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Inhibition of Conidia Germination and Mycelial Growth of *Botrytis cinerea* by Some Alternative Chemicals

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Abstract: Fungal activities of food additives, potassium sorbate, methylparaben, sodium benzoate, propylparaben, sorbic acid and plant activators harpin-protein and potassium dioxide were comparatively examined with ipradion *in vitro* conditions on two isolates of *Botrytis cinerea* obtained from strawberry fruits. Food additives and plant activators showed inhibitory effect at different levels on the mycelial growth of fungus isolates. According to the Minimum Inhibition Concentration (MIC), sorbic acid ($300 \mu\text{g mL}^{-1}$ for each isolate) and comparison fungicide ipradion ($10 \mu\text{g mL}^{-1}$ for each isolate) displayed the highest inhibition effect on the radial mycelial growth of fungal isolates. Similar efficacy was obtained at the highest dose ($1000 \mu\text{g mL}^{-1}$) from other additives propylparaben (for both isolates), potassium sorbet and methylparaben (for isolate 1) and from plant activator potassium oxide. It was detected that all alternative substances has high inhibition activity according to ED_{50} values. Methylparaben, harpin-protein and potassium oxide revealed efficacies similar to ipradion. While harpin-protein did not have any effect on *B. Cinerea* germination, other alternative substances excluding potassium sorbate inhibited germination completely at differing doses. Potassium oxide and food additives except potassium sorbate inhibited the spore germination of isolates by 50% at doses between $25\text{-}345 \mu\text{g mL}^{-1}$. MIC value of ipradion on spore germination of isolate-1 and isolate-2 was determined as 10 and $30 \mu\text{g mL}^{-1}$, while ED_{50} values were determined as 2.9 and $12.5 \mu\text{g mL}^{-1}$, respectively. Data obtained from the research verified that food additives may be used in traditional and organic agriculture individually, or in appropriate combination with each other or with fungicides. Moreover, the study put forth that harpin-protein and potassium oxide not only activate plants against pathogens, but they may also have toxic effect on different growth periods of fungus. Although ipradion works only with two isolates, its MIC and ED_{50} values were considered noteworthy in terms of the resistance risk of *B. cinerea*.

Key words: *Botrytis cinerea*, food additive, plant activators, effectiveness, alternative chemical

INTRODUCTION

Botrytis cinerea Pers: Fr. is an important pathogen that has a broad host range including vegetables, indoor plants, bulbed plants and fruits. If grey mould disease (*B. cinerea*) cannot be prevented adequately, it may cause important losses in efficiency and quality (Coley-Smith *et al.*, 1980; Erkan *et al.*, 1997; Boyd-Wilson *et al.*, 1998; Baraldi, 2002). The disease may be controlled in classical fungicides; however, most of the traditional fungicides embody the risk of leaving remnants on products and have negative effects on human beings, animals and environment (Erkan *et al.*, 1997). Furthermore, *B. cinerea* may easily develop races with high resistance against many synthetic fungicides (AbouJawdah and Itani, 1994; Erkan *et al.*, 1997; Knight *et al.*, 1997; Palmer *et al.*, 1997; Vallejo *et al.*, 2003).

On these grounds, recent studies have focused on alternative chemicals and biologic agents in the control of pathogens (Antonov *et al.*, 1997; Archbold *et al.*, 1997; Boyd-Wilson *et al.*, 1998; Reddy *et al.*, 1998; Ntirampemba, 1998). Plant extracts, sodium and potassium salts and some antimicrobial food additives have gained importance in the control of pathogens.

In this study, the effects of food additives, potassium sorbate, sodium benzoate, methylparaben, propylparaben, sorbic acid and potassium dioxide and harpin-protein *in vitro* on the mycelium growth and conidia germination of *B. cinerea* were determined *in vitro*.

