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## Nitrification Inhibition in Soil and Ecosystem Functioning – An Overview

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**Abstract:** Form ( $\text{NH}_4$  or  $\text{NO}_3$ ) and availability of N has significant implications to the functioning and sustainability of agroecosystems. Most of the fertilizer nitrogen (N) applied to agricultural soils is in the form of  $\text{NH}_4$  or  $\text{NH}_4$ -forming fertilizers. This form of N is rapidly oxidized to  $\text{NO}_3$  by nitrifying microorganisms leading to significant losses of N through  $\text{NO}_3$ -leaching and denitrification. Both denitrification and  $\text{NO}_3$ -leaching have environmental implications and economic concerns. Strategies have therefore been sought to regulate the process of nitrification leading to its complete or partial inhibition. Indeed, climax ecosystems are developed in such a way that the process of nitrification is already fairly inhibited. This paper presents an overview on: I) the process of nitrification, ii) microorganisms involved, iii) the implications of nitrification and nitrification inhibition to ecosystem functioning and finally iv) the methods to inhibit nitrification.

**Key words:** Fertilizer nitrogen, agroecosystems, soil

### Introduction

Nitrogen (N) is one of the most widely distributed elements in nature, with atmosphere as the main reservoir. Of the total N found in nature, 99.96% is present in the atmosphere. Of the remaining 0.04%, biosphere contains only 0.005%. In spite of the small proportion found in living beings, N is most often the limiting nutrient for crop production since only a fraction of atmospheric  $\text{N}_2$  is made available to the plants through biological nitrogen fixation. However, introduction and use of chemical nitrogenous fertilizers has resulted in substantial increases in crop yields. According to statistics by FAO (2001), about 42 million tons of fertilizer N is being used annually on a global scale for the production of 3 major cereal crops i.e., wheat, rice and maize (17, 9 and 16 million tons, respectively). However, fertilizer N is not an unmixed blessing since the crop plants are able to use only about 50% of the applied fertilizer N, while 25% is lost from the soil-plant system through leaching, volatilization and denitrification etc. causing an annual financial loss of ca 3 billion US\$. Incidentally, the losses are similar for the 3 crops mentioned above but twice as much from rice compared to wheat and maize when computed on the basis of per unit of fertilizer N applied.

The soil, which is the major source of plant-available N, accounts for a minute fraction of the lithospheric N, while only a very small proportion is directly available to plants as  $\text{NH}_4$  and/or  $\text{NO}_3$ . Although most plants are able to use both  $\text{NH}_4$ -N and  $\text{NO}_3$ -N, the later is the predominant source of N for plants grown under arable/terrestrial situations. This is primarily because of compulsion rather than preference as most of the  $\text{NH}_4$ -N is rapidly nitrified under optimum conditions of temperature, moisture and

aeration. As a result,  $\text{NH}_4$ -N is available to the plants only for a limited time period and most of the time, plant roots are confronted with  $\text{NO}_3$ -N as a source of N.

Nitrification is a key process in managed agricultural ecosystems because the conversion of  $\text{NH}_4$  to  $\text{NO}_3$  can lead to substantial loss of agricultural N (both applied as chemical fertilizers or present as native soil N) by leaching and/or denitrification. The loss of fertilizer N is of concern not only because of economic reasons but also due to the pollution potential of different N forms. Some of the adverse environmental effects of excessive use of nitrogenous fertilizers include I) methemoglobinemia in infants due to  $\text{NO}_3$  and  $\text{NO}_2$  in waters and food, ii) cancer due to secondary amines, iii) respiratory illness due to  $\text{NO}_3$ , aerosols,  $\text{NO}_2$  and  $\text{HNO}_3$ , iv) eutrophication due to N in surface waters, v) material and ecosystem damage due to  $\text{HNO}_3$  in rain water, vi) plant toxicity due to high levels of  $\text{NO}_2$  and  $\text{NH}_4$  in soils, vii) excessive plant growth due to more available N and viii) depletion of stratospheric ozone due to NO and  $\text{N}_2\text{O}$ . It is because of these concerns that concerted efforts have been and are being made to reduce the use or increase the efficiency of fertilizer N uptake by crop plants.

