

<http://www.pjbs.org>

PJBS

ISSN 1028-8880

**Pakistan
Journal of Biological Sciences**

ANSI*net*

Asian Network for Scientific Information
308 Lasani Town, Sargodha Road, Faisalabad - Pakistan

Effect of Growth Regulators on the Regeneration Potential of Two Sugarcane Cultivars SPF-213 and CPF-237

Farheen Niaz and Azra Quraishi

National Agricultural Research Institute, Park Road, Islamabad, Pakistan

Abstract: The study was conducted to optimize *in vitro* plantlets production from callus in two sugarcane cultivars. Sugarcane (*Saccharum officinarum* cv. SPF-213, CPF-237) embryogenic calli were induced from young leaves cultured on MS basal medium supplemented with 3.0 mg/l 2,4-D and 0.1 mg/l NAA. Three concentrations of two different growth regulators (6, benzylaminopurine and kinetin) were tested with and without NAA to compare their ability to induce regeneration from embryogenic calli. After 4 weeks of culture, the percentage of shoot induction was evaluated while after 6 weeks, the total number of shoots produced was checked. Medium containing BA @1.0 mg performed better than Kinetin with the highest percentage of shoot induction.

Key words: Sugarcane, *Saccharum officinarum*, *in vitro* culture, callus regeneration, BAP, Kn

Introduction

Sugarcane is one of the most important cash crop of Pakistan. The entire sugar requirement of Pakistan along with the multi billion sugar industry is solely dependent on the fate of this crop. Considerable difficulties have been encountered in the improvement of sugarcane through hybridization (Maqbool and Akhter, 2000). High yielding varieties with built in pest and disease resistance have been developed during the past five decades through conventional breeding and selection programme. However this method of breeding requires 10 to 15 years to release a variety because of greater genetic complexity of the crop. Tissue culture techniques play an important role in the genetic improvement of vegetatively propagated crops like sugarcane (Liu and Chen, 1984; Krishnamurthi, 1986; Siddique *et al.*, 1994). The *in vitro* plants regenerated from callus are capable of producing somaclonal variants for different traits like high yield, more sugar recovery, disease resistance, drought tolerance and early maturity etc.

Rapid callus formation has been obtained mostly from young leaves (Brisibe *et al.*, 1994; Farheen and Quraishi, 2002). To promote regeneration, callus was transferred to medium with different growth hormones (Irvine *et al.*, 1991).

The objective of the present research was to study the regeneration potential of two sugarcane varieties with supplementation of cytokinins and auxin in sugarcane germplasm on defined MS media. It has become increasingly clear that under the appropriate culture conditions, a great deal of genetic variability can be recovered in regenerated plants.

Materials and Methods

Sugarcane varieties SPF-213 and CPF-237 selected in this study were taken from sugarcane germplasm field at the National Agricultural Research Center (NARC) in collaboration with Co-ordinated programme on sugarcane during Dec-February. Embryogenic calli were induced from young leaves as described by Farheen and Quraishi (2000). Calli which were 3 months old were cultured on MS (Murashige and Skoog, 1962) medium supplemented with either BA, Kinetin at three different concentrations i.e. 0.1, 1.0, 2.0 with and without 0.2 mg NAA. Sucrose was added at the rate of 2% w/v. pH of the medium was adjusted to 5.7 before autoclaving. Total number of regenerated calli and average number of shoots were recorded after four weeks of culture. Each treatment was replicated 10 times. Cultures were incubated at 25±2°C under 2000 lux of light for 18/8 h light/dark photoperiod.

Results and Discussion

Regeneration is the ability of protoplast to develop the highly structured morphology of a whole plant. When calli of certain size

are transferred from culture medium to regeneration medium, the further regeneration process is not fundamentally novel. It follows either the organogenic or embryogenic pathway depending on genotype, donor tissue and culture medium used (Skarzhinskaya, 1996).

Different experiments were conducted in an attempt to find the optimum culture conditions for shoot regeneration from embryogenic calli. Vigorous calli were induced from leaf explants of *Saccharum officinarum* on MS media supplemented with 2.0 mg 2,4-D in SPF-213 while CPF-237 produced callus on media supplemented with 3.0 mg/l 2,4 D (Farheen and Quraishi, 2002). It was noted that auxin (2,4-D) had already shown a good response to callus induction and proliferation in both the cultivars tested (Chen *et al.*, 1988; Fitch and Moore, 1990; Oropeza and Garcia, 1996). This may be due to its chemical stability (Jha and Roy, 1982). The effect of different concentrations of BAP and Kinetin with and without NAA in MS were studied in terms of shoot initiation from callus of two cultivars. The type and concentration of growth regulators used in culture medium had a significant effect on induction of shoots. Medium containing BAP (1.0 mg/l) had the highest percentage of shoot initiation in all the concentrations in SPF-213 (Fig. 1).