MATERIALS AND METHODS

Origin of *B. cinerea* isolates: *B. cinerea* isolates (I1, I2) were isolated from infected fresh strawberry fruits (C1;

Table 1: Some properties of the test chemicals

Commercial name	Active ingredient	Formulation and ratio of a.i.	Firm
Mesenger	Harpin-protein	10	A.M.C. Chemical, S.L.
Bioclean	Potassium oxide	K ₂ O, 94.2	Green Solutions Co. Ltd.
Mass rodin	Ipradion	WP, 50%	Mass Tar. İlaçları Ltd.
Potassium sorbate	Potassium sorbate	C ₆ H ₇ O ₂ K, 150.22	Selen Kim. Tiç. Ltd.
Methylparaben	Methyl-p-hydroxybenzoat	C ₈ H ₈ O ₃ , 152.15	Tianyu Fine Chem, Ltd.
Sodium benzoate	Sodium benzoate	C ₇ H ₅ NaO ₂ , 144.11	Selen Kim. Tiç. Ltd.
Propylparaben	Propyl-p-hydroxybenzoate	C ₁₀ H ₁₂ O ₃ , %100	Tianyu Fine Chem, Ltd.
Sorbic acid	Sorbic acid	C ₆ H ₈ O ₂ , 112.13	Acetic Acid Chem. Ltd.

Camorosa ve SC2; Sivic Charli) obtained from a greenhouse in Aydın (Turkey). Isolates were developed in an incubator at 21°C in PDA environment and were transferred into tubes with PDA in order to obtain stock cultures.

Natural chemicals: Potassium-sorbate, methylparaben (methyl-p-hydroxybenzoate), harpin-protein, potassium dioxide, sodium benzoate, propylparaben (propyl-p-hydroxybenzoate), sorbic acid and ipradion as comparison fungicide were tested *in vitro* so as to determine the efficiency of alternative chemicals against *B. cinerea*. Test chemicals were used at doses of 0 (control), 0.3, 1, 3, 10, 30, 100, 300 and 1000 µg mL⁻¹. In the experiments, streptomycin sulfate (0.3 µg mL⁻¹) was added to the culture environment (Delen *et al.*, 1984). Some properties of test chemicals and comparison fungicides are given in Table 1.

Efficiency of test chemicals on mycelium growth: Stock isolates were developed on PDA environment and agar discs (Ø4 mm) cut from fungal culture using a sterile cork borer were planted on the surface of MM environment with and without test chemicals by placing the side with fungal culture against the bottom. After inoculation, petri dishes wrapped with parafilm were incubated at 25°C in the dark. Colony diameters were measured at the widest point for 3 days at 24 h intervals. Inhibitory effects of chemicals were detected by comparing the diameters of the control group with the diameters of colonies developed in nutrition environment treated with chemicals. ED₅₀ values were found using semi-logarithmic graphics. The trials were established three repetition and coincidence blocks trail design.

Inhibition of conidia germination: Efficiency of test chemicals on the conidia germination of *B. cinerea* was tested in MM environment and it was compared with the efficiency of ipradion. For this purpose, isolates were transferred from stock cultures to MM environment and they were incubated in the incubators for 10 days at 21°C in the dark for 12 h and in the light for 12 h alternately.

Later, MM environments with and without test chemicals were inoculated in a glass tower (0.35×0.70 m² high) with conidia obtained from fungus cultures (30 conidia mm⁻²). Conidia obtained from cultures were given to the environments by means of an air current provided by an air pump. Inoculated petry dishes were incubated at an optimum temperature of 20°C in the dark for the germination of *B. cinerea* conidia. In order to stop conidial growth at the end of 24 h after inoculation, blotting papers pasted inside petry stoppers were treated with formaldehyde (7%). Conidia germination percentage was determined by observing 200 spores under microscope for each recurrence. Conidia germination was verified if the length of germ tube was equal to or more than the spore (Vicedo *et al.*, 2006). Treatment was established four repetition and coincidence blocks trial design. Inhibitory effects of chemicals were determined by means of Abbot Formula and ED₅₀ values were found using semi-logarithmic graphics.

RESULTS

Efficacies of chemicals on colonial growth of *B. cinerea* (isolate-1) are given in Fig. 1 and 2. Colonial growth of both isolates was generally inhibited to a significant extent even in the lowest doses of test chemicals (Fig. 1 and 2).

