http://www.pjbs.org



ISSN 1028-8880

# Pakistan Journal of Biological Sciences



© 2003 Asian Network for Scientific Information

# Effect of Brassica Vegetable Hosts on Biology and Life Table Parameters of Plutella xylostella under Laboratory Conditions

T.S. Syed and G.H Abro Department of Entomology, Sindh Agriculture University, Tandojam, Pakistan

Abstract: Different bracissca vegetables such as Brassica oleracea botrytis, B. oleracea capitata, B oleracea italica, B. napus, B. compestris, B. chenisis perkensis and Raphanus sativa were provided to newly hatched larvae of Plutella xylostella. The shortest and longest larval period of 9.45 and 10.95 days were recorded on B. oleracea botrytis and Raphanus sativa, respectively. Percent larvae pupating did not differ significantly, the lowest and highest percent survival to adult stage was recorded 58.3 and 76.7% on B. napus and B. oleracea italica. Whereas shortest and longest pupal period was recorded as 6.48 and 5.84 days on B.napus and B.oleracea capitata fed larvae. Pupal mass and fecundity of female showed that highest pupal mass and fecundity was recorded on B. oleracea fed larvae as compared to other Brassica vegetable crops. Similarly P. xylostella females preferred to lay more eggs on B.oleracea botrytis as compared with other hosts. Result also showed that females preferred to lay eggs on lower side as compared to upper side of the leaves. The life table parameters showed that net reproductive rate (Ro), was highest (89.71) when P.xylostella fed on B. olaracea botrytis, while the lowest Ro value of 26.77 was recorded when larvae were fed on B. napus. The intrinsic rate of increase (rm) and finite rate of increase (erm) were highest and lowest on B. olaracea botrytis and B.compestris, respectively.

Key words: Brassica, life table, vegetables, fecundity, host-plant

### INTRODUCTION

The diamondback moth, Plutella xylostella (L.) (Lepidoptera: Plutellidae) is the most destructive pest insect of cruciferous plants throughout the world. It has been recorded since 1746 (Harcourt, 1962) and is believed to have originated in the Mediterranean region (Harcourt, 1954) which is also the place of origin of some of the important crucifer crops (Tsunoda, 1980). Virtually, P. xylostella occurs wherever crucifer crops are grown, however, it has now been recorded from at least 128 countries or territories of the world and is believed to be the most universally distributed of all lepidoptera (CIE, 1968; Salinas, 1972; Lim, 1986; Talekar and Shelton, 1993). The annual cost of controlling P. xylostella on a worldwide basis is estimated to be around US\$ 1 billion (Talekar and Shelton, 1993; Lasota et al., 1996). Abro et al. (1992) have found P. xylostella a very serious pest on cruciferous vegetables in the Southern Sindh, Pakistan. P. xylostella feeds only on plants of cruciferae. Many of the plants of this family are cultivated as vegetables and oil seed crops. There are numerous crucifer plants which are not consumed by man, are considered as weeds and consumed by P. xylostella when its favoured hosts are absent and provide crucial link in maintaining P. xylostella populations (Talekar and Shelton, 1993 and Begum et al., 1996).

Brassica vegetables specially cabbage, Brassica oleracea var. capitata, cauliflower, B. oleracea var. botrytis and radish, Raphanis sativus are important vegetable crops of province of Sindh, Pakistan. The climate of southern Sindh (Hyderabad and Karachi regions) is mild and cruciferous vegetables are grown almost throughout the year, around towns and cities. P. xylostella has become a very serious problem and causes great economic loss to farmers. P. xylostella attack is some times so catastrophic that farmers are unable to save their crops even with the cocktails of most toxic chemicals and are compelled to uproot their cruciferous vegetables and grow other non host crops like onions, garlic and tomatoes to save themselves from complete economic wreck.