Fig. 1: *Saccharum officinarum* cv. SPF-213 showing multiple shoots on MS medium containing BAP @ 1.0 mg.

Niaz and Quraishi: Effect of growth regulators on regeneration

Table 1: Effect of different concentration and combination of BA, NAA and Kin on shoot initiation from embryogenic calli of SPF-231

Growth regulators (mg/l)	No. of calli transferred	No. of calli regenerated	Average no. of shoots formed		Growth response	
			4 wk	6 wk		
BA	10	0.1	5	2	4	Slow
		1.0	9	18	24	Fast
		2.0	-	-	-	Stunted
BA + NAA	10	0.1 + 0.2	6	3	10	Slow
		1.0 + 0.2	8	13	22	Fast
		2.0 + 0.2	-	-	-	Stunted
Kin	10	0.1	3	-	2	Slow
		1.0	6	4	7	Medium
		2.0	-	-	-	Stunted
Kin + NAA	10	0.1 + 0.2	5	3	5	Medium
		1.0 + 0.2	7	15	18	Fast
		2.0 + 0.2	-	-	-	Stunted

Table 2: Effect of different concentration and combination of BA, NAA and Kin on shoot initiation from embryogenic calli of CPF-237

Growth regulators (mg/l)	No. of calli transferred	No. of calli regenerated	Average no. of shoots formed		Growth response	
			4 wk	6 wk		
BA	10	0.1	4	2	4	Slow
		1.0	9	18	24	Fast
		2.0	-	-	-	Stunted
BA + NAA	10	0.1 + 0.2	6	3	10	Slow
		1.0 + 0.2	8	13	22	Fast
		2.0 + 0.2	-	-	-	Stunted
Kin	10	0.1	3	-	2	Slow
		1.0	6	4	7	Medium
		2.0	-	-	-	Stunted
Kin + NAA	10	0.1 + 0.2	5	3	5	Medium
		1.0 + 0.2	8	15	18	Fast
		2.0 + 0.2	-	-	-	Stunted

their ability for callus proliferation but showed similar response towards regeneration. However addition of 0.2 mg NAA to the media did not alter shoot induction when combined with BA. Effectiveness of BAP alone for *in-vitro* shoot regeneration and multiplication was also reported in some other plants (Conover and Litz, 1987).

After 6 weeks of culture, the total number of shoots was noted. Highest number of shoots was observed on media containing BAP (1.0 mg/l) followed by Kinetin + NAA. Same results were reported by Chengalrayan *et al.*, 2000, Prajapati *et al.*, 2000). Minimum number of shoots was observed on medium containing Kinetin (0.1 mg/l) without NAA (0.2 mg/l) followed by BA. However no shoots were observed on media containing BA (2.0 mg/l) and Kinetin (2.0 mg/l) with and without NAA which confirmed that high concentration of growth hormones may hinder the regeneration ability (Siddiqui *et al.*, 1994).

It has surfaced that in media containing Kinetin, addition of NAA significantly increased the number of shoot production. However, with their positive effect on shoot production, these treatments still produced fewer shoots than BAP. This supports the fact that NAA may enhance the number of shoots produced depending upon the cytokinin used, but BAP alone remain the most effective treatment. It was also found that growth rate of callus multiplication and regeneration was dependent upon the concentration of growth hormones and type of explants used (Mujib, 1992). It has been concluded that plantlets formation could be increased after the addition of BAP alone. BAP and Kinetin in combination with NAA induced shoot formation in the same concentration with difference in number of shoots. However high concentration of growth regulators beyond 1.0 mg stunted the shoot formation.

It has also been noted that callus regeneration was also visible on media containing 2, 4-D in various concentration in both the cultivars but a sustained multiplication of callus was not possible on the same medium. It was earlier reported that 2, 4-D had the ability to perform to some extent the function of both auxins and cytokinin. However it is not yet known as to how it duplicate the

function (Bhattacharya *et al.*, 2000).

