen consumption of the various immature stages at all tested temperatures is higher for *L. sericata* than *M. domestica*.

The calculated temperature coefficient values ( $Q_{10}$ ) of the

oxygen consumption for the different stages of both species are given in Table 1. It can be seen from this table that, at low temperatures ( $15\text{-}20^\circ\text{C}$ ), the  $Q_{10}$  values of all stages of both species were generally higher than at the higher temperatures ( $30\text{-}35^\circ\text{C}$ ).

Table 1: Temperature coefficient ( $Q_{10}$ ) calculated for each increment in the temperature range from the oxygen consumption rates of different immature stages of *Musca domestica* and *Lucilia sericata*.

Stage	Temp. ( $^\circ\text{C}$ ) Coefficient ( $Q_{10}$ )			
	15-20	20-25	25-30	30-35
<b>Eggs</b>				
<i>M. domestica</i>	2.08	2.16	2.59	1.05
<i>L. sericata</i>	1.72	1.94	1.92	1.33
<b>Larvae</b>				
<i>M. domestica</i>	2.56	1.20	1.39	2.02
<i>L. sericata</i>	1.78	4.19	1.19	1.46
<b>Post-feeding larvae</b>				
<i>M. domestica</i>	2.39	1.74	2.04	1.96
<i>L. sericata</i>	1.43	1.96	1.12	1.03
<b>Pupae</b>				
<i>M. domestica</i>	1.01	1.11	8.54	0.99
<i>L. sericata</i>	0.99	1.35	5.99	1.25

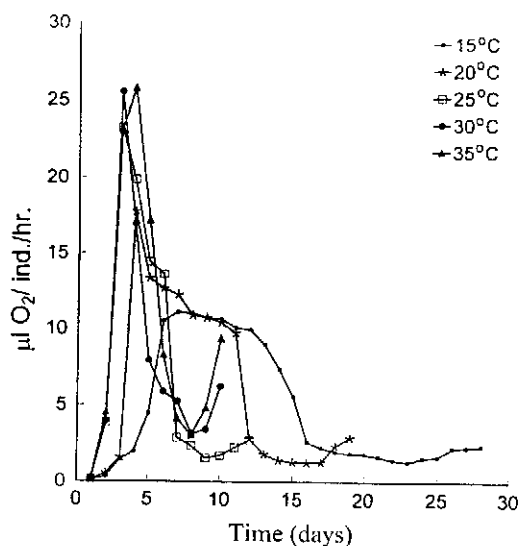


Fig. 2: The daily oxygen consumption ( $\mu\text{O}_2/\text{ind.}/\text{hr.}$ ) of the different immature stages of *L. sericata*, raised at five different constant temperatures.

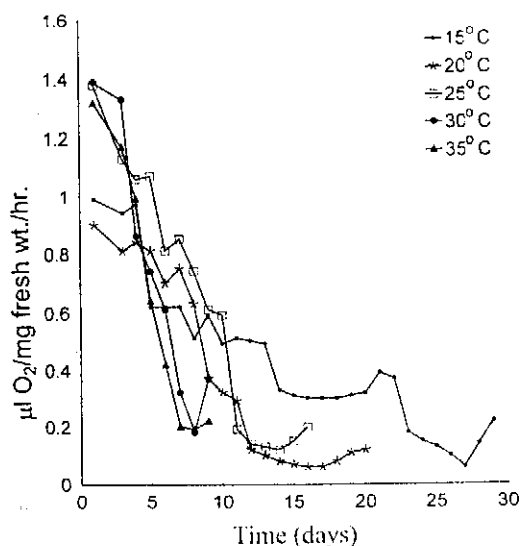


Fig. 3: The daily oxygen consumption ( $\mu\text{O}_2/\text{mg}$  fresh wt./hr.) of the different immature stages of *M. domestica*, raised at five different constant temperatures.

It can be seen from figures 3-6 that when oxygen consumption is expressed on a unit weight basis, the rate of consumption at all tested temperatures decreases in the series feeding larvae-post feeding larvae-pupae.

