

<http://www.pjbs.org>

PJBS

ISSN 1028-8880

Pakistan Journal of Biological Sciences

ANSI*net*

Asian Network for Scientific Information
308 Lasani Town, Sargodha Road, Faisalabad - Pakistan

The Influence of GA₃, Kinetin and Ethylene on Seedling Growth, Nucleic Acids and Protein Content in Sweet Pepper (*Capsicum annuum*)

M.O. Basalah and Sher Mohammad

Department of Botany and Microbiology, College of Science,
King Saud University, Riyadh 11451, Saudi Arabia.

Abstract

A study was made of the influence of GA₃, kinetin and ethrel, individually or in combination, on nucleic acids (RNA and DNA), protein and seedling growth in sweet pepper (*Capsicum annuum*) cv. California Wonder. GA₃ treatment increased nucleic acids and proteins, markedly, as compared to untreated ones. The nucleic acids and protein content were closely related to the length of the seedlings. Kinetin was equally effective while in rest of the treatments, the nucleic acids content were lower than control and small increase was exhibited in protein content.

Introduction

Seedlings of most of the species produce ethylene as well as gibberellins. Gibberellins promote elongation of cells and inhibited by ethylene and kinetin (Pearce *et al.*, 1991, Sanchez-Bravo *et al.*, 1992). It has also been reported that GA₃ treatment will reverse the effect of ethylene on elongation (Stewart *et al.*, 1974). GA₃ treatment of cotton seeds induced synthesis of RNA, DNA and proteins in embryonic axis during period of germination (Khafagy and Mousa, 1982). Similarly GA₃ treatment of pea internodes promoted cell elongation, DNA and RNA in the tissue (Degani *et al.*, 1970), while DNA, RNA and protein synthesis are essential for GA₃ stimulated cell elongation (Broughton, 1969).

Present study was undertaken to see the effect of some of the growth regulators on nucleic acids and protein content in sweet pepper (*Capsicum annuum*) grown as vegetable crop in Saudi Arabia.

Materials and Methods

Sweet pepper (*Capsicum annuum* cv. California Wonder) seeds were purchased from the market. Seeds were sterilized with 1% hypochlorite solution and germinated in 9 cm plastic petri plates containing two, Whatman No. 1, filter papers. 10 ml distilled water or appropriate test solution (Table 1) was added to the petri plates. Fifteen seeds were transferred to each plate and incubated in dark at $\pm 25^{\circ}\text{C}$. Each treatment was replicated three times. After the desired treatment period (10 days in dark), the seedling lengths were measured and harvested. Samples were frozen for subsequent RNA, DNA and protein analysis. For RNA and DNA extraction, the method described by Ogur and Rosen (1950) was followed. Standard orcinol (Munro and Fleck, 1966) and diphenylamine (Burton, 1956) methods were used for estimation of RNA and DNA, respectively. Protein extraction was made following the

method of Torres and Tisserat (1980) and estimated by folin phenol reagent method (Lowry *et al.*, 1951).

Results

The results of the present study are depicted in Table 1. It is observed that the growth regulators, used individually or in combination, did not show primitive effect on seedling growth except for GA₃ and GA₃ + Kinetin which enhanced the growth, significantly, over control.

DNA and RNA contents increased, as compared to control, significantly, with GA, and kin. and non-significantly with ethrel treatment. Similarly protein content increased significantly with all the three growth regulators used. Combination of these growth regulators affected adversely the RNA content while an increase was observed in DNA for GA₃ + Kin and Eth + GA₃. The protein content increased over control, in all the combinations used.

Discussion

In the present study, it is revealed that GA₃ treated seedlings contained much more RNA, DNA and protein as compared to untreated ones. It is also observed that content of RNA, DNA and protein are closely related to the length of the seedlings. In many of the previous studies it is indicated that GA₃ promoted RNA, DNA and protein in germinating cotton seeds (Khafagy and Mousa, 1982) a cortical tissue of pea internodes (Broughton, 1969; Degani *et al.*, 1970). Leaf water soluble protein and nucleic acid contents increased in maize leaves by application GA₃ and kinetin (Dogra and Thukral, 1994) similar to present study. However, kinetin in combination with ethrel did not show much effect. The influence of cytokinins RNA and protein synthesis may affect the fluxes of ions a other material across the plasma membrane of leaf ce (Saeed, 1975). Increase in RNA content by GA₃ treatment is supported by previous studies which show that in barley aleurone layer incorporation into salt soluble RNA is enhanced by GA₃ and that GA₃ enhanced the synthesis of rapidly labeled RNA

