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# The Caper (Capparis ovata Desf.var. palaestina Zoh.) Culture in Turkey

Özlem Tonçer\* and Sezen TANSI\*\*

\*University of Dicle, Faculty of Agriculture, Field Crops Department-21280 Diyarbakir, Turkey \*\*University of Çukurova, Faculty of Agriculture, Field Crops Department-01330 Adana, Turkey

**Abstract:** *Capparis ovata* Desf. is one of the oldest known medicinal plants. It grown as wild in the arid, stony and poor soils of Diyarbakir region in Turkey. The experiments were conducted in open field conditions at University of Dicle, Faculty of Agriculture experiment area. The effect of applications GA<sub>3</sub>, H<sub>2</sub>SO<sub>4</sub> and different sandpaper on germination percentage of seed were studied and, the significant differences were found in terms of germination percentage of seed influenced by application. The maximum germination percentage (55%) was obtained in the seeds scarified by P32OA sandpaper thickness with GA<sub>3</sub> solutions of 400 ppm for 2 hour.

Key words: caper, culture, turkey

## Introduction

Caper (*Capparis ovata* Desf. var. *palaestina Zoh.*), a member of the family *Capparacea* is an important commercial and medicinal crop. In Turkey, it was exported 4.000 tons, accounting to \$ 12.000 (Anonymous, 1996a). Flower buds, fresh leaves, roots and fruits of caper are used in culinary uses such as pasta sauces, pizza, fish, meats and salads. Young shoots bearing immature small leaves may also be eaten as vegetable. Therapeutic effects of caper are anti-rheumatic, diuretic, tonic, anti-flatulence, hepatic stimulant, arteriosclerosis, vermifuges, kidney disinfectants, improving liver function. Derivates of the Bark are used in pharmaceutical preparations (Kontaxis, 1989). Flower buds of caper is processed into sauces, and exported to Europe, accounted \$ 2.5 million in 1996. Caper sauces is only produced in Turkey and Morocco. Thus, Turkey has determinative role in this market with the 50% share (Anonymous, 1996b).

The Southeast Anatolia Project (GAP) which is one of the greatest project of the world consists of Adiyaman, Diyarbakir, Gaziantep, Mardin, Siirt, Sanlurfa, Batman and Sirnak provinces in Turkey. Most experts identified soil erosion in the GAP region as the most urgent environmental problem. A major part of GAP area is subject to erosion, 72.3% to serious erosion and 38.6% to very serious erosion (Celik, 1994). The solutions is related to Flora. Vegetation is a consequence of soil fertility, at the point where vegetative cover is adequate.

Caper seed is difficult to germinate (Kontaxis, 1989). Fresh caper seeds germinate readily but only in low percentage. Dried seeds become dormant and are notably difficult to germinate and therefore require extra measures to grow (Alkire, 1998).

Considering potentially of caper for rural developing as a erosion control agent, new crop and new income sources, this study was aimed to cultivate caper growing under difficult environmental conditions in Southeast Anataolia Region.

#### **Materials and Methods**

Mature fruits of *Capparis ovata* were collected from wild plants in August and September from a population of Diyarbakir conditions in Turkey. The plants were identified as *Capparis ovata* Desf. var. *palaestina* Zoh. at Faculty of Science. Caper was grown under the Diyarbakir conditions prevailing terrestrial type climate which has hot and drought summer and hard winters. Seeds were sepapercentaged from fruit material, rinsed in top water, dried in shade and kept at room. The seed viability was determined by Tetrazolium test using a 400-seed as four replication of 100 seeds. The parts of the seed that are viable were become red, the non-viable parts remained white. Seed treatments were as follows:

- Seed were dipped in  $GA_3$  solutions 0 to 200, 300 and 400 ppm for periods of 0 to 1, 2, 3 and 24 h
- Seeds were scarified by P120A, P220A, P240A, P320A, P360A and P600A sandpaper (sand grain/cm<sup>2</sup>) then dipped in GA<sub>3</sub> solutions of 0 to 200, 300 and 400 ppm for periods of 0 to 1, 2, 3 and 24 h
- Seed were dipped in the concentrated  $H_2SO_4$  for 0, 20, 30 and 40 min then rinsed in top water and dipped in  $GA_3$  solutions of 0 to 200, 300 and 400 ppm for periods of 0 to 1, 2, 3 and 24 h

Seeds were sown at a depth of a few centimeters in open field seed bed in the month of March. The soil mix of 2:1:1 parts soil, sand and organic matter, respectively.

