http://www.pjbs.org



ISSN 1028-8880

Pakistan Journal of Biological Sciences



Asian Network for Scientific Information 308 Lasani Town, Sargodha Road, Faisalabad - Pakistan

Studies on Acute and Lethal Toxicities of Iron and Nickel to the Fish

Muhammad Javed and Sajid Abdullah Fisheries Research Farms, Department of Zoology and Fisheries, University of Agriculture, Faisalabad, Pakistan

Abstract: Laboratory tests were conducted on three fish species viz. Catla catla, Labeo rohita and Cirrhina mrigala to determine their sensitivity to iron and nickel for 96-hr LC₅₀ and lethal toxicity. These tests were performed with three age groups of each fish species at water temperature of 30°C with pH 7 and total hardness of 100 mg L⁻¹. The 96-hr LC₅₀ concentrations of both iron and nickel varied significantly among the three fish species with age. 30-day fish were more sensitive to metals toxicity, followed by that of 60 and 90 day fish, respectively. Among the three fish species, Catla catla were more sensitive to nickel while Cirrhina mrigala showed highest sensitivity to iron. Lethal iron concentrations for all the three fish species ranged between 72.20 mg L⁻¹ (for 30 day *Labeo rohita*) and 164.65 mg L⁻¹ (for 90-day *Catla catla*). However, nickel concentrations fluctuated between 20.20 and 50.11 mg L⁻¹ for 30-day Catla catla and 60 day Cirrhina mrigala, respectively. The 96-hr LC₅₀ nickel concentrations in three fish species varied significantly within age groups. However, the difference between 60 and 90 day old fish was statistically non-significant. Among the three fish species, Catla catla appeared significantly more sensitive than that of Labeo rohita and Cirrhina mrigala to nickel contaminations. The 96-h LC50 concentrations of both metals for all the three fish species were positively and significantly dependent upon fish age. The regressions of water temperature on 96-hr LC₅₀ concentrations of both the metals were negatively significant. However, water pH, hardness, ammonia, sodium and potassium showed significantly direct relationships with 96-hr LC₅₀ concentrations of fish. The ammonia concentrations of test medium increased while dissolved oxygen decreased at high levels of toxicants. At higher 96-hr LC₅₀ values of iron, dissolved oxygen decreased significantly in the test medium.

Key words: Acute, lethal, toxicity, 96-hr LC_{50,} iron, nickel, fish

INTRODUCTION

Rapid industrialization in the province of Punjab has created a lot of problems including contamination of natural waters with metals due to dumping of untreated wastes in the aquatic habitats, causing deleterious effects on fish (Javed, 2004). The accumulation of metals in an aquatic environment has direct consequences to man and to the ecosystem also. Interest in the metals, which are required for metabolic activities in organism, lies in the narrow "window" between their essentiality and toxicity (Fatoki et al., 2002). Fish accumulate metals in its tissue from polluted environment. Metal distribution between different tissues of fish varies depending on the sources of uptake, diet and/or water borne exposure (Kraal et al., 1995). Amongst fish species, considerable differences in sensitivity to metals have been reported. Salmonids are generally sensitive to high cadmium levels (Suresh et al., 1993). Carp (Cyprinus carpio) and Catla catla have the ability to accumulate and concentrate iron to the levels

several orders of magnitude above those found in their environment (Abdullah et al., 2003). Determination of water quality criteria for aquatic life is similar to the solving of most biological problems, in which experimental data are obtained under controlled laboratory conditions in order to predict effects that might occur under natural conditions. Iron, zinc, lead, nickel and manganese are the most common pollutants of the rivers of Punjab province, entering them with industrial and municipal waste waters (Javed and Mahmood, 2001). Although, trace metals are essential for normal physiological process, abnormally high concentrations can be toxic to aquatic organisms (Javed, 2003). Heavy metals being non-biodegradable primarily necessitate knowledge on their uptake, distribution and persistence in tissues of organisms (Lewis et al., 2004). Egg hatchability and hatchling time are more sensitive indicators of toxicity than "standard" end points, like mortality and growth (Pyle et al., 2002). No work has been done on the determination of tolerance limits of major carps viz. Catla catla, Labeo rohita and

Cirrhina mrigala against iron and nickel toxicities. Therefore, present work was planned to investigate the comparative sensitivity of iron and nickel by means of toxicity tests (both LC₅₀ and lethal) of three fish species for different age groups.

