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# Isolation of Saponin from Dried Roots of Gypsophila simonii Hub. Mor

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**Abstract:** In the present research, a saponin was isolated from the roots of *Gypsophila simoni*. The structure was characterized by means of  ${}^{1}H$  NMR,  ${}^{13}C$  NMR, FTIR and EIMS. The findings indicate that the proposed structure of that saponin was as a new Gypsogenin ester ( $C_{31}$   $H_{51}$   $O_{3}$ ).

Key words: Gypsophila simonii, isolation, saponin structure

## INTRODUCTION

Saponins (a group of phytoanticipins) are present constitutively in plants and play important roles in plant defense (Figen, 2006). Some saponins of *Gypsophila* species are broadly used as a drug with extensive medical importance such as an expectorant and diuretic (Ansari and Has, 1989) and are used in treatments of hepatitis, gastritis and bronchitis (Mizutani *et al.*, 1984). A number of saponins were isolated from various *Gypsophila* species and investigated by many researchers (Henry *et al.*, 1991; Frechet *et al.*, 1991; Yaylı *et al.*, 1998; Acebes *et al.*, 1998; Bernadete and Parante, 2004).

Gypsophila simonii, local name Çöven, is an endemic growing plant at Çankırı province in Turkey (Davis, 1982). There is no study conducted on the gypsogenin ester saponins of this plant to date. In this study, we aim to present the isolation and structural elucidation of new unfamiliar saponin from the root of Gypsophila simonii.

### MATERIALS AND METHODS

**General:** Chemical materials that are used in the experiments are in analytical grade. Infrared spectra of the compounds were recorded between 4000 and 400 cm<sup>-1</sup> on Mattson 1000 FT-IR spectrometers which where calibrated using polystyrene bands. The resolution of IR spectrometers is 2 cm<sup>-1</sup> and the number of scan is 20. The samples were prepared as a KBr disc.

The melting point was determined in a glass capillary tube. Mass spectrum was recorded on an electron impact mass spectrometer from Research Institute of Tübitak (Turkey). <sup>1</sup>H NMR (400 MHZ, DMSO-d<sub>6</sub>) and <sup>13</sup>C NMR (100 MHZ, DMSO-d<sub>6</sub>) were explored in Middle East Technical University (METU).

**Plant material:** Çöven (*Gypsophila simonii*) was collected around Çankırı province, Türkiye, in June 1997 and identified by Professor Zeki Aytaç, Department of Biology, Gazi University. The root material was dried in a cool dark place and powdered in the Faculty of Pharmacy of Gazi University.

**Isolation and extraction:** Collected plant's roots were removed and dried. Approximately 3.5 kg of dried materials were placed into a cartridge and then extracted with chloroform in Soxhlet apparatus for 24 h. The cartridge was re-extracted with ethanol for extra 8 h and dried completely at the room temperature (Baytop, 1991).

The extracts containing saponins were evaporated by using a rotary evaporator (Biby, Oklahoma) at 40 rpm, without solvent at the reduced pressure. The dried extracts were dissolved in ethanol and applied on thin-layer chromatograms (TLC) ( $20\times20$ , silica gel  $G_{60}$  Art.7731) in the solvent system (1-Bu OH: 1-PrOH: HAc: H<sub>2</sub>O) (40: 20:7,5:30). Spots on TLC were detected by spraying with 10% H<sub>2</sub>SO<sub>4</sub> followed by heating at 110°C for 5-10 min. Spraying was done in order to identify the points of the separated compounds (Segal *et al.*, 1978).

**Acid hydrolysis of the saponins:** Each of the separated saponins (*Gypsogenin ester*) was refluxed for 7 h with 5% HCl (Okawa *et al.*, 2002). Third spot ( $R_f$ = 0.28) was run up for isolation and identification.

Sugar components were identified on Paper Chromatograms. The sugar in filtrate was identified as D-Glucose by comparison on PC (ethyl-acetate; pyridine; water, 12; 5; 4) with on authentic sugar (Boders, 1972).