It was detected that ipradion at low doses (0.3-3 µg mL⁻¹) with a minimum 3% and maximum 100% inhibition rate is the most effective chemical in inhibiting colonial growth and it is followed by methylparaben, bioclean, harpin-protein and others respectively (Fig. 1). Sorbic acid at 1000 µg mL⁻¹ dose reached the highest efficacy (100%) on colonial growth of isolate-1. On the other hand, harpin-protein at 1000 µg mL⁻¹ dose revealed the lowest efficacy (74.7%) among test chemicals.

Efficacies of test chemicals on the colonial growth of isolate-2 were similar to the efficacies of that of isolate-1. Ipradion, methylparaben, harpin-protein and potassium oxide at low doses displayed important efficacies on colonial growth of isolate-2; however, other chemicals were effective on colonial growth only at doses over

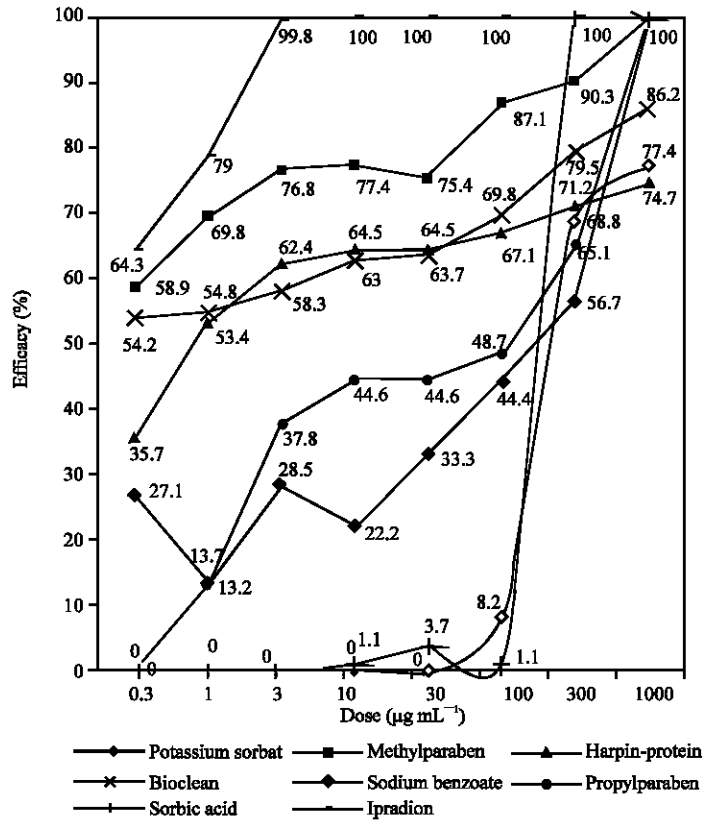


Fig. 1: Effects of the chemicals on colonial growth of *Botrytis cinerea* (Isolate-1)