MATERIALS AND METHODS

Iron and nickel toxicity tests on three fish species viz. Catla catla, Labeo rohita and Cirrhina mrigala were conducted in 50 L glass aquaria in clean water. Ten fish were placed into each aquarium for acclimation. In order to not stress the fish, the metal concentrations in aquarium were increased gradually, 50% concentration being reached in 3.50 h and full toxicant concentration in 7 h. Each test was conducted with three replications. Constant air was supplied to all the test mediums with air pump through capillary system. Chemically pure chloride compounds of iron and nickel were dissolved in distilled water and stock solutions prepared for required metal dilutions. The hardness, pH and electrical conductivity of water were adjusted. For each metal, the concentrations tested for three fish species, separately, were started from zero with an increment of 0.05 and 5 mg L⁻¹ (as total concentration) for low and high concentrations, respectively. In each trial, the observations of fish mortality and physico-chemical variables of the test medium viz. temperature, pH, total hardness, dissolved oxygen, ammonia, sodium, potassium and carbon-dioxide were made at 12 h intervals for 96-h. Dead fish were weighed individually (after being lightly blotted dry) and measured (total length) at the time of mortality observations. No mortality was observed among control fish. At the end of each test, water samples were taken from the aquarium and tested for metal concentrations (Anonymous, 1989). The analytical data obtained confirmed that the determined iron and nickel concentrations coincided with the estimated data quite satisfactorily. Iron and nickel toxicity tests were performed with three age groups of fish viz. 30, 60 and 90 day having following mean total lengths and weights:

Age group	Fish species	Total length (mm)	Weight (g)
(30 day)	Catla catla	43.37±4.44	1.2175±0.45
	Labeo rohita	21.87±2.17	0.1862 ± 0.08
	Cirrhina mrigala	18.32±1.25	0.4203±0.05
(60 day)			
	Catla catla	45.32±1.85	1.4110±0.04
	Labeo rohita	28.25±2.36	0.2650 ± 0.02
	Cirrhina mrigala	42.87±2.69	0.6737 ± 0.14
(90 day)			
	Catla catla	52.25±3.59	1.6900±0.29
	Labeo rohita	30.00±1.05	1.6500±0.39
	Cirrhina mrigala	43.28±2.33	1.0500 ± 0.20

The LC₅₀ and lethal levels, as total metal concentrations, were estimated for three fish age groups viz. 30, 60 and 90 day at temperature of 30°C with pH 7 and 100 mg L⁻¹ total hardness of water. The 96-hr LC₅₀ values and their 95% confidence intervals were estimated by using trimed spear man-Karber method (Hamilton *et al.*, 1977). Physico-chemical constituents of test mediums were also analyzed (Anonymous, 1984). The data obtained from this experiment were subjected to statistical analysis (Steel and Torrie, 1986) through microcomputer. Analysis of variance and Duncan's multiple range tests were performed to find-out statistically significant differences among parameters under study. Correlation and regression analyses were performed to find-out relationships among various parameters also.

RESULTS

Iron 96-hr LC₅₀ for fish: The present data shows that calculated 96-hr LC₅₀ values of three fish species varied within the range of 50.56±1.99-115.38±3.11 mg L⁻¹. The lowest 96-hr LC₅₀ was recorded for 30 day *Labeo rohita* while the highest one for 90-day *Catla catla*. The 95% confidence intervals of 96-hr LC₅₀ for 60 and 90 day *Catla catla* were almost similar (Table 1). Table 2 shows that 96-hr LC₅₀ concentrations varied significantly (p<0.05) among three fish species with age. Thirty day fish was significantly more sensitive to iron contamination than that of 60 and 90 day fish. The 96-hr LC₅₀ concentrations of iron increased with fish age. However, there was non-significant difference between 60 and 90 day old fish. *Labeo rohita* were significantly more sensitive to iron toxicity than that of *Cirrhina mrigala* and *Catla catla*.