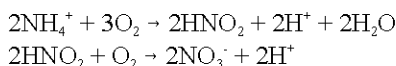
Multi-disciplinary approaches have been identified and put to practice over the past few decades to increase use efficiency and decrease the losses of fertilizer N. These include a) breeding crop varieties with higher fertilizer use efficiency, b) management of fertilizer N, c) use of chemicals inhibitory to specific N transformation processes like urea hydrolysis and nitrification i.e., urease and nitrification inhibitors, respectively, to slow the process of  $\text{NH}_4$  formation from urea and to regulate the process of  $\text{NO}_3$  formation, d) supplementation/integration

of fertilizer N with organic manures e.g., green manures. These approaches applied singly or in combination, have certainly helped alleviate the problems arising as a result of fertilizer N use. Fertilizer management through improved formulations, mode and time of application and placement etc. has also been found helpful to overcome some of the problems. Likewise, a large variety of chemicals have been tested as potent inhibitors of specific N transformation processes including urea hydrolysis and nitrification. With these inhibitors, not only use efficiency of fertilizer N is enhanced, but a significant reduction occurs in losses due to NH<sub>3</sub> volatilization, denitrification and NO<sub>3</sub> leaching. Currently, increasing attention is being paid to the development of controlled-release and stabilized fertilizers that seem more efficient and environment friendly (Trenkel, 1997).

Amongst different N cycle processes vis-à-vis ecosystem functioning and environmental concerns, the process of nitrification stands out to be the most important process. Nitrification inhibition could lead to: i) increased rhizospheric microbial activities including biological N<sub>2</sub> fixation and production of phytohormones, ii) enhanced mineralization of native soil N thus decreasing dependence on chemical fertilizers, iii) increased efficiency of fertilizer N through decreased losses via denitrification and NO<sub>3</sub> leaching and iv) greater photosynthate partitioning to the rhizosphere thus enriching the soil with organic matter. It is important, therefore, to develop an understanding of microbes responsible for nitrification, factors affecting nitrification, methods to regulate the process and its implications to ecosystem functioning.

### Microbiology of nitrification

The biological oxidation of ammonia to nitrate, termed as nitrification, is a two-step process mediated by autotrophic bacteria. Ammonia is first oxidized to NO<sub>2</sub><sup>-</sup> that in turn is oxidized to NO<sub>3</sub><sup>-</sup> as follows:



The source of NH<sub>4</sub><sup>+</sup>-N could be soil organic matter (mineralization by soil microorganisms) and chemical fertilizers. In the case of soil, organic N is used by the ammonifiers, while chemical fertilizers either contain NH<sub>4</sub><sup>+</sup> as such or its precursors.

Several genera and species of ammonium and NO<sub>2</sub><sup>-</sup> oxidizing heterotrophs including fungi (*Aspergillus flavus*, *Neurospora crassa*, *Penicillium* sp.), actinomycetes (*Streptomyces* sp., *Nocardia* sp.) and bacteria (*Arthrobacter* sp., *Azotobacter* sp., *Pseudomonas fluorescens*, *Aerobacter aerogenes*, *Bacillus megaterium*,

*Proteus* sp.) have been reported. However, autotrophic nitrifiers are generally considered as the main organisms responsible for most of nitrification. Ammonium oxidizing autotrophs include *Nitrosomonas*, *Nitrosolobus* and *Nitrosospira*. These organisms have been isolated from a variety of soil environments with ubiquitous distribution. Nitrite produced by the ammonium oxidizing autotrophs is rapidly oxidized to NO<sub>3</sub><sup>-</sup> by *Nitrobacter* species. All nitrifiers are obligate aerobes and hence a restricted nitrification under waterlogged or aquatic environments can be observed. In addition, these microorganisms, especially *Nitrobacter*, are fairly sensitive to acidic pH. As a result nitrification is inhibited in climax ecosystems like forest soils with thick layer of leaf litter and zones of acidic pH (Roberge and Knowles, 1966). The process of nitrification itself may lead to lowering of pH of the medium due to release of H<sup>+</sup> as shown in the equation above.

### Why to inhibit or regulate nitrification?

As mentioned above, nitrification is one of the key N cycle processes under most arable situations on land. The following discussion will highlight the need to inhibit nitrification in order to maintain the economy of agro-ecosystems.