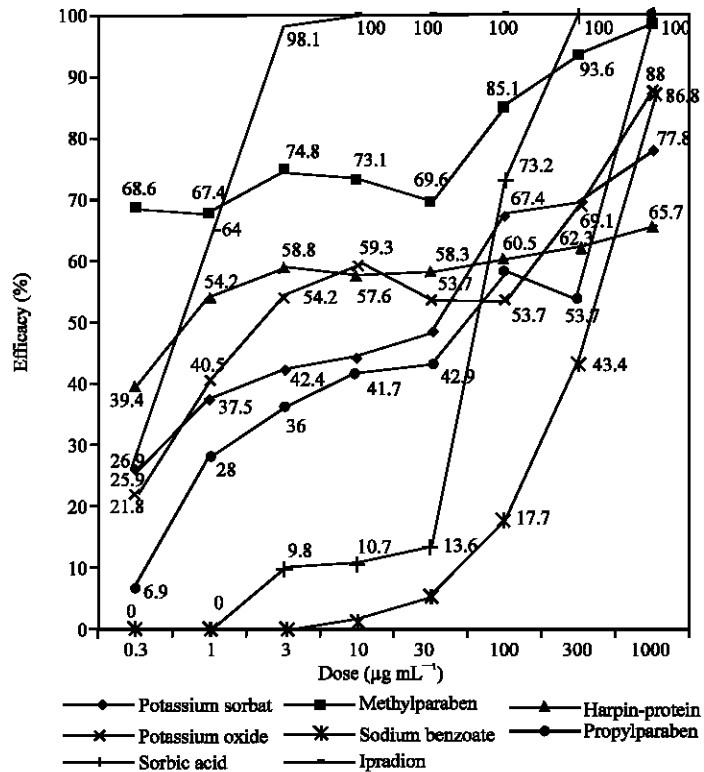


Fig. 2: Effects of the chemicals on colonial growth of *B. cinerea* (Isolate-2)

Table 2: MIC and ED₅₀ values of test chemicals on mycelial growth of *Botrytis cinerea*

Test chemicals	Isolate-1		Isolate-2	
	MIC (µg mL ⁻¹)	ED ₅₀ (µg mL ⁻¹)	MIC (µg mL ⁻¹)	ED ₅₀ (µg mL ⁻¹)
Potassium sorbate	1.000	120	>1.000	65
Methylparaben	1.000	<0.3	>1.000	<0.3
Harpin-protein	>1.000	0.8	>1.000	0.8
Sodium benzoate	>1.000	220	>1.000	352
Sorbic acid	300	157	300	63
Potassium oxide	>1.000	<0.3	>1.000	2.2
Propylparaben	1.000	110	1.000	52
Ipradion	10	<0.3	10	0.85

Table 3: Efficacies of chemicals on Conidia germination of *B. cinerea* (Isolate-1) (%)

Dose (µg mL ⁻¹)	Potassium sorbate	Methylparaben	Harpin protein	Potassium oxide	Sodium benzoate	Propylparaben	Sorbic acid	Ipradion
0.3	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
3	0.0	0.0	0.0	0.0	0.0	0.0	0.0	56.1
10	0.0	0.0	0.0	0.0	0.0	0.0	0.0	100.0
30	1.1	0.0	0.0	0.0	0.0	0.0	64.4	100.0
100	4.0	0.0	0.0	0.0	65.7	4.0	75.7	100.0
300	8.5	62.3	0.0	100.0	100.0	31.5	100.0	100.0
1000	6.2	100.0	0.0	100.0	100.0	100.0	100.0	100.0

Table 4: Efficacies of chemicals on Conidia germination of *B. cinerea* (Isolate-2) (%)

Dose (µg mL ⁻¹)	Potassium sorbate	Methylparaben	Harpin protein	Potassium oxide	Sodium benzoate	Propylparaben	Sorbic acid	Ipradion
0.3	1.0	0.0	0.0	0.0	0.0	1.04	3.08	0.0
1	2.0	0.0	0.0	0.0	0.0	3.08	4.10	0.0
3	1.5	5.0	0.0	1.5	0.0	3.08	5.10	0.0
10	11.5	15.5	0.0	8.5	0.0	4.10	9.20	20.4
30	15.5	51.5	0.0	14.5	7.3	8.10	11.20	100.0
100	16.5	76.5	0.0	15.5	9.5	100.00	100.00	100.0
300	18.5	94.0	0.0	39.0	100.0	100.00	100.00	100.0
1000	24.0	100.0	0.0	100.0	100.0	100.00	100.00	100.0

30 µg mL⁻¹ (Fig. 2). Minimum Inhibition Concentrations (MIC) and ED₅₀ values of test chemicals on radial mycelial growth of *B. cinerea* are introduced in Table 2.