Nickel 96-hr LC₅₀ for fish: The mean nickel 96-hr LC₅₀ concentrations varied significantly within the fish of three age groups as minimum of 6.00±0.25 mg L⁻¹ for 30 day *Catla catla* and a maximum mean value of 38.80±2.15 mg L⁻¹ for 90 day *Cirrhina mrigala*. Lethal concentrations of nickel for fish ranged between 20.20 and 50.11 mg L⁻¹ (Table 1). The 96-hr LC₅₀ nickel concentrations in three fish species varied significantly within age groups. However, the difference between 60 and 90 day old fish was statistically non-significant. Among the three fish species, *Catla catla* were significantly more sensitive than that of *Labeo rohita* and *Cirrhina mrigala* to nickel (Table 2).

Dependence of 96-hr LC₅₀ of metals on fish age: The 96-hr LC₅₀ concentrations of both metals for all the three fish species were positive and significantly dependent upon fish age i.e., with the increase in fish age the 96-hr

Table 1: Calculated 96-hr LC₅₀ as total iron and nickel concentrations for three fish species

Age group	Fish species	96-hr LC ₅₀ (mg L ⁻¹)	95 % confidence interval (mg L ⁻¹)	Lethal concentration (mg L ⁻¹)
Iron				
30 day	Catla catla	92.11±2.26	86.50-97.72	130.32
	Labeo rohita	50.56±1.99	45.62-55.50	72.20
	Cirrhina mrigala	77.95±3.66	68.86-87.04	83.16
60 day	Catla catla	110.02±2.91	102.79-17.25	162.28
	Labeo rohita	48.26±3.14	47.42-62.40	80.20
	Cirrhina mrigala	80.15±1.35	91.62-104.38	150.16
90 day	Catla catla	115.38±3.11	109.45-121.31	164.65
	Labeo rohita	56.20±2.34	50.62-62.01	75.66
	Cirrhina mrigala	100.05 ± 4.01	90.09-10.01	154.60
Nickel				
30 day	Catla catla	6.00±0.25	5.38-6.62	20.20
-	Labeo rohita	20.20±1.11	17.44-22.96	30.25
	Cirrhina mrigala	23.47±2.01	18.48-28.46	40.11
60 day	Catla catla	11.52±2.05	6.43-16.61	19.88
	Labeo rohita	24.94±3.61	15.97-33.91	40.02
	Cirrhina mrigala	34.80±2.25	29.21-40.39	50.11
90 day	Catla catla	20.15±1.64	16.08-24.22	30.36
-	Labeo rohita	27.00±2.00	22.03-31.97	40.28
	Cirrhina mrigala	38.80±2.15	32.96-43.64	50.05

Table 2: Metals 96-hr LC₅₀ responses of three age groups of fish

Fish age	Iron LC ₅₀	Nickel LC ₅₀	Fish species	Iron LC ₅₀	Nickel LC50
30 day	73.54±21.12b	16.56±9.29b	Catla catla	105.84±12.18a	12.56±7.13c
60 day	79.98±30.88a	23.75±11.68a	Labeo rohita	51.67±4.08c	24.05±3.49b
90 day	90.54±30.71a	28.65±9.43a	Cirrhina mrigala	86.05±12.17b	32.36±7.95a

Means with similar letters in a single column are statistically similar at p<0.05

Table 3: Dependence of 96-hr LC₅₀, concentration of three fish species on their age