### RESULTS AND DISCUSSION

The ethanolic extract obtained from the dried roots of Çöven (*Gypsophila simonii*) was purified on TLC

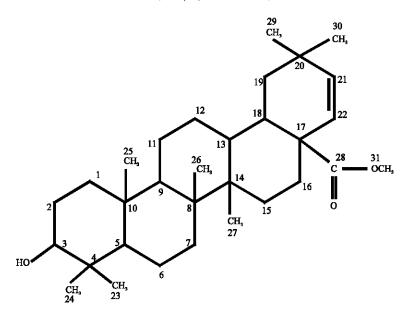


Fig. 1: Gypsogenin ester saponin

Table 1: 13C and 1H NMR data (δ ppm), DMSO-d<sub>6</sub>

| <sup>13</sup> C δ (ppm) | ¹H δ (ppm) |
|-------------------------|------------|
| 37.440                  | 1.60/1.62  |
| 26.208                  | 2.33/2.02  |
| 79.047                  | 4.17       |
| 37.440                  | -          |
| 45.392                  | 1.49       |
| 20.107                  | 2.11/1.85  |
| 30.454                  | 0.98       |
| 36.450                  | -          |
| 45.602                  | 1.73       |
| 45.392                  | <u>-</u>   |
| 25.337                  | 1.93/1.86  |
| 37.440                  | 1.14       |
| 45.000                  | 1.49       |
| 45.392                  | -          |
| 36.100                  | 1.86-1.45  |
| 73.481                  | 2.11-1.85  |
| 48.581                  | -          |
| 30.440                  | 3.06       |
| 34.450                  | 1.75/1.23  |
| 30.000                  | -          |
| 137.244                 | 5.44       |
| 133.533                 | 5.57       |
| 25.300                  | 1.93       |
| 20.100                  | 1.50       |
| 19.500                  | 0.93       |
| 20.250                  | 0.99       |
| 25.450                  | 1.28       |
| 168.000                 | -          |
| 35.450                  | 0.98       |
| 25.337                  | 0.93       |
| 69.648                  | 1.70       |

(R<sub>f</sub> = 0.28). Structure of the isolated Gypsogenin ester was characterized by <sup>1</sup>H NMR, <sup>13</sup>C NMR, IR and EIMS. The results were compared with the similar studies (Malaviya *et al.*, 1991; Zhao *et al.*, 1990; Delay *et al.*,

Table 2: FTIR spectral data (KBr, cm<sup>-1</sup>)

| Wave numbers (cm <sup>-1</sup> ) | Assignment                   |
|----------------------------------|------------------------------|
| 3400                             | -OH stretching               |
| 2967-2877                        | Aliphatic C-H stretching     |
| 1714                             | C = O stretching (for ester) |
| 1656-1618                        | C = C stretching             |
| 1450                             | C-H bending                  |
| 1190                             | -OCH₃ stretching             |

1997; Zhao *et al.*, 1997). It was concluded that the structure of Gypsogenin ester may have the form presented in Fig. 1.

Data of IR spectrum (KBr, cm $^{-1}$ ) exhibited absorptions at 3400 (-OH), 1656-1618 (C = C), 2967-2877 aliphatic C-H stretching and an ester (1714 carbonyl stretching (-C (O)-OH), 1190 asymmetric stretching).

The proton nuclear magnetic resonance, <sup>1</sup>H NMR (400 MHZ, DMSO-d<sub>6</sub> δ ppm), IR spectral data of compound (Fig. 1) showed the doubled vinylic proton [6.50-7.00].

The carbon-13 nuclear magnetic resonance, <sup>13</sup>C NMR (400 MHZ, DMSO-d<sub>6</sub> δ ppm), spectral of compound (Fig. 1) and its derivatives are shown in Table 1.

Gypsogenin ester saponin (Fig. 1); mp: 235°C (uncorrupted); IR spectrum of studied compound (Fig. 1); and its derivatives are shown in Table 2. Molecular ion peak was also observed at EIMS; m/z, [M $^+$ ]: 472. All these results confirm that the proposed structure of that saponin appears as a new one and is called as Gypsogenin ester ( $C_{31}$   $H_{51}$   $O_{3}$ ).

PC results showed the presence of D-Glucose by comparing their retention times with those of authentic sugar ( $R_G$  = 1.00, mp = 204°C).

### CONCLUSIONS

In recent years, although technology and medicine have developed extensively due to the decrease in natural richness and other drawbacks, some countries have made it obligatory to use natural products for many goals (Ertürk *et al.*, 2003). For these reason *Gypsophila* species are used for the treatment of various diseases.