Ipradion (comparison fungicide) at a dose of 10 µg mL⁻¹ completely inhibited the radial mycelial growth of both isolates of *B. cinerea*. It was followed by sorbic acid at a dose of 300 µg mL⁻¹. MIC values of other alternative chemicals were detected at 1000 µg mL⁻¹ and higher doses (Table 2). On the other hand, ED₅₀ values of methylparaben, potassium oxide and harpin-protein were close to the values of Ipradion. ED₅₀ values of other alternative chemicals were slightly higher than that of Ipradion (Table 2). Ipradion at a dose of 3 µg mL⁻¹ reduced conidia germination of *B. cinerea* (Isolate-1) to a significant extent, whereas alternative chemicals at low doses failed to inhibit conidia germination of *B. cinerea* (Table 3). Sorbic acid at a concentration of 30 µg mL⁻¹ inhibited conidia germination considerably and it was followed by sodium benzoate at 100 µg mL⁻¹ dose, potassium oxide at 300 µg mL⁻¹ dose and methylparaben respectively. On the other hand, while harpin-protein did not show any efficacy on conidia germination of *B. cinerea* (isolate-1), potassium sorbate could not reveal

sufficient efficacy on conidia germination of *B. cinerea*. Potassium oxide, sorbic acid and sodium benzoate at 300 µg mL⁻¹ completely inhibited conidia germination (Table 3).

While comparison fungicide Ipradion inhibited conidia germination of Isolate-2 to 20.4% at a dose of 10 µg mL⁻¹, it had a complete inhibitory effect at 30 µg mL⁻¹ and higher doses (Table 4). It was determined that sorbic acid and propylparaben at 100 µg mL⁻¹ dose, among alternative substances, have the lowest minimum inhibition concentration on conidia germination of Isolate-2 and these are followed by sodium benzoate (300 µg mL⁻¹), methylparaben (1000 µg mL⁻¹) and potassium oxide (1000 µg mL⁻¹) respectively. While harpin-protein did not have any efficacy on conidia germination of isolate-2, MIC value of potassium sorbate was detected to be above 1000 µg mL⁻¹ (Table 4).

ED₅₀ values of test chemicals on conidial germination of *B. cinerea* isolates are introduced in Table 5.

Among test chemicals, ipradion at doses of 2.9 and 12.5 µg mL⁻¹ had the lowest ED₅₀ values on conidial germination of both isolates and it was followed by sorbic acid (Table 5). ED₅₀ values of methylparaben,

Table 5: ED₅₀'s of test chemicals on Conidial germination of *Botrytis cinerea*

Chemicals	Isolate 1	Isolate 2
	ED ₅₀ (µg mL ⁻¹)	ED ₅₀ (µg mL ⁻¹)
Potassium sorbate	>1000	>1000
Methylparaben	272	29
Harpin-protein	>1000	>1000
Sodium benzoate	90	135
Sorbic acid	25	40.5
Potassium oxide	172	340
Propylparaben	345	42
Ipradion	2.9	12.5

propylparaben, sodium benzoate and potassium oxide revealed differences depending on the isolates. ED₅₀ values of potassium sorbate and harpin-protein were determined to be 1000 µg mL⁻¹ higher for both isolates.

DISCUSSION

In this study; potassium dioxide and harpin protein besides potassium sorbate, sodium benzoate, methylparaben, propylparaben and sorbic acid, which are food additives that may be alternatives to classical fungicides, were analysed *in vitro* conditions in comparison with a classic fungicide ipradion regarding their efficacies on mycelium growth and conidia germination of *Botrytis cinerea*.

Among alternative substances, food additives potassium sorbate, sodium benzoate, methylparaben, propylparaben and sorbic acid and harpin-protein and potassium oxide showed strong inhibition effect on mycelial growth and conidia germination of *B. cinerea* at increasing doses. Food additives are being used widely for a long time in cosmetics and in the field of medicine, as well as in food preservation due to their anti-microbial effects and they have high efficacy against yeast and mould (Wade and Weller, 1994). Effect mechanisms of sorbic acids, benzoic acids and parabens against yeast and fungi rely on two factors: decreasing of pH in the intracellular part because of becoming ionized of fungi's non-decomposing acid molecules or damaging of substrate transporting due to changing of cell-membrane's permeability (Liewen, 1991; Khan *et al.*, 2001; Vicedo *et al.*, 2006). Enzyme activation in fungi depends on pH value and performance of extracellular enzyme catalysis decreases at pH levels that are not within optimum levels (Dix and Webster, 1995). Therefore, fungus cannot provide the sufficient energy for hif development (Skirdal and Eklund, 1993).