Species	Regression equation	r	\mathbb{R}^2
Catla catla			
	Iron $LC_{50} = 82.57 + 0.388**$ (Fish	age) 0.955	0.9120
	Nickel LC ₅₀ = $-1.59+0.235**$ (Fish	age) 0.992	0.9841
Labeo rohita			
	Iron LC ₅₀ = $48.25 + 0.094**$ (Fish	age) 0.954	0.9101
	Nickel LC ₅₀ = $17.25+0.113**$ (Fish	age) 0.975	0.9506
Cirrhina mrigala			
	Iron LC ₅₀ = $69.90+0.368*$ (Fish a	age) 0.905	0.8190
	Nickel LC ₅₀ = $17.03+0.255**$ (Fish	age) 0.964	0.9293

^{* =} Significant at p<0.05; ** = Significant at p<0.01; r = correlation coefficient; R^2 = coefficient of determination

 $\underline{\text{Table 4: Physico-chemistry of the mediums during 96-hr LC}_{\text{50}} \text{ toxicity tests with three fish species}$

					Dissolved				
		Temperature		Total hardness	oxygen	Ammonia	Sodium	Potassium	CO_2
Age group	Fish species	(°C)	pН	(mg L^{-1})	(mg L^{-1})	$(mg L^{-1})$	(mg L^{-1})	(mg L^{-1})	(mg L^{-1})
IRON									
30 day	Catla catla	30.22±0.34	7.01±0.01	102.35±0.37	4.85±0.50	1.20±0.07	201.00±25.14	10.21±2.34	Nil
60-day	Catla catla	30.21±0.03	7.02 ± 0.03	100.22±0.35	4.68±0.45	1.34±0.08	307.83±31.48	13.47±4.19	Nil
90-day	Catla catla	30.30±0.01	7.08 ± 0.01	103.52±0.40	4.31±0.41	1.62±0.06	311.52±30.25	12.22±3.68	Nil
30-day	Labeo rohita	30.24±0.02	7.04±0.01	99.91±0.32	5.11±0.52	1.05±0.05	240.33±22.35	9.74±2.56	Nil
60-day	Labeo rohita	30.21±0.03	7.02±0.02	100.22±0.31	5.00±0.20	1.40±3.00	380.31±16.17	5.88±1.06	Nil
90-day	Labeo rohita	30.25±0.01	7.00±0.02	105.15±0.40	4.77±0.17	1.70±3.11	221.01±16.45	7.25±2.00	0.01
30-day	Cirrhina mrigala	30.30±0.05	7.08 ± 0.03	100.01±0.37	4.95±0.16	1.02±2.10	245.67±24.68	11.33±3.12	Nil
60-day	Cirrhina mrigala	30.21±0.03	7.02 ± 0.02	100.22±0.35	4.68±0.45	1.34±0.75	307.83±31.48	13.47±4.19	Nil
90-day	Cirrhina mrigala	29.94±0.01	7.00 ± 0.01	101.35±0.37	4.35±0.36	1.45±0.66	230.16±26.68	11.35±3.45	0.01
NICKEL									
30 day	Catla catla	30.12±0.29	6.95 ± 0.01	99.32±0.32	5.01±0.15	0.55±0.33	144.00±8.67	5.02±1.24	Nil
60 day	Catla catla	30.01±0.27	7.02 ± 0.01	100.66±0.33	6.37±0.49	1.63±0.69	160.72±16.09	5.69±0.41	Nil
90 day	Catla catla	29.67±0.28	7.05 ± 0.02	102.12±0.35	6.38±0.43	1.50±0.56	162.35±14.38	5.68±0.42	Nil
30 day	Labeo rohita	30.21±0.30	6.98 ± 0.01	100.02±0.35	5.28±0.40	1.25±0.51	152.00±20.31	5.82±1.00	0.12
60 day	Labeo rohita	30.01±0.31	7.02 ± 0.01	100.66±0.31	6.88±0.43	1.24±0.58	162.04±20.00	5.96±0.40	Nil
90 day	Labeo rohita	29.65±0.28	7.05 ± 0.03	101.00 ± 0.32	6.90±0.34	1.30±0.48	140.95±18.94	5.97±0.57	0.01
30 day	Cirrhina mrigala	30.57±0.34	7.01±0.01	99.80±0.30	4.85±0.15	0.31±0.25	77.77±11.22	2.55±0.42	0.01
60 day	Cirrhina mrigala	30.01±0.31	7.02±0.01	100.66±0.30	5.64±0.17	0.89±0.35	90.00±9.01	2.88±0.30	0.09
90 day	Cirrhina mrigala	30.04±0.29	7.04±0.03	101.28±0.34	4.38±0.15	1.20±0.33	92.34±8.99	3.11±0.25	0.03