Germination was recorded at a week intervals within the experimental period. Pre-treatments effects were assessed by split split plot design. LSD test was carried out to test significances of all differences.

#### **Results and Discussion**

Tetrazolium test showed that the percentage of viable seed was more than 87%. The germination percentage of caper seeds showed differences according to concentrations of  $GA_3$  and application of different sandy paper (Table 1). The highest germination percentage was obtained in the seeds scarified by P320A sandpaper thickness with  $GA_3$  solutions of 400 ppm for 2 hour in first year. Also, in the second year, the highest germination rations were obtained from the control, P120A and P240A sandpaper treatments, followed by 400 ppm  $GA_3$  applications (Table 1).

The germination percentage of caper seeds showed differences according to concentrations of  $GA_3$  and dipping durations (Table 2). In the first year the highest germination percentage 35% was determined in the seeds dipped 400 ppm  $GA_3$  for 24 hour. The highest germination percentage was obtained from 400 ppm  $GA_3$  application for long duration such as 24 h is probably caused by thick seed testa (Table 2).

The germination percentage of caper seeds showed differences according to concentrations and dipping durations of  $GA_3$  and dipping durations of  $H_2SO_4$  (Table 3). In this first year, the highest percentage was determined in the seeds dipped 400 ppm  $GA_3$  non-applied  $H_2SO_4$ . In second year, the highest germination

GA <sub>3</sub> Dipping	Sandpaper Thickness	GA <sub>3</sub> Concentrations (ppm)									
Duration (h)		0		200		300		400			
		1.Year	2.Year	1.Year	2.Year	1.Year	2.Year	1.Year	2.Year		
0	Control	5.0d	13.0b	5.0d	13.0b	5.0d	13.0b	5.0d	13.0b		
	P120A	20.0a	8.0c	20.0a	8.0c	20.0a	8.0c	20.0a	8.0e		
	P220A	20.0a	17.0a	20.0a	17.0a	20.0a	17.0a	20.0a	17.0a		
	P240A	5.0d	10.0bc	5.0d	10.0bc	5.0d	10.0bc	5.0d	10.0bc		
	P320A	15.0b	8.0c	15.0b	8.0e	15.0b	8.0e	15.0b	8.0e		
	P360A	10.0c	20.0a	10.0c	20.0a	10.0c	20.0a	10.0e	20.0a		
	P600A	0.0e	0.0d	0.0e	0.0d	0.0e	0.0d	0.0e	0.0d		
1	Control	5.0d	13.0b	0.0e	21.0d	15.0c	13.0d	0.0d	27.0c		
	P120A	20.0a	8.0c	0.0e	29.0b	10.0b	8.0e	10.0c	29.0b		
	P220A	20.0a	17.0a	50.0a	29.0b	20.0b	42.0a	10.0c	25.0c		
	P240A	5.0d	10.0bc	15.0c	29.0b	20.0b	29.0b	0.0d	17.0d		
	P320A	15.0b	8.0c	10.0d	25.0c	40.0a	13.0d	20.0b	33.0a		
	P360A	10.0c	20.0a	0.0e	38.0a	0.0e	29.0b	30.0a	29.0b		
	P600A	0.0e	0.0d	0.0d	29.0b	15.0c	21.0c	10.0c	13.0e		
2	Control	5.0d	13.0b	0.0d	13.0e	0.0c	29.0a	10.0c	17.0e		
	P120A	20.0a	8.0c	5.0c	21.0c	5.0b	8.0d	0.0d	17.0e		
	P220A	20.0a	17.0a	20.0a	25.0b	0.0c	21.0c	0.0d	25.0c		
	P240A	5.0d	10.0bc	10.0b	21.0c	0.0d	25.0b	10.0c	33.0b		
	P320A	15.0b	8.0c	20.0a	17.0d	0.0c	21.0c	0.0d	21.0d		
	P360A	10.0c	20.0a	0.0d	17.0d	0.0c	25.0b	55.0a	17.0e		
	P600A	0.0e	0.0d	5.0c	21.0c	0.0c	21.0c	35.0b	38.0a		
3	Control	5.0d	13.0b	5.0d	17.0b	0.0d	13.0c	0.0g	25.0a		
	P120A	20.0a	8.0c	15.0c	8.0d	45.0a	8.0d	50.0a	8.0c		
	P220A	20.0a	17.0a	15.0c	17.0b	0.0d	17.0b	35.0b	21.0b		
	P240A	5.0d	10.0bc	20.0b	21.0a	5.0c	17.0b	5.0f	4.0d		
	P320A	15.0a	8.0c	0.0e	17.0b	0.0d	8.0d	15.0d	25.0a		
	P360A	10.0c	20.0a	30.0a	8.0d	0.0d	0.0e	20.0c	21.0b		
	P600A	0.0e	0.0d	0.0e	13.0e	10.0b	21.0a	10.0e	25.0a		
24	Control	5.0d	13.0b	0.0d	46.0a	0.0c	42.0a	35.0a	33.0d		
	P120A	20.0a	17.0a	10.0b	46.0a	0.0c	33.0b	0.0d	46.0a		
	P220A	20.0a	8.0c	0.0d	42.0b	0.0c	42.0a	20.0b	38.0e		
	P240A	5.0d	8.0c	5.0c	42.0b	15.0a	42.0a	20.0b	46.0a		
	P320A	15.0b	0.0d	5.0c	33.0c	0.0e	42.0a	15.0c	42.0b		
	P360A	10.0c	10.0bc	20.0a	42.0b	5.0b	42.0a	15.0c	42.0b		
	P600A	0.0e	20.0a	0.0d	42.0b	0.0c	33.0b	20.0b	42.0b		

