

<http://www.pjbs.org>

PJBS

ISSN 1028-8880

**Pakistan
Journal of Biological Sciences**

ANSI*net*

Asian Network for Scientific Information
308 Lasani Town, Sargodha Road, Faisalabad - Pakistan

Influence of Methylxanthines on the Labeling of Blood Elements with ^{99m}Tc

¹Jacques Natan Grinapel Frydman, ¹Márcia Betania Nunes de Oliveira ¹Ana Emília Oliveira dos Santos,
²Adenilson de Souza da Fonseca, ²Ricardo Santos and ¹Mario Bernardo-Filho
¹Department of Biophysics and Biometry, ²Department of Pharmacology and Psychobiology,
Institute of Biology Roberto Alcantara Gomes, State University of Rio de Janeiro, Av. 28 de Setembro 87,
fundos, 5º andar, 20551-030, Rio de Janeiro, Brazil

Abstract: This work evaluated the effect of some methylxanthines on the fixation of ^{99m}Tc on blood elements. Blood was incubated with different concentrations of the drugs before ^{99m}Tc. Plasma (P), blood cells (BC), insoluble (IF-P, IF-BC) and soluble (SF-P, SF-BC) fractions were separated and percentage of radioactivity (%ATI) bound were determined. The %ATI in IF-P and SF-P was altered (p<0.05) by caffeine at the highest concentrations used. Data showed that methylxanthines in concentrations found in humans not modify the fixation of ^{99m}Tc.

Key words: Aminophylline, caffeine, plasma proteins, red blood cells, theophylline, technetium-99m

INTRODUCTION

Some drugs can modify the labeling of red blood cells (RBC) with ^{99m}Tc and alter the results obtained in the daily routine procedure in nuclear medicine laboratories^[1-4]. High labeling yields and good *in vivo* stability of the *in vitro* labeling procedure gives superior images, while *in vivo* labeling is more convenient and thus, quite widely used. In addition to pool imaging and other uses in nuclear cardiology, applications of ^{99m}Tc -RBC have also included diagnosis of deep vein thrombosis, gastrointestinal bleeding, hepatic hemangionas and splenic reticuloendothelial system^[5,6] (Bernardo-Filho *et al.*, 1983; Bernardo-Filho *et al.*, 1994).

The labeling of RBC with ^{99m}Tc has been influenced by patient medications^[7] or labeling conditions^[6,8]. Thus, the presence of disease may be missed and/or underestimated^[7,9].

Aminophylline, caffeine and theophylline are methylxanthines that share in common several pharmacological actions of therapeutics interest. Caffeine and theophylline are naturally occurring methylxanthines and aminophylline is formed by complex between theophylline and ethylenediamine. Their actions include: (I) relax bronchial and cardiac muscles, (ii) stimulate central system and (iii) diuretic effect on the kidney^[10]. Aminophylline and theophylline are used as bronchodilators to relieve the spasm of bronchial smooth muscle in conditions such as bronchial asthma, chronic bronchitis, emphysema and cystic fibrosis. Caffeine is

present in human diet through various sources (coffee, chocolate, soft drinks) and it produces subjective and behavioral effects as such feelings well-being, motivation for work, energy, concentration, delays sleep and enhances vigilance performance on psychomotor tasks^[11-14].

The aim of this work was to investigate the *in vitro* effects of aminophylline, caffeine and theophylline on the labeling of blood elements with ^{99m}Tc.

MATERIALS AND METHODS

Animals: Adult male Wistar naive rats (3-4 month of age, body weight 250-350 g) were housed, five per cage, in an environment controlled room with inverted light/dark cycle conditions (12 h light/12 h dark; lights on at 6:00 a.m.), for an acclimatization period of at least 3 weeks. Animals had free access to water and food and ambient temperature was kept at 25±2°C. Experiments were conducted in accordance with the Department Committee of Animal Care.

Drugs: Aminophylline and theophylline were purchased from Sanofi Wintrop Farmacêutica Ltda (Brazil) and caffeine was purchased from Sigma (USA).

Study protocol: An *in vitro* technique employed to label RBC described elsewhere^[6] was used with minor modification.