### Nitrification as a source of atmospheric N<sub>2</sub>O:

Concentrations of N<sub>2</sub>O in the atmosphere have increased markedly in recent years and are continuing to increase at 0.25% per annum (Denmead 1991). This increase is attributed mainly to biospheric processes. Flood irrigation leads to rapid nitrification/denitrification (Bacon *et al.*, 1986) resulting in considerable amounts of N<sub>2</sub>O being emitted to the atmosphere (Lindau *et al.*, 1990) which may amount to 35-45% of the applied N. On the global level, >65% of the atmospheric N<sub>2</sub>O comes from the soil, which is twice the amount produced by burning fossil fuels and four times the amount evolved from the oceans. Rates of denitrification in the soil may be as high as 30 kg N lost ha<sup>-1</sup> day<sup>-1</sup>. However, more realistic values could be 3 kg N ha<sup>-1</sup> day<sup>-1</sup>. Being a greenhouse gas (Bouwman, 1990), N<sub>2</sub>O contributes substantially to the destruction of stratospheric ozone (Crutzen, 1981). Efforts have therefore been devoted to identifying the processes that contribute to N<sub>2</sub>O emissions and possible mitigation strategies.

Among different processes, nitrification and denitrification are reported to be the main contributors to atmospheric N<sub>2</sub>O (Bouwman, 1990; Azam *et al.*, 2002). However, since the two processes occur simultaneously (aerobic and anaerobic microsites can develop within the same aggregate, supporting nitrification and denitrification, respectively), it is not easy to ascertain the

real contribution of either process to the observed  $N_2O$  fluxes (Arah, 1997). Nevertheless, nitrification is reported to make a substantial contribution to the  $N_2O$  emissions under aerobic conditions (Williams *et al.*, 1998). Higher  $N_2O$  emissions are often reported from fertilized (a routine practice in the present-day agriculture) than unfertilized soils, rates of emission being greatest following application of  $NH_4$  or  $NH_4$ -forming fertilizers (Breitenbeck *et al.*, 1980; Flessa *et al.*, 1996; Azam *et al.*, 2002). In several studies, using isotope methodology and nitrification inhibitors, this increase is attributed to losses of  $N_2O$  occurring during the process of nitrification (Bremner and Blackmer, 1978; Arah, 1997; Abbasi and Adams, 2000). Estimates of the amount of  $N_2O$  resulting from nitrification are variable but generally account for <1% of the fertilizer N applied (Breitenbeck *et al.*, 1980). In the case of anhydrous  $NH_3$ , however, the losses may increase to 6–7% (Smith and Chalk, 1980). In most studies, the onset of  $N_2O$  emission is observed very early during the incubation, while nitrification continues for extended periods of time (Simarmata *et al.*, 1993). Williams *et al.* (1998) reported active nitrification 7–12 days after application of  $NH_4NO_3$ , while a flush of  $N_2O$  emission from soil was observed around day 1, followed by a decline. These authors showed very low molar ratios of NO to  $N_2O$  and suggested that denitrification was the dominant process involved in  $N_2O$  emission. Azam *et al.* (2002) have reached to a similar conclusion from model experiments conducted under laboratory conditions.

Under the agro-climatic conditions of Pakistan, contribution of nitrification to  $N_2O$  emissions may be higher than generally reported. Urea, which is the major nitrogenous fertilizer in Pakistani agriculture, is rapidly hydrolyzed followed by a quick nitrification of the resultant  $NH_4$ , especially when applied under relatively warmer conditions. Thus not only nitrification contributes to  $N_2O$  emissions but the process of denitrification is fairly well supported by sustained availability of  $NO_3$ . Because of the low organic matter content of the soils that would limit the process of denitrification, a major source of  $N_2O$  evolved may still be nitrification. However, hardly any work has been reported from Pakistan on these aspects, whereas some good studies have been reported recently on denitrification from wheat-maize cropping systems (Mahmood *et al.*, 1999 and 2000). In most soils, formation and emissions of  $N_2O$  to the atmosphere are enhanced by an increase in available mineral N through increased rates of nitrification and denitrification. Therefore addition of N in organic or inorganic compounds eventually leads to enhanced  $N_2O$  emissions; N derived from BNF and returned to the soil in residues is also susceptible to such losses.

**Pollution of ground water:** Uncontrolled and excessive nitrification may lead to ground water contamination with  $NO_3$  and  $NO_2$  as well as increased concentration of the later in eatables, especially vegetables leading to human health hazards. Nitrate itself is not a threat while  $NO_2$  is definitely a potential health hazard and that too when found at places at a wrong time. In water bodies, however,  $NO_3$  and other forms of N may encourage the growth of algae and subsequently the bacteria leading to exhaustion of molecular oxygen thereby affecting animal life. Indeed whole ecological balance of water bodies may change due to the so-called “eutrophication”.