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Table 1: Germination percentage of caper seeds as influenced by sandpaper application soaking duration and GA <sub>3</sub> concentration	Table	1:	Germination	percentage of	caper see	eds as influenced	d by	/ sandpaper	application	soaking	duration	and GA	concentration
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 Table 2: Germination percentage of caper seeds as influenced by Ga<sub>3</sub> concentration and soaking duration

 GA<sub>2</sub> (ppm)
 GA<sub>2</sub> Dipping Durations (b)

GA <sub>3</sub> (ppm)		GA <sub>3</sub> Dipping Durations (h)								
	0	1	2	3	24	Average				
1997										
0	5.0a	5.0b	5.0b	5.0a	5.0b	5.0				
200	5.0a	0.0c	0.0e	5.0a	0.0e	2.0				
300	5.0a	15.0a	0.0e	0.0b	0.0c	4.0				
400	5.0a	0.0c	10.0a	0.0b	35.0a	10.0				
Average	5.0	5.0	4.0	3.0	10.0	5.0				
1998										
C	13.0a	13.0c	13.0c	13.0c	13.0d	13.0				
200	13.0a	21.0b	13.0c	17.0b	46.0a	22.0				
300	13.0a	13.0e	29.0a	13.0c	42.0b	22.0				
400	13.0a	27.0a	17.0b	25.0a	33.0c	23.0				
Average	13.0	19.0	18.0	17.0	34.0	20.0				
S.D. (Int.) (	% 5): 1. Year: 2	.6, 2, Year: 2.4								

percentage was determine in the seeds dipped 200 ppm  $\mbox{GA}_3$  for 24 hour after treatment with  $H_2SO_4$  for 40 min and dipped 400 ppm GA\_3 for 24 h after treatment with  $\rm H_2SO_4$  for 20 min (Table 3). In other study, GA<sub>3</sub> and KNO<sub>3</sub> applications significantly increased the germination percentage (Toncer, 1999). In contrast to literature reviews, leaving seeds into  $H_2SO_4$  for 40 min. has no negative effect on germination (Orphanos, 1983). This can be attributed seed testa of C. ovata is more thicker then other