# MATERIALS AND METHODS

**Biological studies:** Studies on the effect of host plants on some biological parameters were carried out under laboratory conditions, at the department of Entomology, Sindh Agriculture University Tandojam during 1996-97. Different Brassica vegetable crops that is, cauliflower, *Brassica oleracea botrytis*; cabbage, *Brassicae oleracea capitata*; radish, *Raphanis sativa*; turnip, *Brassica napus*; Chinese cabbage, *Brassica chinesis perkensis*;

broccoli, Brassica italica and mustered, Brassica compestris were sown in a completely randomized block design on ridges. The leaf discs approximately 5.0 cm diameter of each host plant were kept in petridishes (10.0 cm dia) separately and 10 newly hatched larvae were introduced with fine hair brush in to each petridishes. The experiment was replicated five times. Fresh food was provided daily and old food removed till pupation. One day after pupation, pupal weight of randomly selected 25 pupae of each host plant were weighed on electrical balance (Ohas, Galexy 160). After adult emergence one male and female were paired and kept in glass chimney.(17.0 cm high) for egg laying. A small leaf of cauliflower was kept in vial containing water for egg laying and 10% (w/v) glucose solution in a beverage bottle lid impregnated on to cotton wool was provided to adult as food. Obsevations on egg laying were recorded daily till the end of oviposition or death of females.

Egg laying preference: For egg laying preference, newly emerged (10 pairs) male and female adults were released in a egg laying chambers. Fully expanded leaves of different host plants in a vial containing water were placed individually in a chamber for egg laying. Leaves were replaced daily and the eggs laid on subsequent days were considered as replications.

**Egg laying distribution:** The experiment was conducted on egg laying distribution. Newly emerged adult pairs were introduced in a cage, cauliflower leaves were provided for egg laying. Eggs laid on upper and lower leaf surfaces were counted daily. The experiment was replicated 12 times.

Life table parameters: To study the effect host plants on life table parameters of *P. xylostella* under laboratory conditions, the fresh laid eggs were collected from the culture and kept for hatching. After hatching, 50 neonate larvae (10.0 h old) were transferred with fine camel hair brush onto different host pants leaf discs in Petridishes (10.0 cm dia). Each petridish contained 10 larvae (5 replications). The leaf discs were changed daily till all larvae went into pupation. The emerged adults were paired and released in separate cages for egg laying. The fecundity of the females on subsequent days was noticed daily till all the females died. The life table parameters under laboratory conditions were constructed as suggested by Southwood (1978).

Net reproductive rate= Ro. =  $\sum$  Lxmx

 $\label{eq:logerout} \begin{array}{c} LogeRo \\ The \ intrinsic \ rate \ of \ population \ increase \ (rm) = ----- \\ T \end{array}$ 

The finite rate of population increase = (,rm).....

### RESULTS

Biological parameters: The effect of host plants on some biological parameters of Plutella xylostella showed the effect of different Brassica vegetables crops on larval and pupal period, percent larvae pupating and percent survival to adults (Table 1) .There was significant effect (F=173.01,DF=6,12; P<0.001) of host plants on larval development of P. xylostella. The shortest and longest larval period of 9.45 and 10.95 days was observed on cauliflower, Brassica oleracea botrytis and radish, Raphanus sativa, respectively. However, pupal periods were significantly different from each other. The percent larvae pupating did not differ significantly (F=1.61, df=12; p>0.05). The lowest and highest survival to adult stage was recorded as 58.3 and 76.7% on turnip, Brassica napus and broccoli, Brassia oleracea italics, respectively and it was significantly different (F=60.11, df=6,12; P< 0.001) from each other.

The effect of different host plants on pupal mass and fecundity are shown in Table 2. which indicated that there

Table 1: Biological parameters of *P.xylostella* on different host species in a detached leaf bioassay<sup>1</sup>