It was verified in the study that especially sorbic acid inhibits mycelial growth of both isolates (isolate-1, isolate-2) of *B. cinerea* at low doses. Similarly, it was noted that different forms of sorbic

acid play a considerable role in the inhibition of fungal growth of *Penicillium chrysogenum*, *Cladosporium cladosporioides* and *Ulocladium atrum* (Palmer *et al.*, 1997). Sorbic acid at doses of 100 and 300 µg mL⁻¹ completely inhibited conidia of both isolates of *B. cinerea* and proved to be successful in the inhibition of mycelial growth. It was also detected that sorbic acid has low ED₅₀ values, such as 25 µg mL⁻¹ (for isolate-1) and 40.5 µg mL⁻¹ (for isolate-2), besides low MIC values on chemical germination. Although sorbic acid effectively inhibits radial mycelia growth and spore germination of *B. cinerea*, its hydrophobic structure makes it difficult to use it in practice as antifungal agent. Dudman (1963) notes that using water-soluble salts of sorbic acid, such as sorbate and sodium sorbate, may provide a solution in practice. Although potassium sorbate in this study could not show sufficient efficacy on conidia germination (ED₅₀ and MIC values for both isolates were higher than 1000 µg µg mL⁻¹), it inhibited mycelial growth of fungus to a great extent. When the significance of mycelial growth in disease development is taken into consideration, potassium sorbate may be regarded as successful.

Other additives, parabens (methylparaben, propylparaben) and sodium benzoate, showed high inhibitory effect on radial mycelia growth and conidia germination of both isolates (isolate-1 and isolate-2) of the pathogen. Sodium benzoates and parabens are used in additives to inhibit fungal mould (Khan *et al.*, 2001).

Sodium benzoate could not totally inhibit mycelia of isolates *in vitro*, but ED₅₀ values were found to be at a low level (220 µg mL⁻¹ for isolate-1; 352 µg mL⁻¹ for isolate-2). Sodium benzoate displayed a higher efficacy on conidia germination of *B. cinerea* isolates. Clausen and Yang (2003) noted that sodium benzoate and potassium sorbate at doses of 5% inhibited the spores of *Penicillium chrysogenum*, *Aspergillus niger* and *Trichoderma viride* and also stated that sodium nitrate at dose combinations of 0.3%, in addition to especially these two chemicals, totally inhibited the spores of all three fungi in malt agar environment.

Other additives, methylparaben and propylparaben, had important inhibitory effects on radial mycelia growth and conidia germination of *B. cinerea* isolates. It was observed that efficacy of methylparaben on mycelia growth of fungus was close to that of comparison fungicide ipradion. While LD₅₀ value of the chemical in isolate-1 (ED₅₀<0.3 µg mL⁻¹) was the same with the value of ipradion, it was noteworthy that this value was at lower doses (ED₅₀<0.3 µg mL⁻¹) than ipradion (ED₅₀ = 0.85 µg mL⁻¹) in isolate-2. However, ED₅₀ values on spore germination were observed to be slightly

higher than ipradion in both isolates. MIC values of the chemical were determined to be 1000 $\mu\text{g mL}^{-1}$ on mycelia growth and spore germination of both isolates. In a study carried out with mould fungi, it was observed that methylparaben displayed different MIC values depending on fungus type; for example, 600 $\mu\text{g mL}^{-1}$ against *Aspergillus oryzae*, 1000 $\mu\text{g mL}^{-1}$ against *Aspergillus niger* and 250 $\mu\text{g mL}^{-1}$ against *Trichoderma lignorum* (Haag and Loncrini, 1984).

It was observed that propylparaben at low doses had a higher inhibitory effect on mycelia growth of *B. cinerea* isolates than the effect of sorbic acid and sodium benzoate. Yet, MIC value determined as 1000 $\mu\text{g mL}^{-1}$ against both isolates was higher than sorbic acid. Propylparaben completely inhibited conidia germination of isolate-1 and isolate-2 at doses of 1000 and 100 $\mu\text{g mL}^{-1}$, respectively. ED_{50} values on germinations were similar to the values of methylparaben.