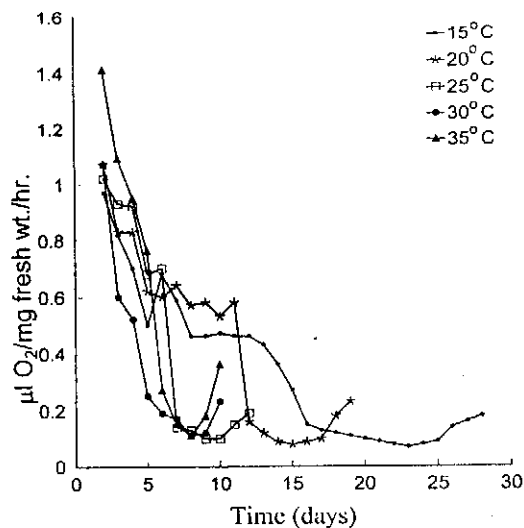


Fig. 4: The daily oxygen consumption ( $\mu\text{O}_2/\text{mg}$  fresh wt./hr.) of the different immature stages of *L. sericata*, raised at five different constant temperatures.

In the present study the instantaneous and cumulative food assimilation as well as the instantaneous and cumulative growth efficiency for the feeding larval stage of *M. domestica* and *L. sericata* raised at different five constant temperatures are presented in Tables 2-6. It can be observed from the data cited in tables 2 & 3, that the instantaneous rate of assimilation ( $A_i$ ) increased with the increase of weight and age of the feeding larvae. Furthermore,  $A_i$  of both species increases with temperature. Person correlation coefficient indicated significant correlation ( $r = 0.95, 0.96$  for *M. domestica* and *L. sericata* respectively).

From the present data, it can be recorded that 25 and 30°C represent an optimum production in *M. domestica* and *L. sericata* respectively. Also, it can be observed that food assimilation at all tested temperatures is higher for *L. sericata* than *M. domestica*.

### Discussion

The results presented clearly show that oxygen consumption rates of the various stages of the tested species are temperature dependent; the rate increases with temperature. These results add support to similar studies by other workers on a variety of insects (Keister and Budd 1961; Tribe and Bowler, 1968; and Cairns, 1978) and a wide variety of poikilotherms (Dawson and Bartholomew 1956; Grainger, 1956; Kanugo and Prosser, 1959; Beamish, 1964; Edney, 1964; McFarland and Pickens 1965 and Davies, 1966).

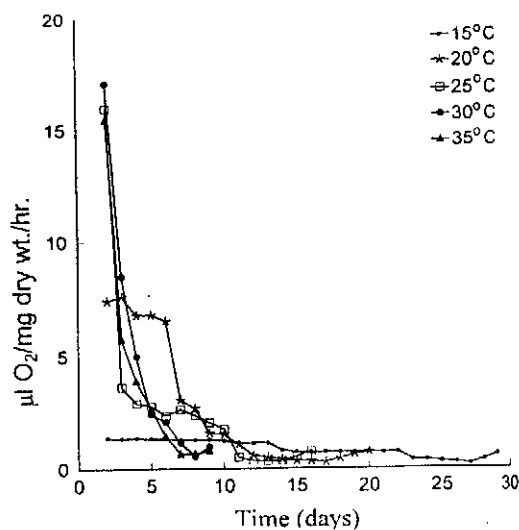


Fig. 5: The daily oxygen consumption ( $\mu\text{O}_2/\text{mg}$  dry wt./hr.) of the different immature stages of *M. domestica* raised at five different constant temperatures.

Table 2: Mean  $\pm$  SE of instantaneous rate of assimilation ( $A_1 = P_1 + R_1$ ) of the feeding larval stage of *M. domestica*, raised at five different constant temperatures.