Table 5: Relationships between metals toxicity (y) and physico-chemical variables (x) of the test medium

Metal	Regression equation $(y = a + bx)$		r	\mathbb{R}^2	
Iron					
	96-hr LC ₅₀ of fish (y)	=	30.20 - 0.0014* (Water temp.)	- 0.692	0.4788
		=	7.01 + 0.0001* (pH)	0.565	0.3192
		=	104.81 + 0.032* (Hardness)	0.537	0.2884
		=	5.48 – 0.009** (D. O.)	- 0.790	0.6241
		=	1.19 + 0.002* (Ammonia)	0.502	0.2520
		=	256.30 + 0.184 ^{NS} (Sodium)	0.179	0.0320
		=	3.28 + 0.087** (Potassium)	0.826	0.6823
Vickel					
	96-hr LC ₅₀ of fish (y)	=	30.15 - 0.0011* (Temp.)	- 0.614	0.3770
		=	6.97 + 0.002* (pH)	0.611	0.3733
		=	99.76 + 0.037* (Hardness)	0.654	0.4277
		=	6.04 - 0.013* (D.O.)	- 0.545	0.2970
		=	1.06 + 0.002* (Ammonia)	0.641	0.4109
		=	178.61 + 2.061* (Sodium)	0.608	0.3696
		=	6.46 + 0.075 NS (Potassium)	0.523	0.2735

^{* =} Significant at p < 0.05; ** = Significant at p < 0.01; r = 0

LC₅₀ values of metals increased. The high values of R² (coefficient of determination) for all the regression equations computed for the relationship of metal's toxicity with age predict highly reliable regression models (Table 3).

Physico-chemical variables of the test medium: The mean values for physico-chemical variables of the test mediums used during 96-hr toxicity tests are presented in Table 4. Table 5 shows the dependence of fish 96-hr LC₅₀ of iron and nickel on physico-chemical variables of the test medium. The regression equations reveal negative regression of water temperature on 96-hr LC₅₀ concentrations of both iron and nickel. However, water pH, hardness, ammonia and potassium showed direct relationships with 96-hr LC₅₀ concentrations of fish. The ammonia concentrations of test medium increased while dissolved oxygen decreased at high levels of toxicity. At higher 96-hr LC₅₀ concentrations of iron, dissolved oxygen decreased significantly. The 96-hr LC₅₀ of nickel for the fish showed positively significant correlation with sodium and potassium.

DISCUSSION

The present investigation reveals that 96-hr LC₅₀ concentrations of iron and nickel varied significantly among the three fish species with age. Thirty day fish exhibited more sensitivity than 60 and 90 day fish to metallic ion concentrations. Among the three fish species, Catla catla were the most sensitive to nickel toxicity, followed by Labeo rohita and Cirrhina mrigala. Cirrhina mrigala exhibited highest tolerance to nickel. Labeo rohita had the lowest 96-hr LC₅₀ to iron while the same was maximum for Catla catla. Amongst fish species, considerable differences in sensitivity to metals have been reported. Salmonids are generally sensitive to high