Apart from the food additives included in the study, potassium oxide and harpin-protein displayed a medium level of inhibition effect on mycelia growth of fungus. While ED_{50} values of both substances on mycelia growth were detected to be quite low, their MIC values were above the dose of 1000 $\mu\text{g mL}^{-1}$. Harpin-protein did not have any efficacy on conidia germination of *B. cinerea* isolates, whereas potassium oxide showed efficacy only at high doses. ED_{50} value of harpin-protein was above 1000 $\mu\text{g mL}^{-1}$ for both isolates as observed in mycelia growth, but the values of potassium oxide were detected to be lower. Although it is known that harpin-protein is not directly effective on microorganisms, but stimulates resistance against pathogens in plants (EPA, 2006); it was considered noteworthy that it inhibited especially mycelia growth of fungus in this study. This result indicates that harpin-protein can have direct toxic effect on mycelia of fungi.

Similar to harpin-protein, potassium oxide also strengthens the defence mechanism of plants rather than providing a direct effect on the pathogens. The efficacy of potassium oxide on fungal growth may be considered as a direct toxic effect.

In the study, comparison fungicide ipradion revealed high inhibitory effect both on mycelia growth and conidia germination of *B. cinerea*. Erkan *et al.* (1997) determined in their study, in which they examined the sensitivity of *B. cinerea* isolates obtained from vineyards towards dicarboximide fungicides, that isolates have high sensitivity towards ipradion and their ED_{50} values on mycelia growth of fungus and MIC value on spore germination are both lower than 0.3-1.7 $\mu\text{g mL}^{-1}$. However, it is significant that ED_{50} values obtained on spore

germination (2.9 $\mu\text{g mL}^{-1}$ for isolate-1; 12.5 $\mu\text{g mL}^{-1}$ for isolate-2) indicate that isolates are sensitive to ipradion at low and medium levels (Erkan *et al.*, 1997).

Data obtained from the study have proved that food additives *in vitro* inhibit mycelia growth and spore germination of *B. cinerea* to a significant extent especially at high doses. This shows that food additives, alone or in combinations, may replace classical fungicides in organic and traditional agriculture in practice. Furthermore, synthetic fungicides may also be used in combination with food additives in order to increase their efficacies. Yet, in the first place, it requires researches to be carried out on plants. Subsequently, it has been determined in this study that harpin-protein and potassium oxide not only activate plants against disease factors in practice, but also have toxic effects on pathogens.

REFERENCES

- Abou-Jawdah, Y. and H. Itani, 1994. Resistance of *Botrytis cinerea* isolates to fungicides used in Lebanon. In: Proceedings of 9th Congress of the Mediterranean Phytopathology Union, 18-24 September 1994, Kuşadası-Aydın, Türkiye, pp: 373.
- Antonov, A., A. Stewart and M. Walter, 1997. Inhibition of conidium germination and mycelial growth of *Botrytis cinerea* by natural products. In: Proceedings of 50th N.Z. Plant Protection Conference, 18-21 August 1997, Lincoln, pp: 159-164.
- Archbold, D.D., T.R. Hamilton-Kemp, M.M. Barth and B.E. Langlois, 1997. Identifying natural volatile compounds that control gray mold (*Botrytis cinerea*) during postharvest storage of strawberry, blackberry and grape. J. Agric. Food Chem., 45: 4032-4037.
- Baraldi, E., P. Bertolini, E. Chierici, B. Truffelli and D. Luiselli, 2002. Genetic diversity between *Botrytis cinerea* isolates from unstored and cold stored kiwi fruit. J. Phytopathol., 150: 629-635.
- Body-Wilson, K.S.H., J.H. Perry and M. Walter, 1998. Persistence and Survival of Saprophytic Fungi Antagonistic to *Botrytis cinerea* on Kiwifruit Leaves. In: Proceedings of 51st N.Z. Plant Protection Conference, 11-13 August 1998, Lincoln, New Zealand.
- Clausen, C.A. and V.W. Yang, 2003. Mold inhibition on unseasoned southern pine. In: Proceedings of 34th Annual Meeting of International Research Group on Wood Preservation, 18-23 May 2003, Brisbane, Australia.
- Coley-Smith, J.R., K. Verhoeff and W.R. Jarvis, 1980. The Biology of *Botrytis*. Academic Press, London.