	Larval duration	Rate of larval	Pupal duration	Rate of survival
Host species	(days)	pupating/%	/d	to adult/%
B. oleracea botrytis	9.45±0.08b	88.3±1.7a	6.00±0.15e	75.0±0.58d
<ul> <li>B. oleracea capitata</li> </ul>	$9.63\pm0.10b$	78.3±4.4a	$5.84\pm0.15a$	63.3±1.116b
B. oleracea italica	10.68±0.11c	90.0±2.9a	$5.89\pm0.13b$	76.7±1.16d
B. napus	$10.57 \pm 0.19c$	78.3±6.0a	$6.48\pm0.15f$	58.3±0.58a
B. compestris	$10.90\pm0.15d$	78.3±1.7a	$5.94\pm0.19d$	60.0±1.16a
B. chinesis Pekinesis	9.13±0.18a	85.0±2.9a	$6.00\pm0.19e$	68.3±0.17c
R. sativa	$10.95\pm0.13d$	76.7±6.0a	$5.92\pm0.15c$	63.3±1.16
LSD 0.05	0.20	-	0.01	2.84

1) Mean  $\pm$  SE followed by the same letter within a same column are not significantly different from each other (P< 0.05; LSD) method)

Table-2: Effect of different host species on pupal mass and fecundity of \*P.xylostella\* under laboratory conditions<sup>1)</sup>

The state of the s					
Host	Pupal mass mg <sup>-1</sup>	Fecundity per female			
B. oleracea Botrytis	$6.08\pm0.13f$	212.3±23.8b			
B. oleracea capitata	5.89±0.11e	190.0±6.6b			
B. oleracea italica	$5.58\pm0.08d$	97.7±18.5a			
B. napus	5.50±0.09c	82.0±20.5a			
B. compestris	$4.95\pm0.15a$	118.7±19.3a			
B. chinesis Pekinesis	4.94±0.13a	83.7±11.6a			
R. sativa	5.04±0.16b	88.3±20.0a			
LSD 0.05	0.04	38.06			

 $<sup>^{(1)}</sup>$  Mean  $\pm$  SE followed by the same letter within a same column are not significantly different from each other (P< 0.05; LSD)

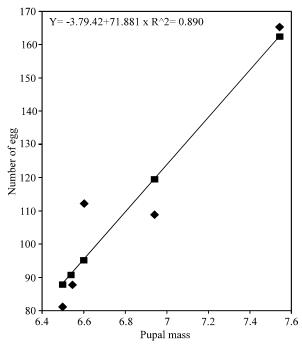


Fig. 1: Linear Regression between the eggs per female and average pupal mass of *P. xylostella* when larvae were reared on different host plants

was significant (F=13.75, df=6,54, P<0.01) effect of host species on pupal mass and the highest pupal mass was recorded from larvae reared on cauliflower (6.08 mg) followed by cabbage (5.89 mg). While the lowest pupal mass was found of larvae reared on Chinese cabbage (4.94 mg). Similarly, the highest fecundity of females was recorded when larvae were reared on cauliflower (212.3) followed by cabbage (190.0) which was significantly (F=8.82, df=6,12; P<0.01) higher than larvae reared on remaining host plants. The lowest fecundity of 82.0 eggs per female was recorded when larvae were fed on turnip as host. The correlation studies carried out between pupal mass and fecundity also indicated a highly significant (P<0.01) relationship (Fig. 1).

**Egg laying preference:** Host plant preference for egg laying by *P. xylostella* (Table 3) indicated that significantly (F=8.55, df=6,12; P<0.01) more eggs were laid on cauliflower leaf followed by cabbage indicating that cauliflower is preferred host of *P.xylostella* as compared with other hosts for egg laying.

**Egg laying distribution:** The egg laying distribution of *P. xylostella* on cauliflower leaves under laboratory conditions showed that more eggs were laid on lower leaf surface when compared with upper leaf surface, however, the difference was statistically non significant (Table 4).

Table 3: Egg-laying preference of *P.xylostella* females on different host species in a detached-leaf bioassay<sup>1)</sup>

Host	Eggs	%Eggs	
B.oleracea botrytis	803.3±232.1b	47.7	
B.oleracea capitata	233.3±23.9a	13.8	
B.oleracea italica	154.0±10.0a	9.1	
B.napus	127.3±49.1a	7.5	
B.compestris	133.3±14.1a	7.9	
B.chinesis pekinesis	126.7±43.6a	7.57	
Raphanus sativa	106.7±35.9a	6.3	
Total	1685.6	100	
LSD 0.05	132.04	-	