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Table 3: Germination percentage of caper seeds as influenced by soaking duration of H<sub>2</sub>SO<sub>4</sub> concentrations and soaking duration of GA<sub>3</sub>

GA <sub>3</sub> (ppm)	H₂SO₄ (min)												
(ppm)			0	1		2		3	}	2	4		
		1.Year	2.Year										
0	0	5.0b	13.0c	5.0b	13.0c	5.0b	13.0c	5.0b	13.0c	5.0b	13.Oc		
	20	15.0a	25.0a										
	30	5.0b	21.0b										
	40	5.0b	21.0b										
200	0	5.0b	13.0c	0.0c	21.0a	0.0a	13.0c	5.0b	17.0a	0.0b	46.0b		
	20	15.0a	25.0a	10.0b	17.0b	0.0a	25.0a	0.0b	4.0c	5.0a	38.0d		
	30	5.0b	21.0b	0.0c	17.0b	0.0a	13.0a	5.0a	13.0b	0.0b	42.0c		
	40	5.0b	21.0b	25.0a	21.0a	0.0a	17.0b	0.0b	17.0a	0.0b	50.0a		
300	0	5.0b	13.0c	15.0a	13.0c	0.0b	29.0a	0.0c	13.0c	0.0b	42.0b		
	20	15.0a	25.0a	5.0c	25.0a	0.0b	25.0b	15.0a	25.0a	0.0b	42.0b		
	30	5.0b	21.0b	10.0b	8.0d	20.0b	25.0b	5.0b	17.0b	0.0b	46.0a		
	40	5.0b	21.0b	0.0d	21.0b	0.0b	21.0c	5.0b	13.0c	5.0a	33.0c		
400	0	5.0b	13.0c	0.0c	17.0b	10.0b	17.0d	0.0b	25.0a	35.0a	33.0d		
	20	15.0a	25.0a	0.0c	29.0a	20.0a	21.0c	0.0b	8.0d	15.0a	50.0a		
	30	5.0b	21.0b	20.0a	13.0c	0.0c	29.0a	5.0a	21.0b	5.0c	46.0b		
	40	5.0b	21.0b	5.0b	17.0b	0.0c	25.0b	0.0b	13.0c	15.0b	42.0c		

(Toker et al., 1994). In both experiment year, when hard seed coat removes by scarifying, the caper seeds would performe subsequent germination. Orphanos (1983) reported that the seed coat is distributed trought microbial action during the winter so that the seed may germinate in spring. It is understood that dormancy of caper seed is imposed by its covering (Sozzi and Chiesa, 1995). Germination of seeds is influenced not only by temperature, but also by light, breaking of the seed coat. The presence of a hard seed coat that prevents absorption of oxygen or water. In nature, when enough rain falls to leach inhibitors from the seed (Bewley and Black, 1985). Also, the structure of caper seed and mucilage layer can prevent to germination. Therefore, available water in seed bed greatly enhanced germination percentage in our trials. In contrast to this study, other research was obtained low result in open conditions (Barbera et al., 1991).

There is evidence that soaking in GA<sub>3</sub> for 24 h generally produce possitive results for germination percentage in all treatments. Caper seed could only germinate if GA<sub>3</sub> was treated (Table 1). Also, GA<sub>3</sub> treatments after mechanical or chemical scarification gave quite well results. Thus, it is likely that GA<sub>3</sub> action speeds germination percentage by increasing O<sub>2</sub> gases uptake (Bewley and Black, 1985).

In general, all treatments used in this study have possitive effects on the germination percentage of caper seeds. Caper seedling after emergence grows very slowy, in this period, very rare and weak rooting systems needs regular irrigations, and it is necessary to keep soil at field capacity, if the irrigation is below the requirement soil surface dries rapidly, causing the wilting, over irrigation leads to decaying at seedlings, in both case young plants easily die. Also, keeping the soil moisture constant under field conditions enhanced and stimulated the germination. Consequently, caper will important role in the improving the export income of Turkey, also it will provide additional income of people living in the Southeast Anatolia. It is expected that erosion can be reduced by its cultivation, and it will bring the marjinal lands under relatively cropping.

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