Corresponding Author: Mario Bernardo-Filho, Department of Biophysics and Biometry, Institute of Biology Roberto Alcantara Gomes, University of State of Rio de Janeiro, Av. 28 de Setembro, 87, fundos, 5º andar, 20 551-030, Rio de Janeiro – RJ - Brazil, Phone (Fax) : (5521)2587-6432, E-mail: bernardo@uerj.br

Heparinized whole blood was withdrawn from Wistar rats. Samples of 0.5 ml were incubated with 100 µl of aminophylline, or theophylline (50.0 µg ml⁻¹) or different caffeine concentrations (5.0, 50.0, 625.0, 1250.0, 2500.0, 5000.0 and 10000 µg ml⁻¹) for 1 h at room temperature. A sample of heparinized whole blood was incubated with NaCl 0.9% (Reagen, Rio de Janeiro, Brazil) as a control. Then, 0.5ml of stannous chloride (1.2 µg ml⁻¹) (Sigma Chemical Co., St Louis, USA) was added and the incubation continued for another 1h. After this period, ^{99m}Tc (0.1 ml), as sodium pertechnetate, recently milked from a ⁹⁹Molybdenum/^{99m}Technetium generator (Instituto de Pesquisas Energéticas e Nucleares, Comissão Nacional de Energia Nuclear, São Paulo, Brazil), was added and the incubation continued for another 10 minutes. These samples were centrifuged and plasma (P) and blood cells (BC) were separated. Samples (20 µl) of P and BC were precipitated with 1 ml of trichloroacetic acid (TCA) 5% and soluble (SF) and insoluble fractions (IF) were separated. The radioactivity in P, BC, IF-P, SF-P, IF-BC and SF-BC were determined in a well counter (Clinigamma, gamma counter, LKB, Wallac, Finland). After that, the percentage of radioactivity (%ATI) was calculated, as previously described^[6].

Statistical analysis: Data was reported as means±SE of %ATI were compared between the treated and control groups by One way analysis of variance - ANOVA, followed by Bonferroni post test with a *p*<0.05 as significant level. InStat Graphpad software was used to perform statistical analysis (GraphPad InStat version 3.00 for Windows 95, GraphPad Software, San Diego California, USA).

RESULTS

The results in the Table 1 showed the distribution of the radioactivity on blood cells, insoluble fraction of blood cells (IF-BC) and insoluble fraction of the plasma (IF-P) from whole blood treated with aminophylline, caffeine or theophylline (50.0 µg ml⁻¹). The data presented the treatment with these drugs not significantly modify the uptake of ^{99m}Tc by blood cells, IF-BC or IF-P.

The Table 2 showed the fixation of the radioactivity on plasma and blood cells from whole blood treated with different concentrations of caffeine. The treatment with this drug not significantly modify the uptake of ^{99m}Tc by blood elements.

Table 3 showed the fixation of the radioactivity on plasma proteins from whole blood treated with the various concentrations of caffeine. The treatment with caffeine at

Table 1: Effect of aminophylline, caffeine and theophylline on the labeling of blood elements with ^{99m}Tc

%ATI			
Drug	Blood Cells	IF-BC	IF-P
Saline	93.0±0.4	88.0±3.1	70.0±1.7
Aminophylline	92.7±1.9	94.6±3.8	78.2±2.9
Caffeine	98.3±0.2	92.5±1.0	52.1±5.2
Theophylline	97.2±0.2	85.3±1.8	65.2±2.9

Table 2: Effect of Caffeine on the labeling of plasma and blood cells with ^{99m}Tc

%ATI		
Caffeine (µg ml ⁻¹)	Blood cells	Plasma
0.0	96.6±0.5	3.4±0.7
5.0	97.9±0.3	2.1±0.3
50.0	98.3±0.2	1.7±0.2
625.0	98.6±0.2	1.4±0.2
1250.0	96.8±0.9	3.2±0.9
2500.0	97.9±0.3	2.1±0.3
5000.0	97.5±0.8	2.5±0.8
10000.0	98.7±0.2	1.3±0.2

Table 3: Effect of Caffeine on the labeling of plasma proteins with ^{99m}Tc

%ATI		
Caffeine (µg ml ⁻¹)	IF-P	SF-P
0.0	68.3±3.0	31.7±3.0
5.0	53.5±5.0	46.5±5.0
50.0	52.1±5.1	47.9±5.1
625.0	46.4±4.4	53.6±4.4
1250.0	39.2±7.3	60.8±7.3
2500.0	43.9±4.9	56.1±4.9
5000.0	45.0±4.1	55.0±4.1
10000.0	33.1±0.8	66.9±0.8

Table 4: Effect of Caffeine on the labeling of blood cells proteins with ^{99m}Tc

%ATI		
Caffeine (µg ml ⁻¹)	IF-BC	SF-BC
0.0	92.8±1.6	7.2±1.6
5.0	89.1±2.2	10.9±2.2
50.0	92.5±1.0	7.5±1.0
625.0	89.2±2.0	10.8±2.0
1250.0	91.4±1.7	8.6±1.7
2500.0	95.0±0.5	5.0±0.5
5000.0	93.6±0.7	6.4±0.7
10000.0	94.6±0.6	5.4±0.6

highest concentrations (625 up to 10000 µg ml⁻¹) modify significantly (*p*<0.05) the fixation of ^{99m}Tc on insoluble (IF-P) and soluble (SF-P) fractions of plasma. The data suggest that caffeine at lowest concentrations (5 and 50 µg ml⁻¹) also may change the labeling of this plasma fractions. However, the difference with control is not significant (*p*>0.05).