- Delen, N., M. Yildiz and H. Maraite, 1984. Benzimidazole and dithiocarbamate resistance of *Botrytis cinerea* on greenhouses crops in Turkey. Med. Fac. L andbouww. Rijksuniv. Gent., 49: 153-161.
- Dix, N.J. and J. Webster, 1995. Fungal Ecology. Published by Chapman and Hall, London.
- Dudman, W.F., 1963. Sorbic Hydroxamic Acid, an Antifungal Agent Effective over a Wide pH Range. Applied Microbiol., 11: 362-364.
- EPA, 2006. Harpin Protein (006477) Fact Sheet.
- Erkan, M., T. Demir and S. Öz, 1997. Investigations on the sensitivities of gray mold (*Botrytis cinerea*) isolates on grapes against some fungicides. J. Turk. Phytopathol., 2623: 87-96.
- Haag, T.E. and D.F. Loncrini, 1984. Esters of para-hydroxybenzoic acid. In: Cosmetic and Drug Preservation. Kabara, J.J. (Ed.), New York: Marcel Dekker, pp: 63-77.
- Khan, S.H., J. Aked and N. Magan, 2001. Control of the anthracnose pathogen of banana (*Colletotrichum musae*) using antioxidants alone and in combination with thiabendazole or imazalil. Plant Pathol., 50: 601-608.
- Knight, S.C., V.M. Anthony, A.M. Brady, A.J. Greenl, S.P. Heaney, D.C. Murray, K.A. Powell, M.A. Schulz, C.A. Spinks, P.A. Worthington and D. Youle, 1997. Rationale and perspectives in the development of fungicides. Ann. Rev. Phytopathol., 35: 349-372.
- Liewen, M.B., 1991. Antifungal food additives. In: Arora, D.K., K.G. Mukerji and E.H. Marth (Eds.), H and Book of Applied Mycology, Vol. 3. Foods and Feeds. New York, USA: Marcel Dekker, pp: 541-52.
- Ntirampemba, G., B.E. Langlois, D.D. Archbold, T.R. Hamilton-Kemp and M.M. Bart, 1998. Microbial Population of *Botrytis cinerea* Inoculated Strawberry Fruit Exposed to four Volatil Compound. J. Food Prot., 61: 1352-1357.
- Palmer, C.L., R.K. Horst and R.W. Langhans, 1997. Use of bicarbonates to inhibit *in vitro* colony growth of *Botrytis cinerea*. Plant Dis., 81: 1432-1438.
- Reddy, M.V.B., P. Angers, A. Gosselin and J. Arul, 1998. Characterization and use of essential oil from *Thymus vulgaris* against *Botrytis cinerea* and *Rhizopus stolonifer* in Strawberry Fruits. Phytochemistry, 47: 1515-1520.
- Skirdal, I.M. and T. Eklund, 1993. Microculture model studies on the effect of sorbic acid on *Penicillium chrysogenum*, *Cladosporium cladosporioides* and *Ulocladium atrum* at different pH levels. J. Applied Bacteriol., 74: 191-195.
- Vallejo, I., F. Munoz, M. Carbu, L. Rebordinos, F.J. Fernandez-Acero and J.M. Cantrol, 2003. Study on fungicides resistance of *Botrytis cinerea* isolates from diseased strawberry plants. Arch. Phytopath. Plant Prot., 36: 13-67.
- Vicedo, B., M.O. Leyva, V. Flors, I. Finiti, G.A.D. Walters, M.D. Real, P. Garcia-Agustin and C.G. Bosch, 2006. Control of the phytopathogen *Botrytis cinerea* using adipic acid monoethyl ester. Arch. Microbiol., 184: 316-326.
- Wade, A. and P.J. Weller, 1994. Handbook of Pharmaceutical Excipients. The Pharmaceutical Press. London.