cadmium levels. Juvenile trout (Orcorhynchus mykiss) have higher 48-hr LC_{50} (Handy, 1992). In water hardness of 100 mg L⁻¹ Ca²⁺, carp fry and fingerling (Cyprinus carpio) had the 96-hr LC₅₀ of 4.3 and 17.10 mg L⁻¹ of cadmium, respectively (Suresh et al., 1993). Other fish characteristics, such as age, body size, feeding habits and sex can also be considered for variable LC_{50} of metals for different species of fish (Witeska and Jezierska, 2003). Therefore, it is important to consider the physico-chemical characteristics of test medium along with biotic factors to know the mechanisms affecting LC50 concentrations of fish in toxicity tests. During present investigation, total hardness showed significantly direct correlation with 96hr LC₅₀ concentration of metals for fish as these values escalated with increasing hardness of test medium. Zinc toxicity to rainbow trout showed positively significant dependence upon hardness and pH of water (Hansen and Welsh, 1996). At 8°C trout was less sensitive than that of at 12°C. Environmental conditions such as oxygen concentration, temperature, hardness, salinity and presence of other metals may modify metal's toxicity to fish. Hypoxic conditions, temperature increase and acidification usually render the fish more susceptible to intoxication (Witeska and Jezierska, 2003).

The ammonia excretion by the fish increased significantly at higher concentrations of metals. An inverse relationship between pH and the concentration of mobile magnesium, iron, manganese and cobalt has been reported (Boqomazov *et al.*, 1991). Increase in water temperature enhanced the uptake of metals by the aquatic organisms also (Jackson, 1988). Water-borne metals generally exhibit their greatest toxicity to aquatic organisms in soft waters of low pH (Penttinen *et al.*, 1995). This is because, the hardness cat-ions (Mg and Ca) compete with heavy metal cat-ions for binding sites within the organism. Acidic water, with high concentration of hydrogen ions, may also compete with metal cat-ions for

binding sites in the same manner as magnesium and calcium in hard waters. A negative correlation between pH and exchangeable zinc in water has been reported (Davies, 1992). Bioaccumulation of metals in fish is a function of metal bioavailability which can vary with pH, uptake and toxicokinitics (Spry and Wiener, 1991). Mercury uptake enhanced by increasing water temperature, reduced salinity and pH, at increased levels of zinc and cadmium in water (Eisler, 1985). Zinc toxicity to aquatic organisms was the highest at conditions of low pH, alkalinity, dissolved oxygen and elevated temperature (Weatherley et al., 1988). Heavy metal toxicity may also decrease the oxygen consumption by the fish (Sarkar, 1999) while water hardness had a significantly positive effect on heavy metals toxicity (Rathore and Khangranot, 2003).

ACKNOWLEDGMENT

This research work was supported by a grant (P-AU/Env.62) received from the Pakistan Science Foundation, Islamabad, Pakistan.

REFERENCES

- Abdullah, S., M. Javed and A. Iram, 2003. Bioaccumulation of iron in *Catla catla* from river Ravi. I. J. P. Sci., 2: 54-58.
- Anonymous, 1984. Official Methods of Analysis of the Association of Official Analytical Chemist, 14th Edn., Arlington, Virginia.
- Anonymous, 1989. Standard Methods for the Examination of Water and Wastewater, 17th Edn., APHA, Washington DC.
- Boqomazov, N.P., I.A. Shilnikov, S.M. Soldatov, S.N. Lebedev and Shilnikov, 1991. Effect of the pH of leached chernozem on mobility of iron and micronutrients. Sov. Soil. Sci., 23: 44-46.
- Davies, B.E., 1992. Trace metals in the environment, retrospect and prospect: In Bibliography of Trace Metals in Invertebrates (Adriano, D.C. Ed.). Lewis, Boca Raton, FL., pp. 383-399.
- Eisler, R., 1985. Selenium hazards to fish, wildlife and invertebrates: A symposium review US Fish and Wildlife Service, Biological Report, 85: 57.
- Fatoki, O.S., N. Lujiza and A.O. Ogunfowokan, 2002. Trace metal pollution in Umtata river. Water SA., 28: 183-189.
- Hamilton, M.A., R.C. Russo and R.V. Thurston, 1977.
 Trimmed spearman-Karber method for estimating median lethal concentrations in toxicity bioassays.
 Environ. Sci. Technol., 11: 714-719.