The Table 4 showed the fixation of the radioactivity in blood cell proteins from whole blood treated with different concentrations of caffeine. The treatment with caffeine not modify significantly the fixation of the ^{99m}Tc on insoluble (IF-BC) and soluble (SF-BC) fractions of blood cells.

DISCUSSION

The interaction of drugs with radiopharmaceuticals is of interest. However, the data from these studies are relatively scarce and the effects of pharmacologically active agents on the diagnostic radiopharmaceuticals can be evaluated. Thus, a therapeutic drug can modify the nature or amount of the ^{99m}Tc -radiopharmaceutical bound to blood elements and this may result in unexpected behavior of the radiopharmaceutical.

The analysis of data obtained in this work showed that the methylxanthines (aminophylline, caffeine and theophylline) at concentration of $50 \mu\text{g ml}^{-1}$ not modify the fixation of ^{99m}Tc in the blood elements (Table 1). Aminophylline, caffeine and theophylline in the plasma concentrations more than 8, 15 and $20 \mu\text{g ml}^{-1}$, respectively, are associated with toxicity in human^[10]. Thus, the results obtained in this work showed that methylxanthines in the plasma concentrations highest than founded in humans not alter the uptake of the ^{99m}Tc .

Caffeine is obtained in the diet through coffee, tea, chocolate and cola-flavored drinks. Its plasma concentrations in humans can be highly variable due to consuming habits and amounts ingested.

Caffeine at lowest concentration used not modify the fixation of the ^{99m}Tc on the blood cells and plasma as well as in insoluble and soluble fractions of blood cells proteins (IF-BC and SF-BC) (Table 2 and 4). However, in high concentrations (highest than $625 \mu\text{g ml}^{-1}$), caffeine alters the fixation of ^{99m}Tc on plasma protein fractions (IF-P and SF-P).

The antioxidant ability of caffeine was demonstrated to be similar to that of the established biological antioxidant glutathione and significantly higher than ascorbic acid^[15]. Others data shown that this compound competes with oxygen for electrons and inhibits lipid peroxidation and protein oxidation as a function of concentration in mitochondrial membranes exposed to the gamma-radiation^[16]. Thus, the influence of caffeine on the fixation of ^{99m}Tc on plasma protein fractions observed at highest concentrations used may be related to a redox effect.

Another possibility that could explain the caffeine effect on fixation of ^{99m}Tc on plasma protein fractions is the competition between this drug and the radionuclide by the same binding sites. This hypothesis is based on other data that suggested that the labeling of blood elements with ^{99m}Tc can be altered by some drugs or extracts of plants^[17-19].

In conclusion, the data presented in this work showed that methylxanthines (aminophylline, caffeine and theophylline) at concentrations similar to the concentrations usually found in the plasma of human

beings not modify the labeling of blood elements with ^{99m}Tc in rats.