Day	15°C	20°C	25°C	30°C	35°C
1	0.39 $\pm$ 0.03	0.63 $\pm$ 0.07	1.19 $\pm$ 0.05	1.49 $\pm$ 0.12	2.38 $\pm$ 0.08
2	0.62 $\pm$ 0.02	1.07 $\pm$ 0.10	4.35 $\pm$ 0.17	12.91 $\pm$ 0.85	27.68 $\pm$ 1.19
3	0.79 $\pm$ 0.02	2.37 $\pm$ 0.05	9.76 $\pm$ 0.43	15.84 $\pm$ 0.94	28.15 $\pm$ 2.10
4	1.04 $\pm$ 0.08	8.09 $\pm$ 0.15	13.40 $\pm$ 0.22	18.71 $\pm$ 1.91	
5	1.63 $\pm$ 0.04	11.05 $\pm$ 0.26	16.07 $\pm$ 0.51		
6	7.10 $\pm$ 0.47	16.29 $\pm$ 0.28			
7	7.70 $\pm$ 0.22				
8	8.93 $\pm$ 0.13				
9	8.93 $\pm$ 0.50				
10	8.94 $\pm$ 0.41				
11	8.74 $\pm$ 0.96				
12	8.90 $\pm$ 0.15				
Mean $\pm$ S.E.	5.31 $\pm$ 0.11	7.97 $\pm$ 0.10	8.95 $\pm$ 0.13	12.25 $\pm$ 0.81	19.40 $\pm$ 0.39

Table 3: Mean  $\pm$  SE of instantaneous rate of assimilation ( $A_1 = P_1 + R_1$ ) of the feeding larval stage of *Lucilia sericata*, raised at five different temperatures.

Day	15°C	20°C	25°C	30°C	35°C
1	0.42 $\pm$ 0.03	0.65 $\pm$ 0.01	4.94 $\pm$ 0.09	4.68 $\pm$ 0.28	5.33 $\pm$ 0.32
2	1.73 $\pm$ 0.14	1.89 $\pm$ 0.46	28.89 $\pm$ 1.31	35.70 $\pm$ 2.90	27.51 $\pm$ 1.88
3	2.14 $\pm$ 0.17	22.60 $\pm$ 1.23			29.41 $\pm$ 0.91
4	5.91 $\pm$ 0.18				
5	12.28 $\pm$ 0.66				
6	12.34 $\pm$ 0.75				
7	12.78 $\pm$ 0.26				
Mean $\pm$ S.E.	6.80 $\pm$ 0.08	8.38 $\pm$ 0.40	16.92 $\pm$ 0.67	20.19 $\pm$ 1.47	20.75 $\pm$ 0.64

Table 4: Mean  $\pm$  SE of instantaneous growth efficiency ( $K_{21} = P_1/A_1 \cdot 100$ ) of the feeding larval stage of *Musca domestica*, raised at five different constant temperatures.

Day	15°C	20°C	25°C	30°C	35°C
1	33.59 $\pm$ 2.74	11.11 $\pm$ 1.41	5.04 $\pm$ 0.21	4.69 $\pm$ 0.37	5.88 $\pm$ 0.43
2	25.81 $\pm$ 0.78	4.67 $\pm$ 0.51	20.23 $\pm$ 1.19	9.99 $\pm$ 0.56	14.38 $\pm$ 0.59
3	17.72 $\pm$ 0.41	8.02 $\pm$ 0.11	18.34 $\pm$ 0.89	9.47 $\pm$ 0.58	7.80 $\pm$ 0.56
4	16.35 $\pm$ 2.33	9.27 $\pm$ 0.03	11.27 $\pm$ 0.02	18.33 $\pm$ 2.16	
5	20.25 $\pm$ 0.83	4.79 $\pm$ 0.23	11.08 $\pm$ 0.37		
6	18.03 $\pm$ 1.26	17.19 $\pm$ 0.31			
7	5.84 $\pm$ 0.30	6.13 $\pm$ 0.09			
8	7.92 $\pm$ 0.16				
9	4.48 $\pm$ 0.81				
10	4.52 $\pm$ 0.52				
11	3.66 $\pm$ 0.55				
12	2.02 $\pm$ 0.03				
Mean $\pm$ S.E.	12.26 $\pm$ 0.38	8.74 $\pm$ 0.23	13.19 $\pm$ 0.16	10.62 $\pm$ 0.67	9.35 $\pm$ 0.14

Table 5: Mean  $\pm$  SE of instantaneous growth efficiency ( $K_{21} = P_1/A_1 \cdot 100$ ) of the feeding larval stage of *Lucilia sericata*, raised at five different constant temperatures.