Consumption of water and vegetables containing excessive amounts of  $NO_3$  may lead to the production of  $NO_2$  in the stomach and the later becomes particularly dangerous for the babies. Methaemoglobonaemia (blue-baby syndrome) may occur in 1-year old babies taking diet with too much  $NO_3$ . Stomach cancer has also been associated with the concentration of  $NO_3$  in potable water. Again, it is  $NO_2$  that reacts with amines to form N-nitroso compounds which are reported to cause stomach cancer. Such an illness may result from consumption of vegetables containing high concentrations of  $NO_3$  originating from soil or irrigation water.

**Nitrification and ecosystem functioning:** Nitrification inhibition and consequent accumulation of  $NH_4$  would lead to:

- 1 Increased microbial activities including biological nitrogen fixation, a process that is known to be inhibited more by  $NO_3$ -N than  $NH_4$ -N.
- 2 Greater photosynthate partitioning to the rhizosphere thus enriching the soil with organic matter
- 3 Enhanced mineralization of native soil N through the so-called priming effect or added nitrogen interaction that is more with  $NH_4$ -N than  $NO_3$ -N
- 4 Increased efficiency of fertilizer N use by plants through decreased losses via denitrification and  $NO_3$  leaching

Microorganisms are known to prefer  $NH_4$  over  $NO_3$  as a source of N (Jansson, 1958). However, this preference is consequential rather than the reason. This is because of the fact that not all organisms possess nitrate reductase to enable them assimilate  $NO_3$ , while almost all of them will be able to assimilate  $NH_4$ . In addition, assimilation of  $NO_3$  is more energy intensive as compared to that of  $NH_4$ . Hence, for efficient assimilation of  $NO_3$ , ample supply of easily oxidizable C will be required. Studies involving the use of glucose as a C source indeed reveal similar

assimilation of both  $\text{NH}_4$  and  $\text{NO}_3$  by the soil microorganisms (Azam *et al.*, 1988, 1993). Nevertheless, presence of  $\text{NH}_4$  leads to an enhancement in microbial activities in terms of respiratory response. In experiments aimed at studying the mineralization of native soil N,  $\text{NH}_4$ -N is reported to have a significantly higher effect as compared to  $\text{NO}_3$ -N. This so-called "priming" effect or added nitrogen interaction (ANI) has frequently been reported using both isotopic as well as non-isotopic methods and is found to increase with the amount of applied N (Jenkinson *et al.*, 1985; Azam, 1990). Azam (1990) suggested that an indirect effect of chemical fertilizers as well as green manures is their positive influence on the mineralization and plant availability of N from the soil organic reserves.

One of the processes reported to be substantially affected by mineral N is the biological  $\text{N}_2$  fixation. This has been unequivocally proved in legumes at each level of the process starting from nodule initiation to actually fixation at the level of nitrogenase (Ledgard *et al.*, 1987; Blumenthal *et al.*, 1997). The effect is more intense with  $\text{NO}_3$  than  $\text{NH}_4$  leading to studies aimed at successful production of  $\text{NO}_3$  tolerant legume types (Herridge and Bergerson, 1988). This strategy is particularly important under conditions where legumes are grown together with non-legumes and the later are fertilized with chemical fertilizers. The nature has also endowed the rhizobia with the ability to avoid accumulation of inhibitory levels through dissimilatory  $\text{NO}_3$  reduction i.e., denitrification. Several studies suggest substantial denitrifying ability of these bacteria (Rosen *et al.*, 1996).

Plant species vary in their ability to utilize  $\text{NH}_4$  or  $\text{NO}_3$  as the sole source of N although a majority of them grow best with a mixture of the two. However, when used as an exclusive source of N,  $\text{NH}_4$  may cause growth inhibition in many species, particularly in those grown under arable conditions (Cramer and Lewis, 1993; Marschner, 1999). Under these conditions, nitrification is generally quite rapid and hence deleterious effects of  $\text{NH}_4$  are avoided. Under saline conditions also,  $\text{NH}_4$  is reported to increase the sensitivity of plants whereas  $\text{NO}_3$  has been reported to moderate the negative effects of salinity (Khan *et al.*, 1994). However, the plants are bound to face higher concentrations of  $\text{NH}_4$  under saline conditions because of the inhibitory effects of salts on the process of nitrification. It would appear therefore that nitrification inhibition would be a blessing for arable plants grown on normal agricultural soils whereas it may be an added problem for those grown on salt-affected lands. Several studies indeed show a positive effect of nitrification inhibitors on plant growth and N use efficiency (Crawford

and Chalk, 1993; Lodhi and Azam, 1998; Lodhi *et al.*, 1996 a,b). The mechanisms involved include i) decrease in losses through denitrification and  $\text{NO}_3$  leaching, ii) increased microbial activities in the presence of prolonged availability of  $\text{NH}_4$  and iii) increased rhizospheric activities etc. Nitrification inhibition may not only decrease in the loss of N through denitrification and  $\text{NO}_3$  leaching, but conserve the applied N through enhanced immobilization of the persistently available  $\text{NH}_4$ -N.