1) Means $\pm$ SE followed by the same letter within a column are not significantly different from each other (P< 0.05; LSD)

Table 4: Egg distribution of *P. xylostella* on cauliflower leaves under laboratory condition

laboratory condition					
Treatment	n	$X \pm SE$ eggs	x %	t-a	P
Lower leaf surface	12	300.0+49.4	53.1	0.00	>0.05
Upper leaf surface	12	265.0+61.4	46.9	0.23	>0.05

a = Paired two tailed "t" test

Table 5: Effect of host plants on the life-table parameters of *P. xylostelia* under laboratory conditions

Host plants	F*	Ro	T(d)	rm	$e^{rm}$
B. oleracea botrytis	212.3+22.8b	89.71	18.75	0.2398	1.27
B. oleracea capitata	190.0+6.6b	73.96	18.88	0.2288	1.25
R. sativa	88.3+20.0a	45.54	21.03	0.181	1.199
B. napus	82.0+20.5a	26.77	20.54	0.160	1.173
B. compestris	118.7+19.3a	31.797	21.69	0.159	1.173
B. chinesis pekinesis	83.7+11.6a	31.79	19.697	0.176	1.192
B. oleracea italica	98.7+18.5a	39.95	21.67	0.170	1.185

<sup>\* =</sup> Mean + S.E

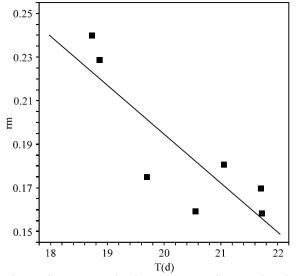


Fig. 2: Linear regression between rm and generation time of *P. xylostella* under laboratory conditions

**Life table parameters:** The life table parameters of *P. xylostella* on different host plants are shown in Table 5. The net reproductive rate (Ro) which is a product of mean

p = figures are statistically non-significant (P>0.05)

n = Replications

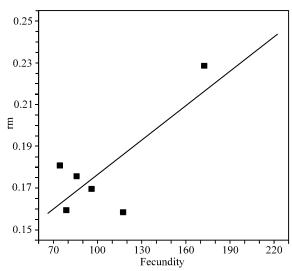


Fig. 3: Linear regression between rm and fecundity of P. xylostella under laboratory conditions

total fecundity, survival rate and sex ratio was the highest (89.71) expected females per female obtained with cauliflower as a host plant, while the lowest Ro value of 26.77 expected females per female was recorded when *P. xylostella* larvae were fed on turnip. There was some variation in the mean generation time T(d) of *P. xylostella* fed on different host plant. This value was (21.69 days) when individuals were fed mustard; T(d) was shorter (18.75) on cauliflower. The intrinsic rate of increase (rm) and the finite rate of increase (e<sup>rm</sup>) were the highest (0.2398 and 1.27 respectively) when *P. xylostella* were fed cauliflower and lowest (0.159 and 1.173 respectively) on mustard as a host plant.

Multiple linear regression analysis between mean generation time (T) and mean total fecundity (independent variable) yielded R2=0.839 (P=0.05). The following equation was derived for *P. xylostella* at laboratory temperature rm=0.378+0.000146 F-0.0116G. This model indicated that 83.9% of the variance of the obtained rm values could be attributed to total fecundity and generation time. A higher total fecundity value would have a positive effect on the obtained rm value, where as, a higher value of generation time would have a negative effect on the rm value. (Fig. 1 and 2).

## DISCUSSION

Life-cycle of diamondback moth varies considerably depending upon various factors such as the environmental conditions under which it develops and host plants (Ooi, 1986; Ko and Fang, 1979; Biever and Boldt, 1971; Shelton *et al.*, 1991). This difference in

developmental time may be due to rearing of *P. xylostella* on different host plants. Differences in male and female developmental times and pupal weight observed in present studies were also reported by various workers (Kanervo, 1936; Umeya and Yamada, 1973).