Day	15°C	20°C	25°C	30°C	35°C
1	26.19 $\pm$ 2.33	21.54 $\pm$ 0.13	22.67 $\pm$ 0.07	16.67 $\pm$ 1.02	15.95 $\pm$ 1.03
2	18.49 $\pm$ 0.45	17.46 $\pm$ 2.06	19.76 $\pm$ 0.99	28.58 $\pm$ 2.43	16.24 $\pm$ 1.15
3	8.41 $\pm$ 0.96	21.64 $\pm$ 0.44			12.48 $\pm$ 0.42
4	25.04 $\pm$ 0.74				
5	14.58 $\pm$ 0.75				
6	10.45 $\pm$ 0.76				
7	14.24 $\pm$ 0.29				
Mean $\pm$ S.E.	16.77 $\pm$ 0.35	20.21 $\pm$ 0.71	21.22 $\pm$ 0.51	22.62 $\pm$ 1.34	14.89 $\pm$ 0.16

Table 6: The cumulative growth efficiency ( $K_{2c} = P_c/A_c \cdot 100$ ) of feeding larval stage of *Musca domestica* and *Lucilia sericata*, raised at five different constant temperatures.

Temp. (°C)	Mean ± S.E. of cumulative growth efficiency	
	<i>M. domestica</i>	<i>L. sericata</i>
15	62.44 ± 0.65	69.79 ± 0.20
20	62.14 ± 0.73	80.20 ± 0.14
25	72.75 ± 0.43	77.52 ± 1.03
30	62.69 ± 0.99	90.63 ± 0.58
35	62.82 ± 0.74	82.04 ± 0.56

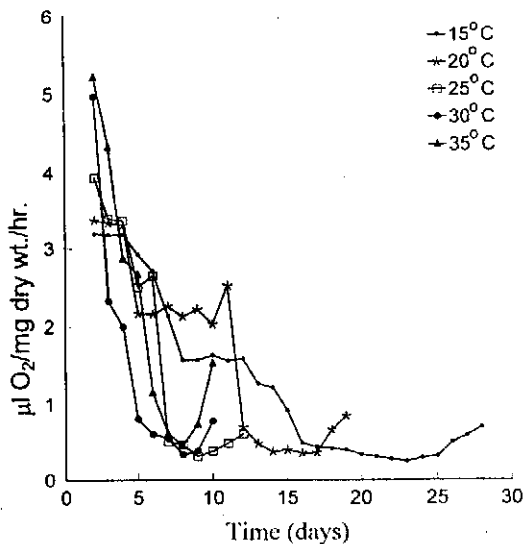


Fig. 6: The daily oxygen consumption ( $\mu\text{O}_2/\text{mg dry wt./hr.}$ ) of the different immature stages of *L. Sericata*, raised at five different constant temperatures.

It was recorded that as weights of feeding larvae increase there is a corresponding increase in oxygen consumed per individual. These results are in conformity with those obtained by Drummond and Chamberlain (1961), Gromysz-Kalkowska (1970), and Khalil (1994). The results also showed that the respiratory rates of the pupae follow a U-shaped curve. These results are similar to those reported by Needham (1950), Agrell (1953), Keister (1953), Roeder (1953); Park and Buck (1960), Guerra and Cochran (1970) and Jensen (1978). In this connection, Shaurub (1993) supposed that the descending and ascending arms of the U-shaped curve represent histolysis and histogenesis of tissues respectively.

The present study revealed values of  $Q_{10}$  not less than unity for all stages of both species at higher temperatures and not be very high at lower temperatures. This could suggest that the two species under investigation are adapted to high temperatures.

Also, the present study adds support to the view "as the weights increase, the consumption per unit weight decreases". Similar results have been reported by Drummond and Chamberlain, (1961); Guerra and Cochran (1970); Swift (1976); and Ziser and Nettles (1979).

It was reported that the instantaneous rate of assimilation of the feeding larvae of both species increased with increasing weight and age. These results may be explained according to the view of Hanski (1977) who stated that when the larvae are at the stage of the highest production rate, their metabolism is channeled to the synthesis of new tissues.

Also, it was recorded that larvae raised at high temperatures had an increased rate of assimilation. Williams and Richardson, (1984) suggested that instantaneous rate of assimilation increases with temperature due in part to increasing respiratory costs ( $R_i$ ) and since production ( $P$ ) also increases with temperature, consumption and digestive rates must increase.