- Handy, R.D., 1992. The effects of cadmium and copper enriched diets on tissue contaminant analysis in rainbow trout (*Oncorhynchus mykiss*). Arch. Environ. Contam. Toxicol., 22: 82-87.
- Hansen, J.L and P.G. Welsh, 1996. Acute responses of bull trout (*Salvelinus confluentus*) to cadmium, copper and zinc. Boulder Colo., 15: 11-16.
- Jackson, T.A., 1988. Accumulation of mercury by plankton and benthic invertebrates in riverine lakes of northern Manitoba (Canada): Importance of regionally and seasonally varying environmental factors. Can. J. Fish. Aqua. Sci., 45: 1744-1757.
- Javed, M. and G. Mahmood, 2001. Metal toxicity of water in a stretch of river Ravi from Shahdera to Baloki headworks. Pak. J. Agric. Sci., 38: 37-42.
- Javed, M., 2003. Relationships among water, sediments and plankton for the uptake and accumulation of heavy metals in the river Ravi. I. J. P. Sci., 2: 326-331.
- Javed, M., 2004. Studies on metals toxicity and physicochemistry of water in the stretch of river Ravi from Baloki headworkd to Sidhnai barrage. I. J. Biol. Sci., 1: 106-110.
- Kraal, M.H., M.H.S. Kraak, C.J. DeGroot and C. Davids, 1995. Uptake and tissue distribution of dietary and aqueous cadmium by carp (*Cyprinus carpio*). Ecotoxicol. Environ. Saft., 31: 179-183.
- Lewis, M.A., R.L. Quarles, D.D. Dantin and J.C. Moore, 2004. Evaluation of a Florida Coastal Golf Complex as a local and water shed sources of bio-available contaminants. Mar. Pollut. Bull., 48: 254-262.
- Penttinen, S., J. Kukkonen and A. Oikari, 1995. The kinetics of cadmium in *Daphnia magna* as affected by humic substances and water hardness. Ecotoxicol. Environ. Saft., 30: 72-76.
- Pyle, G.B., S.M. Swanson and D.M. Lehmkuhl, 2002. Toxicity of uranium mine receiving waters to early life stage fathead minnows (*Pimaphales promelas*) in the laboratory. Environ. Pollut., 16: 243255.
- Rathore, R.S. and B.S. Khangarot, 2003. Effects of water hardness and metal concentration on a freshwater muller (*Tubifex tubifex*). Water Air Soil Pollut., 42: 341-356.
- Sarkar, S.K., 1999. Effects of two heavy metals (copper sulphate and cadmium sulphate) on the oxygen consumption of the fish *Cyprinus carpio*. U. P. J. Zool., 19: 13-16.
- Spry, D.J. and J.G. Wiener, 1991. Metal bioavailability and toxicity to fish in low alkalinity lakes: A critical review. Environ. Pollut., 71: 243-304.
- Steel, R.G.D. and J.H. Torrie, 1986. Principles and Procedures of Statistics: A Biometrical Approach, 2nd Edn., McGraw Hill Comp. Inc., New York.

- Suresh, A., B. Sivaramakrishna and K. Radhakrishna, 1993. Patterns of cadmium accumulation in the organs of fry and fingerlings of freshwater fish *Cyprinus carpio* following cadmium exposure. Chemosphere, 26: 945-953.
- Weatherley, A.H., P.S. Lake and P.L. Stahal, 1988. Zinc pollution and ecology of the environment: In Zinc in the Environment and Ecological Cycling. John Wiley, pp: 337-417.
- Witeska, M. and B. Jezierska, 2003. The effects of environmental factors on metal toxicity to fish. Fresenius Environ. Bull., 12: 824-829.