The growth and development of P. xylostella varied considerably on different hosts plants in present study which may be due to presence of nutritional and phagostimulant factors in a most appropriate ratio for optimal growth and development of it in some of its host plants as compared with others. In present study cauliflower and cabbage were found to be better hosts for P. xylostella as larval period was completed in shorter time when compared with other hosts. Chand and Choudhary (1977) noted that P. xylostella attacked all the cruciferous plants, however, it exhibited a marked preference for cauliflower and cabbage. These two plant species possess fleshy succulent leaves that provide both olfactory and gustatory stimuli. Similarly, Singh and Singh (1982) studied the influence of various cruciferous host plants on survival and development of P. xylostella and found that pest completed its larval and pupal development in the shortest time on cauliflower. Harcourt (1957) studied the larval population of P. xylostella on various crucifers growing in the field and found the highest pest population on collards, cabbage and cauliflower crops as compared with other hosts. Ramachandran et al. (1998) also found significant differences in biological parameters of P. xylostella feeding on different Brassica spp. leaves. Present studies also show the highest P. xylostella survival when fed cauliflower followed by cabbage, almost similar observations were also recorded by other workers (Dube and Chand, 1977; Singh and Singh, 1982).

Highest number of eggs were laid by female P. xylostella when larvae were reared on cauliflower followed by cabbage (Table 4). Similarly, pupal weight of P. xylostella was also higher for those insect, which were fed cauliflower and cabbage in larval stage. Fecundity of an insect depends apart from other factors on pupal size (Wan, 1970). The fecundity-pupal weight regression equations obtained in present studies indicated that P. xylostella fecundity increased at a rate of 71.88 and 85.61 eggs for each milligram increase in pupal weight, regardless of the host on which female was reared. A correlation carried out between pupal weight and fecundity showed a significant positive relationship between two factors (Fig. 3). A significant correlation was noted between the fecundity of adults and pupal weight or length of P. xylostella (Wan, 1970). Similarly, Yamada and Umeya (1972) concluded that fecundity is determined by pupal size. The linear relationship of fecundity to body

weight or size of pupa or both, has also been observed in other insects. Miller (1957) noted that fecundity of spruce budworm, Choristoneusa fumiferana Clemens, could be estimated on pupal weight or size, because both factors are related linearly. Williams (1963) observed that the correlation between pupal weight and fecundity was greater than that between pupal length and fecundity in Chilo sacchariphagus Bojer. Begum et al. (1996) showed that female P.xylostella grown on wild crucifers (Rorippa indica and Lipidium virginicum) was less fecund than those reared on cabbage. Moreover, fecundity, longevity, flight activity and morphological characters of adult were positively correlated with pupal weight.

In the present study more eggs were recorded on the lower leaf surface than upper leaf surface (Table 4). This difference in egg laying behaviour may be due to difference in leaf morphology including leaf waxes, contour (grooves) etc. supporting egg impaction on leaf surface. These findings are in agreement with Robertson (1939).

P. xylostella feeds on a wide range of cruciferous host plants (Harcourt, 1957; Dube and Chand, 1977; Singh and Singh, 1982; Siemens and Mitchellolds, 1996; Ramachandran et al., 1998). The effect of host plants on development and reproduction of P. xylostella vary (Wakisaka et al., 1992; Salas et al., 1993; Begum et al., 1996). Van Lanteren and Noldus (1990) have stated that shorter developmental times and greater total oviposition (fecundity) on a host reflected the suitability of the host plant. In present study, P. xylostella fed on cauliflower and cabbage showed higher intrinsic rates of increase (rm) resulting from faster development (shorter generation time), higher survivorship and higher fecundity rates. These two host plants are presumably more suitable hosts as compared to other hosts. Wakisaka et al. (1992) studied the rm value of P. xylostella on different host plants and found that 'rm' value ranged between 0.2778 and 0.1362 respectively on broccli and a wild crucifer, Capsella bursa-postoris. Salas et al. (1993) investigated the life table parameter of P. xylostella on different host plants and observed that the highest rm was on cauliflower. Also, there are many other studies where host plants have affected biology and life table parameters of many insects/organisms (Root and Olson, 1969; Bessin and Reagan, 1990; Fouly et al. 1995; Tsai and Wang, 1996).