Generally *L. sericata* exhibited high assimilation efficiency than *M. domestica*. This can be attributed to the Protein rich diet offered to *L. sericata* larvae. Also, Welch (1966) stated that animals with high assimilation efficiencies tend to be carnivorous, whereas those with lower assimilation efficiencies tend to be herbivore/detritivores. The present study supports the principle, that the detritivores *M. domestica* has a lower assimilation efficiencies than the carnivores *L. sericata*.

In addition, the present study revealed that the  $Q_{10}$  consumption and assimilation rate of both *M. domestica* and *L. sericata* larvae are age and temperature dependent. Hence, these age-markers could be used for accurate determination of postmortem interval in criminal investigations.

## References

- Agrell, I., 1953. The aerobic and anaerobic utilization of metabolic energy during insect metamorphosis. *Acta Physiol. Scand.*, 28: 306-335.
- Beamish, F.W.H., 1964. Respiration of fishes with special emphasis on standard oxygen consumption. II-Influence of weight and temperature on respiration of several species. *Can. J. Zool.*, 42: 177-188.
- Block, W. and P.J. Tilbrook, 1975. Respiration studies on the Antarctic collembola *Cryptopygus antarcticus*. *Oikos*, 26: 15-25.
- Cairns, S.C., 1978. Growth, respiration and the utilization of assimilated energy in the larvae of *Sericestus nigrolineata* (Coleoptera). *Oikos*, 31: 142-152.