Table 1: Effect of GA₃, Kinetin and Ethylene (Ethrel) alone or in combination, on growth, nucleic acids and protein content of sweet pepper (*Capsicum annuum*) seedlings. Concentrations of GA₃, Kin. and Eth. were 50 mg/L, 10 mg/L and 250 mg/L, respectively

Experimental solution	Length (cm/seedling)	RNA (Abs/seedling)	DNA (Abs/seedling)	Protein (mg/seedling)
Control	5.8	1.542	0.163	4.17
GA ₃	9.2	2.624	0.236	6.50
Kinetin	4.5	2.000	0.203	5.66
Ethrel	1.8	1.698	0.195	5.50
GA ₃ + Kin.	7.0	0.689	0.186	5.14
GA ₃ + Eth.	2.9	0.618	0.195	6.08
Kin. + Eth.	2.4	0.802	0.135	5.16
GA ₃ + Kin + Eth.	1.8	0.653	0.133	5.93

synthesis in barley aleurone layer (Ho and Varner, 1974). Direct evidence of GA₃ control of RNA, DNA and protein comes from many sources (Broughton, 1969). It is also reported that DNA synthesis is necessary for elongation of certain plant cells as the inhibition of DNA synthesis also inhibited the shoot elongation induced by GA₃ (Nitsan and Lang, 1966). Degani *et al.* (1970) studied that certain fraction of DNA synthesis is pre-requisite for hormone induced elongation or as another possibility, hormone-induced DNA is another manifestation of enhanced growth rather than pre-requisite. Similar mechanism may be operating in the seedlings used in the present study, as a close relationship is observed in length of seedling and nucleic acids and protein content.

References

- Broughton, W., 1969. Relations between DNA, RNA and protein synthesis and the cellular basis of the growth response in gibberellic acid-treated pea internodes. *Ann. Bot.*, 33: 227-243.
- Burton, K., 1956. A study of the conditions and mechanism of the diphenylamine reaction for the colorimetric estimation of deoxyribonucleic acid. *Biochem. J.*, 62: 315-323.
- Degani, Y., D. Atsmon and A.H. Halevy, 1970. DNA synthesis and hormone-induced elongation in the cucumber hypocotyl. *Nature*, 228: 554-555.
- Dogra, R. and A.K. Thukral, 1994. Proteins, nucleic acids and some enzyme activities in maize plants as affected by presowing seed treatment with steroids. *Indian J. Plant Physiol.*, 37: 164-168.
- Ho, D.T.H. and J.E. Varner, 1974. Hormonal control of messenger ribonucleic acid metabolism in barley aleurone layers. *Proc. Natl. Acad. Sci. USA*, 71: 4783-4786.
- Khafagy, E.Z. and A.M. Mousa, 1982. Nucleic acids and protein metabolic changes during germination of cotton seed. *Zeitschrift Pflanzenphysiologie*, 107: 321-328.
- Lowry, O.H., N.J. Rosebrough, A.L. Farr and R.J. Randall, 1951. Protein measurement with the Folin phenol reagent. *J. Biol. Chem.*, 193: 265-275.
- Munro, N.H. and A. Fleck, 1966. The Determination of Nucleic Acids. In: *Methods of Biochemical Analysis*, Glick, D. (Ed.). Vol. 14, Interscience, New York, USA., pp: 113-176.
- Nitsan, J. and A. Lang, 1966. DNA synthesis in the elongating nondividing cells of the lentil epicotyl and its promotion by gibberellin. *Plant Physiol.*, 41: 965-970.
- Ogur, M. and G. Rosen, 1950. The nucleic acids of plant tissues; the extraction and estimation of desoxypentose nucleic acid and pentose nucleic acid. *Arch. Biochem.*, 25: 262-276.
- Pearce, D.W., D.M. Reid and R.P. Pharis, 1991. Ethylene-mediated regulation of gibberellin content and growth in *Helianthus annuus* L. *Plant Physiol.*, 95: 1197-1202.
- Saeed, A.F.H., 1975. The distribution of mineral elements in *Xanthium pennsylvanicum*. Ph.D. Thesis, University of Sussex, UK.
- Sanchez-Bravo, J., A.M. Ortuno, M. Perez-Gilabert, M. Acosta and F. Sabater, 1992. Modification by ethylene of the cell growth pattern in different tissues of etiolated lupine hypocotyls. *Plant Physiol.*, 98: 1121-1127.
- Stewart, R.N., M. Lieberman and A.T. Kunishi, 1974. Effects of ethylene and gibberellic acid on cellular growth and development in apical and subapical regions of etiolated pea seedling. *Plant Physiol.*, 54: 1-5.
- Torres, A.M. and B. Tisserat, 1980. Leaf isozymes as genetic markers in date palms. *Am. J. Bot.*, 67: 162-167.