### REFERENCES

Abro, G.H., R.A. Soomro and T.S. Syed, 1992. Biology and behaviour of diamond back Moth *Plutella xylostella* (L). Pakistan. J. Zool., 24: 7-10.

- Begum, S., T. Ritsuko, K. Fujisaki and F. Nakasuji, 1996. The effects of wild cruciferous host plants on morphology, reproductive performance and flight activity in the diamondback moth, *Plutella xylostella* (Lepidoptera: Yponomeutidae). Res. Popul. Ecol., 38: 257-263.
- Bessin, R.T. and T.E. Reagan, 1990. Fecundity of sugarcane borer (Lepidoptera: Pyralidae), as affected by larval development on gramineous host plants. Environ. Entomol., 19: 635-639.
- Biever, K.D. and P.E. Boldt, 1971. Continuous laboratory rearing of the diamondback moth and related biological data. Ann. Entomol. Soc. Am., 64: 651-655.
- Chand, P. and R. Choudhry, 1977. Patterns of insect plant relationships determining susceptibility of food plants in the diamondback moth, *Plutella xylostella* (L.) Cuyrtis. Mysore. Agric. Sci., 11: 547-549.
- CIE, 1968. Distribution maps of pests: *Plutella xylostella* (L.) (Maculipennis Curt). Common Wealth Inst. of Entomol. CAB, Series A. Map No.32.3.
- Dube, R.B. and P. Chand, 1977. Effects of food plants on the development of *Plutella xylostella* (L.) (Lepidoptera: Plutellidae). Entomon., 2: 139-140.
- Fouly, A.H., M.N. Abou-Setta and C.G. Childers, 1995. Effects of diet on the biology and life tables of *Typhlodormalus peregrinus* (Acari: Phytoseiidae). Environ. Entomol., 24: 870-874.
- Harcourt, D.G., 1954. The biology and ecology of the diamondback moth, *Plutella maculipennis*, Curtis, In Eastern Ontario, Ph.D. Cornell Univ., Ithaca, N.Y. pp: 107
- Harcourt, D.G., 1957. Biology of the Diamondback moth, Plutella xylostella (Curt.) (Lepidoptera: Plutellidae), in Eastern Ontario. II. Life-History, Behaviour and Host Relationships. Can. Entomol., 89: 554-564.
- Harcourt, D.G., 1962. Biology of cabbage caterpillars in Eastern Ontario. Proc. Entomol. Soc. Ont., 93: 67-75.
- Kanervo, V. 1936. The diamondback moth (P. maculipennis Curt.) as a pest of cruciferous plants in Finland Valt. Maatalousk. Julk., 86: 1-86.
- Ko, L.T. and J.L. Fang, 1979. Studies on the biology of the diamondback moth, *Plutella xylostella* L. Life history, annual generations and temperature relations. Acta Entomol. Sin., 22: 310-319.
- Lasota, J.A., A.M. Shelon, J.A. Bolognese and R.A. Dybas, 1996. Toxicity of avermectin to diamondback moth (Lepidoptera: Plutellidae) populations: Implications for susceptibility monitoring. J. Econ. Entomol., 89: 33-38.
- Lim, G.S., 1986. Biological control of diamondback moth. In: Talekar, N.S., Griggs, T.D., (eds.) Diamondback moth management. Proc. 1st Int. Workshop, Taiwan: AVRDC., pp. 159-172...