- Davies, P.S., 1966. Physiological ecology of *Patella*. I. The effect of body size and temperature on metabolic rate. *J. Marine Biol.*, 46: 647-658.
- Dawson, W.R. and G.A. Bartholomew, 1956. Relation of oxygen consumption to body weight, and temperature acclimation in lizards *Uta stansburiana* and *Sceloporus occidentalis*. *Physiol. Zool.*, 29: 40-51.
- Dixon, M., 1951. *Manometric Methods, as applied to the measurement of cell respiration and other processes*. Third Edition. Cambridge Press, London, U.K., pp: 1-68.
- Drummond, R.O. and W.F. Chamberlain, 1961. Studies on respiration of cattle grubs. *Ann. Entomol. Soc. Amer.*, 54: 524-526.
- Doney, E.B., 1964. Acclimation to temperature in terrestrial isopods. II. Heart rate and standard metabolic rate. *Physiol. Zool.*, 38: 378-394.
- Drainager, J.N.R., 1956. Effects of temperature on the respiration of certain Crustacea. *Nature, Lond.*, 178: 930-931.
- Fromysz-Kalkowska, K., 1970. The influence of body weight, external temperature, seasons of the year and fasting on respiratory metabolism in *Polydesmus complanatus* L. (Diplopoda). *Folia Biol.*, 18: 311-326.
- Guerrea, A.A. and D.G. Cochran, 1970. Respiration during the life cycle of the face fly. *J. Econ. Entomol.*, 63: 918-921.
- Ganski, I., 1976. Assimilation by *Lucilia illustris* (Diptera) larvae in constant and changing temperatures. *Oikos*, 27: 288-299.
- Ganski, I., 1977. An interpolation model of assimilation by larvae of the blow fly *Lucilia illustris* (Calliphoridae) in changing temperatures. *Oikos*, 28: 187-195.
- Gealy, I.N., 1967. The energy flow through a population of soil Collembola, pp. 695-707. In K. Petruszewicz (ed.), *Proc. Working meeting on principles and methods of secondary productivity of terrestrial ecosystems*, Warszawa, Poland.
- Garbar, W.S., 1966. *General and Comparative Physiology*. Prentice-Hall, Englewood Cliffs, New Jersey, U.S.A.
- Gjansen, T.F., 1978. An energy budget for a field population of *Formica pratensis* (Retz.) (Hymenoptera: Formicidae). *Natura Jutlandica*, 20: 203-226.
- Granugo, M.S. and C.L. Prosser, 1959. Physiological and Biochemical adaptation of goldfish to cold and warm temperatures. I. Standard and active oxygen consumptions of cold-and warm-acclimatized goldfish at various temperatures. *J. Cell. Comp. Physiol.*, 54: 259-274.
- Gristner, M.L., 1953. Some observations on pupal respiration in *Phormia regina*. *J. Morph.*, 93: 573-587.
- Gristner, M.L. and J.B. Buck, 1961. Respiratory rate in relation to temperature, pp: 630-639. In *Physiology of Insecta* [M. Rockstein (ed.)], Vol. 3. Academic Press, New York, U.S.A.
- Hall, M.A., 1994. Functional ecological activities of some invertebrate soil communities in relation to pollution. Ph.D. Thesis., University of Tanta.
- Hejdukowski, R.Z., T. Prus and H. Zyromska-Rudzka, 1967. Elements of energy budget of *Tribolium castaneum* (Hbst.) in its developmental cycle, pp: 859-879. In *Secondary productivity of terrestrial ecosystems; Principles and Methods*. [K. Petruszewicz (ed.)] PWN, Warszawa-Krakow.
- Martin, P.J., 1991. Respiration of the ant *Leptothorax unifasciatus* (Hymenoptera: Formicidae) at individual and society levels. *J. Ins. Physiol.*, 37: 311-318.
- McFarland, W.N. and P.E. Pickens, 1965. The effects of season, temperature and salinity on the standard and active oxygen consumption of the grass shrimp, *Palaeomonetes vulgaris* (Say). *Can. J. Zool.*, 43: 571-585.
- Needham, J., 1950. The biochemistry of insect metamorphosis, 285 pp. In: *Biochemistry and Morphogenesis, Part 2*, Cambridge University Press, U.K.
- Otto, C., 1974. Growth and energetics in larval population of *Potamophylax cingulatus* (Steph.) (Trichoptera) in a south Swedish Stream. *J. Anim. Ecol.*, 43: 339-361.
- Park, H.D. and J. Buck, 1960. The relation of oxygen consumption to ambient oxygen concentration during metamorphosis of the blow fly, *Phormia regina*. *J. Ins. Physiol.*, 4: 220-228.
- Putman, R.J., 1977. Dynamics of the blow fly, *Calliphora erythrocephala*, within carrion. *J. Anim. Ecol.*, 46: 853-866.
- Roeder, K.D., 1953. *Insect Physiology*. John Wiley and Sons. Inc., New York. pp: 96-146.
- Shaurub, E.H., 1993. Effect of the carbamate insecticide propoxur, on O<sub>2</sub> consumption and CO<sub>2</sub> output during metamorphosis of *Chrysomya albiceps* (Wied.) (Diptera: Calliphoridae). *J. Egypt. Ger. Soc. Zool.*, 12: 43-53.
- Shishin'ova, M.D., 1986. Oxygen consumption during pupal development in *Musca domestica* L. (Diptera: Muscidae). *Ekologiya, Bulgaria*, 19: 44-49.
- Swift, M.C., 1976. Energetics of vertical migration in *Chaoborus trivittatus* larvae. *Ecology*, 57: 900-914.
- Tilbrook, P.J. and W. Block, 1972. Oxygen uptake in an Antarctic collembola *Cryptopygus antarcticus*. *Oikos*, 23: 313-317.
- Tribe, M.A. and K. Bowler, 1968. Temperature dependence of "standard metabolic rate" in a poikilotherm. *Comp. Biochem. Physiol.*, 25: 427-436.
- Umbreit, W.W., R.H. Burris and J.F. Stauffer, 1957. *Manometric Techniques*. Burgess publishing Co., Mineapolis, pp: 1-76.
- Waldbauer, G.P., 1968. The consumption and utilization of food by insects. *Adv. Ins. Physiol.*, 5: 229-288.
- Welch, H.E., 1968. Relationships between assimilation efficiencies and growth efficiencies for aquatic consumers. *Ecology*, 49: 755-758.
- Williams, H. and A.M. Richardson, 1984. Growth energetics in relation to temperature for larvae of four species of necrophagous flies (Diptera: Calliphoridae). *Australian J. Ecol.*, 9: 141-152.
- Ziser, S.W. and W.C. Nettles, 1979. The rate of oxygen consumption by *Eucelatoria* sp. in relation to larval development and temperature. *Ann. Entomol. Soc. Amer.*, 72: 540-543.