Similar results were also observed on MS media containing Kinetin with NAA (Table 1). It has been observed that BAP and Kinetin had similar intermediate effects. Earlier reports showed that combination of NAA with Kinetin promoted rapid regeneration from sugarcane callus (Irvine *et al.*, 1991). Similar results have been achieved in both cultivars (Table 2). Both cultivars varied in

Acknowledgment

The authors wish to thank Dr. Maqbool Akhtar for providing the source material.

References

- Bhattacharya, S., S. Dasgupta and P. Chatterjee, 2000. *In vitro* regeneration of plantlets from seedling explant of different species of Bauhinia L. *Pl. Tiss. Cult.*, 10: 103-109.
- Brisibe, E.A., H. Miyake, T. Taniguchi and E. Maeda, 1994. Regulation of somatic embryogenesis in long term callus cultures of sugarcane (*Saccharum officinarum* L.). *New Phytol.*, 126: 301-307.
- Conover, R.A. and R.W. Litz, 1987. Progress in breeding papaya with tolerance to papaya ring spot virus. *Proc. Ha. State. Hort. Soc.*, 91: 182-184.
- Chen, W.H., M.R. Davey, J.B. Power and E.C. Cocking, 1988. Control and maintenance of plant regeneration in sugarcane callus cultures. *J. Exp. Bot.*, 39: 251-261.
- Chengalrayam, K. and M. GalloMeagher, 2001. Effect of various growth regulators on shoot regeneration of sugarcane. *In vitro Cell Dev. Biol. Pl.*, 37: 434-439.
- Fitch, M.M.M. and P.H. Moore, 1990. Comparison of 2, 4-D and Picloram for selection of long term totipotent green callus cultures of sugarcane. *Plant Cell Tiss. Organ. Cult.*, 20: 157-163.
- Farheen, N. and A. Quraishi, 2002. Studies on Somatic Embryogenesis in sugarcane. *OnLine J. Biol. Sci.*, 2: 67-69.

Niaz and Quraishi: Effect of growth regulators on regeneration

- Irvine, J.E., G.T.A. Benda, B.L. Legendre and G.R. Machado, 1991. The frequency of marker changes in sugarcane plants regenerated from callus culture 2. Evidence for vegetative and genetic transmission, epigenetic effects and chimeral disruption. *Pl. Cell Org. Cult.*, 26:115-125.
- Jha, T.b. and S.C. Roy, 1982. Chromosomal behaviour in cultures of *Vicia fave*. *Cytologia*, 47: 465-470.
- Krishnamurthi, M., 1986. Sugarcane improvement through tissue culture process. *ISSCT*. XXIX: 23-28.
- Liu, M.C. and W.H. Chen, 1984. Tissue and cell culture an aid to sugarcane breeding-III. High sucrose and vigorously growing cell clone 71-489. *Taiwan Sugar*, 31: 77.
- Maqbool, A. and M.E. Akhtar, 2000. Problems and prospects of sugarcane research and development in Pakistan. *Pal. Sugar J.*, 15: 98-97.
- Mujib, A., 1992. Tissue culture studies on some bulbous ornamental plants. Ph.D. Thesis. University of Kalyani, India.
- Murashige, T. and F. Skoog, 1962. A revised medium for rapid growth and bioassay with tobacco tissue cultures. *Physiol. Pl.*, 15: 473-497.
- Oropeza, M., P. Guevara, E. Garcia and J.Z. Ramirez, 1996. Identification of somaclonal variants of sugarcane (*Saccharum* spp.) resistant to sugarcane mosaic virus via RAPD. *Pl. Mol. Biol. Rep.*, 13: 182-191.
- Prajapati, B.S., C.L. Patel, S.R. Patel and A.A. Patel, 2000. Regeneration of tissue culture plantlets through callus culture in sugarcane cultivar CoC671. *Indian J. Gene. Pl. Breed.*, 60: 255-257.
- Siddiqui, S.H., A. Khatri, A.I. Khan, M.A. Javed, N.A. Dhar and G.S. Nizamani, 1994. *In vitro* cultures; A source of genetic variability and an aid to sugarcane improvement. *Pak. J. Agric. Res.*, 15: 127- 133.
- Skarzhinskaya, M., M. Landgren and M. Glimelius, 1996. Production of intertribal somatic hybrids between *Brassica napus* L. and *Lesquerella fendleri* (Gray) Wats. *Theor. Appl. Genet.*, 94: 204-212.