Form of N plays a significant role in affecting root growth, rhizodeposition and the concomitant changes in different rhizospheric microbial functions including root-induced N mineralization. In wheat and maize, root growth may be restricted in  $\text{NH}_4$  compared to  $\text{NO}_3$ -fed plants and may be attributed to an increased root respiration, greater allocation of photosynthates to nitrogenous than structural component and increased export of carbon (probably as amino acids) from root to shoot than that occurring under  $\text{NO}_3$  nutrition. In addition,  $\text{NH}_4$  nutrition leads to a higher rhizodeposition (Giordano *et al.*, 1994) thereby enlarging the below-ground sink for photosynthates, most probably at the expense of plant tops thereby reducing the biomass yield. However, increase in rhizodeposition due to increased/sustained availability of  $\text{NH}_4$  may also prove beneficial to plants in terms of increased microbial activities and especially the mineralization of native soil N. In laboratory experiments, a significant increase in the mineralization of soil N has been observed following addition of easily oxidizable C (Azam *et al.*, 1989).

It has also been suggested above, that mineralization of N from soil organic matter is more intense in the presence of  $\text{NH}_4$  than  $\text{NO}_3$ . Jenkinson *et al.* (1985) attributed this to "pool substitution" whereby the native N stands proxy for the applied N giving the impression of enhanced mineralization of the later. The fact remains, however, that applied N (especially  $\text{NH}_4$ ) leads to an increase in the availability of soil N. Inhibition of nitrification may therefore lead to a higher mineralization of native soil N thereby augmenting N supplies to plants. In addition, microorganisms responsible for the synthesis of aggregation-adhesion macromolecules may be encouraged by higher availability of carbonaceous materials in the rhizosphere. This will result in better soil structure as well as improved moisture holding capacity of the soil at the root surface. The later may help the plants withstand drought stress at least temporarily. Thus, in spite of the negative effects of  $\text{NH}_4$ , inhibition of nitrification may still exert beneficial effects on plant growth as mentioned above. The negative effects can be overcome to a significant extent by developing plant

types more efficient in using  $\text{NH}_4$ ; the so-called “ammoniphilic plants” (Prasad *et al.*, 1983). Plants like rice, sugarcane and kallar grass (*Leptochloa fusca*) etc. growing under high soil moisture conditions can be considered as ammoniphilic plants. Efforts are needed, however, to engineer arable crops (like wheat) for improved tolerance to  $\text{NH}_4$  while employing nitrification inhibitors.

Another aspect worth consideration is the susceptibility of  $\text{NO}_3$  to leaching beyond the effective root zone after being converted to  $\text{Ca}(\text{NO}_3)_2$  in the presence of ionic Ca. In Pakistan, the soils are mostly calcareous and hence the conditions are quite conducive to this mode of  $\text{NO}_3$  escape especially following organic amendment that helps in the release of Ca. The leaching is more pronounced in clayey soils at near neutral pH as negative charge on the clays repels  $\text{NO}_3$  thereby facilitating the process of leaching. Hence, not only the use efficiency of  $\text{NO}_3$  will remain low under these conditions but N economy of the system will be negatively affected.

#### How to inhibit nitrification?

The underlying concept in using nitrification inhibitors (NIs) is to decrease the availability of  $\text{NO}_3$  and hence its vulnerability to escape mechanisms as the later is directly proportional to the former. A great deal of work has been done and reported on the ways to retard/inhibit the rate of nitrification not only to reduce fertilizer N losses (Aulakh *et al.*, 1984) but also to prolong the persistence of fertilizer N in ammoniacal form (Crawford and Chalk, 1993). In recent years a large number of chemicals, including pesticides, have been used to inhibit nitrification (Bremner and Bundy, 1974; Feng and Barker, 1990; McCarty and Bremner, 1989; Lodhi *et al.*, 1996b). Some effective nitrification inhibitors along with their inhibition potential are listed in Table 1.