- Miller, C.A., 1957. A technique for estimating the fecundity of natural populations of the spruce budworm. Can. J. Zool., 35: 1-3.
- Ooi, P.A., 1986. A diamondback moth in Malaysia. In: N.S. Talekar and T.D. Griggs (Ed.). Proc. 1st Intern. Workshop on Diamondback moth management AVRDC. Taiwan, pp: 25-34.
- Ramachandran, S., G.D. Buntin, J.N.All and P.L. Raymer, 1998. Diamondback moth (Lepidoptera: Plutellidae) Resistance of *Brassica napus* and *B. oleraceae* Lines with differing leaf characteristics. J. Econ. Entomol., 91: 987-992.
- Robertson, P.L., 1939. Diamond back moth investigation in New Zealand. N. Z. J. Sci. Technol., 20: 330-364.
- Root, R.B. and A.M. Olson, 1969. Population increase of the cabbage aphid, Brevicoryne Brassicae, on different host plants. Can. Entomol., 101: 768-773.
- Salas, M.D., B. Mendoza, E. Salazar and V.M. Rivera, 1993.
  Supervivencia y Reproduction de la Palomilla Dorso de Diamante en Cruciferas. Turrialba, 43: 242-246.
- Salinas, P.J., 1972. Studies on the ecology and behaviour of the larvae of *Plutella xylostella* (Linnaus) (Lepidoptera: Plutellidae). Ph.D. Thesis, University of London, London, pp. 356.
- Shelton, A.M., R.J. Cooley, M.K. Kroening, W.T. Wilsey and S.D. Eigenbrode, 1991. Comparative Analysis of Two Rearing Procedures for Diamondback moth (Lepidoptera: Plutellidae). J. Entomol. Sci., 26: 17-26.
- Siemens, D.H. and T. Mitchelloids, 1996. Glucosinolates and herbivory by specialists (Coleoptera: Chrysomelidae, Lepidoptera: Plutellidae): Consequences of Concentration and Induced Resistance. Environ. Entomol., 25: 1344-1353.
- Singh, S.P. and D. Singh, 1982. Influence of cruciferous host plants on the survival and development of *Plutella xylostella*. L. J. Res. Punjab Agric. Univ., 19: 100-104.
- Southwood, T.R.E., 1978. Ecological Methods. 2nd Edn., Chapman and Hall, London, Pages: 524.
- Talekar, N.S. and A.M. Shelton, 1993. Biology, ecology and management of the diamondback moth. Ann. Rev. Entomol., 38: 275-301.

- Tsai, J.H. and K. Wang, 1996. Development and reproduction of *Bemisia argentifolii* (Homoptera: Aleyrodidae) of Five Host Plants. Environ. Entomol., 25: 810-816.
- Tsunoda, S., 1980. Eco-physiology of wild and cultivated forms in Brassica and allied genera. In: Brassica crops and wild allies. Biology and breeding, (Eds) S. Tsunoda, K. Hinata and C. Gomez-Campo, PP-109 Scientific Societies Press, 20. Tokyo; Japan, pp. 354.
- Umeya, K. and H. Yamada, 1973. Threshold temperature and thermal constants for development of the diamondback moth, *Plutella xylostella* L. with reference to their local differences. Jpn.J. Appl. Entomol. Zool., 17: 19-24.
- Van Lanteren, J.C. and L.P.J.J. Noldus, 1990. Whitefly Plant Relationships, Behavioural and Ecological Aspects. In: White Flies: Their Bionomics, Pest Status and Management, Gerling, D. (Ed.). Intercept Ltd., Hamshire, England, pp. 47-89.
- Wakisaka, S., R. Tsukuda and F. Nakasuji, 1992. Effects of natural enemies, rainfall, temperature and host plants on survival and reproduction of the diamondback moth and other crucifer pests. In: Talekar, N.S. (ed) diamondback moth and other crucifer pests. Proc. 2nd Intern. Workshop AVRDC Taiwan, pp. 15-36.
- Wan, M.T.K., 1970. The bionomics and control of the diamondback moth, *Plutella xylostella* L. (*P. maculipennis* Curt.) (Lepidoptera: Plutellidae) in Sarawak (Malaysian Borneo). Sarawak. Mus. J., 18: 377-378.
- Williams, J.R., 1963. The reproduction and fecundity of the sugarcane stalk borer, *Plutella saccharipagus* Baj. (Lepidoptera: Crambidae) Proc. Int. Soc. Sugarcane Technol., 11: 611-625.
- Yamada, H. and K. Umeya, 1972. Seasonal changes in wing length and fecundity of the diamondback moth, *Plutella xylostella* (L.) Jpn. J. Appl. Entomol. Zool., 16: 180-186.