The above studies the saponins were extracted from  $Gypsophila\ simonii\ (\Coven)\ dried\ roots\ and\ then\ separated its components by thin layer chromatography. NMR, FTIR and EIMS were carried out to investigate unknown saponin presents in this plant. The comparision of this investigation results with similar studies shows that the proposed structure of that saponin is a new type and could be named as Gypsogenin ester (<math>C_{31}\ H_{51}\ O_{3}$ ). However little is known about sugar presence and sugar linkage pattern in saponins of  $Gypsophila\ simonii$ . Paper chromotograms showed that, in  $Gypsophila\ simonii\ (\Coven)$ , D-Glucose was identified by comparison with on authentic sugar.

In conclusion, the studies on the saponins highlight the necessity for a comprehensive detailed investigation on the other gypsogenins saponin types which have a broad medical and industrial potential.

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## REFERENCES

- Acebes, B., A.M.A. Az-Lanza and M. Bernabeâ, 1998. A saponin from the roots of *Gypsophila bermejoi*. Phytochemistry, 49: 2077-2079.
- Ansari, A.K. and S. Has, 1989. Structural studies on a saponin isolated from the seeds *Nigella sativa*. Phytochemistry, 27: 377-379.
- Baytop, A., 1991. Farmasötik botanik ders kitabı. Yay No. 58, Ist Üniv. Eczacılık Fakültesi, Istanbul.
- Bernadete, P. da Silva and J.P. Parante, 2004. New steroidal saponins from rhizomes of *Cortus spiralis*. Z. Naturforsch, 59: 81-85.

- Boders, C. L., 1972. Descending paper chromatography of oligosaccharides. J. Chem. Educ., 49: 437-438.
- Davis, P.H., 1982. Flora of turkey and the east aegean islands. Vol. 7, Edinurgh Univ. Press, Edinburgh.
- Delay, C., J.A. Gavin, A. Aumelas, P.A. Bonnet and C. Roumestand, 1997. Isolation and structure elucidation of a highly haemolytic saponin from the Merck saponin extract using high-field gradientenhanced NMR techniques. Carbohydr. Res., 302: 67-78.
- Ertürk, Ö., H. Katı, N. Yaylı and Z. Demirbağ, 2003. Antimicrobial activity of *Viscum album* L. subsp. *abietis* (Wiesb), Turk. J. Biol., 27: 255-258.
- Figen, M.T., 2006. Saponins versus plant fungal pathogens. J. Cell Mol. Biol., 5: 13-17.
- Frechet, D., C. Bruno and D.S. Bertrand, 1991. Four triterpenoid saponins from dried roots of *Gypsophila* species. Phytochemistry, 30: 927-931.
- Henry, M., M. Rochd and B. Bennini, 1991. Biosynthesis and accumulation of saponins in *Gypsophila paniculata*. Phytochemistry, 30: 1819-1821.
- Malaviya, N., R. Pal and M.N. Khanna, 1991. Saponins from *Deutzia corymbosa*. Phytochemistry, 30: 2798-2800.
- Mizutani, K., K. Othani, J.-X. Wei and O. Tanaka, 1984. Saponins from *Anemone rivularis*. Planta Medica, 51: 327-331.
- Okawa, M., R.R. Yamaguchi, H. Delger, R. Tsuchihashi, T. Nohara, J. Kinjo, S. Isoda and Y. Ida, 2002. Five Triterpene Glycosides from *Oxytropis myriopylla*. Chem. Pharm. Bull., 50: 1097-1099.
- Segal, R., I. Milo-Goldzweig, D.V. Zaitschek and M. Noam, 1978. A facile method for detection of the genuine sapogenin from a saponin extract. Anal. Biochem., 84: 78-84.
- Yaylı, N., C. Baltacı, A. Zengin, M. Küçükislamoğlu and H. Genç, 1998. A triterpenoid saponin from *Cyclamen coum*. Phytochemistry, 48: 881-884.
- Zhao, L., W.M. Chen and Q. Fang, 1990. Triterpenoide saponins from *Anemone flaccida*. Planta Medica, 56: 92-93.
- Zhao, W.M., J.L. Wolfender, K. Hostettmann, K.F. Cheng, R.S. Xu and G.W. Qin, 1997. Triterpenes and triterpenoid saponins from *Mussaenda pubescens*. Phytochemistry, 45: 1073 1078.