Among the different inhibitors, N-Serve and DCD are indeed the most frequently used (Hauck, 1980; Chalk *et al.*, 1990; Bronson *et al.*, 1992; Guiraud *et al.*, 1992; Abbes *et al.*, 1995; Lodhi *et al.*, 1996b; McTaggart *et al.*, 1997). The recommended rate of N-Serve application is 1.1 kg  $\text{ha}^{-1}$ , while DCD can be applied together with the fertilizer at 1-5% on N content basis. Dicyandiamide is an efficient inhibitor (Rodgers and Ainsworth, 1982). Being nonvolatile, nonhygroscopic and relatively soluble in water (23 g  $\text{L}^{-1}$  at 13 °C), it is one of the most convenient inhibitors to be used in formulations with a variety of fertilizers (Gioacchini *et al.*, 2002).

In addition to the synthetic inhibitors, allelochemicals released by plants are also reported to have an inhibitory effect. Rice (1984) postulated that because inhibition of nitrification results in conservation of both energy and

Table 1: Some commonly used nitrification inhibitors with variable inhibitory effect in different soils (adopted from McCarty and Bremner, 1989)

Compound	Soil 1	Soil 2	Soil 3
	% inhibition of nitrification		
2-Ethynylpyridine	79	80	100
Etridiazole	61	70	97
Nitrapyrin (N-Serve)	45	56	94
3-Methylpyrazole-1-carboxamide	43	53	93
4-Amino-1,2,4-triazole	41	52	92
Dicyandiamide (DCD)	8	20	41

nitrogen, vegetation in late succession or climax ecosystems contains plants that release allelochemicals that inhibit nitrification in soil. Some natural products like neem (*Azadirachta indica*) cake are reported to inhibit the activity of nitrifiers (Sahrawat and Parmar, 1974).

An ideal nitrification inhibitor should be mobile, persistent and economical in use. A major consideration during the selection of NIs is their high effectiveness at the lowest possible application rates with a minimum of side effects. Thus the availability of an inhibitor at effective concentrations is essential. This can be achieved by coating fertilizer granules with the inhibitor or by incorporating the latter into granules (Trenkel, 1997; Zerulla *et al.*, 2001). The aim of both the approaches is to ensure an intimate and uniform interaction of the substrate with the inhibitor. The application of an effective concentration of an NI to soil, together with N fertilizer, is a difficult task since it involves different crops, different forms and rate of N application etc. As a result, different concentrations of NI will reach the nitrifiers, particularly if N is applied as granules (Azam *et al.*, 2001). Among the different inhibitors, N-Serve and DCD are indeed the most frequently used (Hauck, 1980; Chalk *et al.*, 1990; Walters and Malzer, 1990; Bronson *et al.*, 1992; Guiraud *et al.*, 1992; Abbes *et al.*, 1995; Lodhi *et al.*, 1996b; McTaggart *et al.*, 1997). Dicyandiamide is an efficient inhibitor (Rodgers and Ainsworth, 1982). Being nonvolatile, nonhygroscopic and relatively soluble in water (23 g  $\text{L}^{-1}$  at 13 °C), it is one of the most convenient inhibitors to be used in formulations with a variety of fertilizers (Gioacchini *et al.*, 2002). The inhibitory effect is fairly dependent on the persistence of the chemical in soil. For example, DCD is highly susceptible to degradation at temperatures above 25 °C. Likewise, 50% of the N-Serve may be lost 5 weeks after application to the soil.

**Conclusion:** While application of chemical fertilizers to agricultural crops has resulted in tremendous increase in yield, problems arising due to the escape to the environment of different nitrogen species, especially  $\text{N}_2\text{O}$ ,  $\text{NO}_2$  and  $\text{NO}_3$ , have raised serious economic and environmental concerns. Of the different processes

responsible for these concerns, nitrification and denitrification are of prime importance. Hence, efforts have been made to regulate the process of nitrification (major source of different N species) as a means to enhancing the use efficiency of N, decreasing environmental/economic concerns and optimizing the functioning of agro-ecosystems. Use of nitrification inhibitors has been found helpful in mitigating the negative effects of fertilizer application. However, continued efforts need to be made for finding more efficient and environment friendly inhibitors to suit the ever-changing agro-climatic conditions. Fertilizer formulations containing efficient nitrification inhibitors could prove an appropriate way of application.

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