REFERENCES

1. Marini, R.P., R.J. Callahan, L.R. Jackson, S. Jyawook, M.I. Esteves, J.G. Fox, R.A. Wilkinson and H.W. Strauss, 1997. Distribution of technetium 99m-labeled red blood cells during isoflurane anesthesia in ferrets. *American J. Veterinarian Res.*, 58: 781-785.
2. de Oliveira, J.F., A.S. Avila, A.C. Braga, M.B. de Oliveira, E.M. Boasquevisque, R.L. Jales, V.N. Cardoso and M. Bernardo-Filho, 2002. Effect of extract of medicinal plants on the labeling of blood elements with Technetium-99m and on the morphology of red blood cells: I--a study with *Paullinia cupana*. *Fitoterapia*, 73: 305-312.
3. Capriles, P.V., A.P. Dias, T.E. Costa, M.B. Oliveira, M.V. Faria, E.G. Moura, B.A. Abreu and M. Bernardo-Filho, 2002. Effect of eggplant (*Solanum melongena*) extract on the *in vitro* labeling of blood elements with technetium-99m and on the biodistribution of sodium pertechnetate in rats. *Cellular and Molecular Biology*, 48: 771-776.
4. Fonseca de Oliveira, J., M.B. Nunes de Oliveira, A.S. Avila, A.C. Braga, M.T. Jansen de Almeida Catanho, R.L. Cavalcanti Jales, V.N. Cardoso and M. Bernardo-Filho, 2003. Assessment of the effect of *Fucus vesiculosus* extract on the labeling of blood constituents with technetium-99m and the histological modifications on the shape of the red blood cells. *Food and Chemical Toxicology*, 41: 15-20.
5. Bernardo-Filho, M., I.N.S. Moura and E.M. Boasquevisque, 1983. ^{99m}Tc -labeled red blood cells "*in vitro*". *Archives of Biological Technology*, 26: 455-461.
6. Bernardo-Filho, M., B. Gutflen and O.S. Maciel, 1994. Technetium-99m binding on plasma proteins and red blood cells: role of various precipitating agents. *Biomedical Letters*, 50: 17-24.
7. Sampson, C.B., 1996. Complications and difficulties in radiolabeling blood cells: a review. *Nuclear Medicine Communications*, 17: 648-658.
8. Garringer, M.A., 1996. The effect of cyclosporine concentration on the labeling efficiency of an *in vitro* technetium-99m red blood cell labeling procedure. *J. Nuclear Med. Technol.*, 24: 232-235.
9. Santos, J.S., E.F. Paula, T.G. Correa, L.C. Freitas, L.M.B. Fonseca, Gutflen and M. Bernardo-Filho, 1995. Effect of cyclophosphamide on the binding of $^{99m}\text{TcO}_4$ and $^{99m}\text{Tc-MDP}$ to blood cells and plasma proteins. *Brazilian J. Med. Biol. Res.*, 28: 131-135.

10. Hardman, J.G., L.E. Limbird and A.G. Gilman, 2001. *Goodman and Gilman's: The Pharmacological Basis of Therapeutics*. 10th ed. McGraw-Hill, New York.
11. Lieberman, H.R., R.J. Wurtman, G.G. Emde, C. Roberts and I.L.G. Covielle, 1987. The effects of low doses of caffeine in human performance and mood. *Psychopharmacology*, 92: 308-312.
12. Griffiths, R.R., S.M. Evans, S.J. Heishman, K.L. Preston, C.A. Sannerud, B. Wolf and P.P. Woodson, 1990. Low-dose caffeine discrimination in humans. *Journal of Pharmacology and Experimental Therapeutics*, 252: 970-978.
13. Landolt, H.P., D.J. Dijk, S.E. Gaus and A.A. Borbély, 1995. Caffeine reduces low-frequency delta activity in the human sleep EEG. *Neuropsychopharmacology*, 12: 229-238.
14. Garrett, B.E. and R.R. Griffiths, 1997. The role of dopamine in the behavioral effects of caffeine in animals and humans. *Pharmacology, Biochemistry and Behavior*, 57: 533-541.
15. Devasagayam, T.P., J.P. Kamat, H. Mohan and P.C. Kesavan, 1996. Caffeine as an antioxidant: inhibition of lipid peroxidation induced by reactive oxygen species. *Biochemica and Biophysica Acta*, 1282: 63-70.
16. Kamat, J.P., K.K. Bloor, T.P. Devasagayam, B. Jayashree and P.C. Kesavan, 2000. Differential modification by caffeine of oxygen-dependent and independent effects of gamma-irradiation on rat liver mitochondria. *Int. J. Radiation Biol.*, 76: 1281-1288.
17. Oliveira, J.F., A.C.S. Braga, A.S. Ávila, L.M.B. Fonseca, B. Gutflen and M. Bernardo-Filho, 1996. Effect of *Thuya occidentalis* on the labeling of red blood cells and plasma proteins with technetium-99m. *Yale J. Biol. Med.*, 69: 489-494.
18. Rusckowski, M., T. Qu, F. Chang and D.J. Hnatowich, 1997. Technetium-99m labeled epidermal growth factor-tumor imaging in mice. *J. Peptide Res.*, 50: 393-401.
19. Maranhão, R.C., B. Garicochea, E.L. Silva, P. Dorlhiac-Llacer, S.M. Cadena, I.J. Coelho, J.C. Meneghetti, F.J. Pileggi and D.A. Chamone, 1994. Plasma kinetics and biodistribution of a lipid emulsion resembling low-density lipoprotein in patients with acute leukemia. *Cancer Research*, 54: 4660-4666.