http://www.pjbs.org



ISSN 1028-8880

# Pakistan Journal of Biological Sciences



# Physiological Responses of Broiler Chicks Fed Various Level of Dopamine and Zinc

Cyrus Farasati and Reza Tohidi

Agricultural Research Center, Jam-e-Jam St., Sepah Sq., P.O. Box 67145-1661, Kermanshah, Iran

Abstract: The influences of dopamine (DA) and zinc (Zn) on some physiological parameters of plasma such as growth hormone, cholesterol, 3-glyceride and growth rate, feed conversion ratio and abdominal fat of broilers between the ages of 21 to 42 day were investigated. The experiment was designed on a total of 180 commercial meat-type chicks as a randomized complete block with a 3×2×2 factorial arrangement of treatments with three levels of DA (0, 50 and 100 mg kg<sup>-1</sup>) and two levels of Zn (0 and 40 mg kg<sup>-1</sup>) in each sex. At the first day of trial, chicks were randomly distributed in 36 boxes at the same condition until day 15. Then, the treatments were administered in a low dosage for acclimating of chicks to them. From day of 21, the main dose of treatments was used. The effects of DA and Zn and interaction between them on cholesterol concentration of plasma were significant (p<0.05). But the effects on concentration of growth hormone (GH) and 3-glyceride of plasma were not significant. The maximum concentration of cholesterol in plasma was observed in male chicks administered 40 mg kg<sup>-1</sup> of Zn and the minimum concentration was related to female chicks administered 50 mg kg<sup>-1</sup> of DA. In this period, the main and interaction effects of all treatments on weight gain mean of chicks were significant. Also, Zn and DA and interaction of DA×sex improved feed conversion ratio (p<0.05). The results of this experiment confirmed the lipolytic effect of DA on plasma cholesterol and abdominal fat and also showed that consumption of 100 mg kg<sup>-1</sup> of DA plus 40 mg kg<sup>-1</sup> of Zn cause to improve growth performance of male chicks.

Key words: Dopamine, zinc, broiler, physiological parameters, performance

### INTRODUCTION

Modern broiler-type chickens have been intensively selected for rapid body weight and high meat yield. As a result they grow more rapidly and eat greater than layer-type chickens. It is well known that feeding behavior is regulated by the Central Nervous System (CNS). Especially the hypothalamus plays an important role in the feeding regulation in chickens through its content of monoamines (Yamasaki *et al.*, 2003).

Dopamine, a sympathomimetic amine vasopressor, is the naturally occurring immediate precursor of epinephrine. As a chemical messenger, DA is similar to adrenaline with a molecular formula of C<sub>8</sub>H<sub>11</sub>NO<sub>2</sub>. Several DA cell groups have been identified among a variety of hypothalamic region, suprachiasmatic nucleus, lateral hypothalamic area, paraventricular nucleus, lateral mamillaris nucleus and dorsomedial nucleus (Reiner et al., 1994; Chaiseha and El Halawani, 2005). Dopamine receptors have been identified in the chick brain (Sun and Reiner, 2000), although the intestinal dopaminergic system is believed to be a localized no neuronal system (Lucas-Teixeira et al., 2000). Dopamine D2 receptors have a role to inhibit of ceramide formation by growth hormone (Liu et al., 2002). DA administration has been reported to decrease intestinal contractions and motility in the chicks (Lot, 1993; Dive et al., 2000). DA

decreases Na<sup>+</sup>, K<sup>+</sup>-ATPase in rat jejunal enterocytes (Lucas-Teixeira *et al.*, 2000). Experiment is shown that DA can decrease significantly serum triglyceride and cholesterol level (Kimura *et al.*, 2002).

Zinc has been reported to act as a neuromodulator at excitatory synapses and has a considerable role in the stress response (Patricia and Pandolf, 2005). It is believed that Zn is essential in all aspects of immunity (Chandra and Dayton, 1982; Sherman, 1992) and function through its association with the enzymes is critical for the integrity of the cells involved in the immune response (Dardenne *et al.*, 1985). Reports by Miller *et al.* (1968) and Swinkels *et al.* (1994) showed that diets low in Zn lead to depressed appetite resulting in lowered feed intake and reduced weight gain.

Because of not enough investigations on effect of dopamine and zinc on broilers function, the present experiment was performed to describe the effect of these treatments on some physiological parameters, feed conversion ratio and abdominal fat of broiler chicks.

## MATERIALS AND METHODS

**Location of experiment:** This study was conducted in Poultry Research Section of Animal Research Station belonged to Ferdowsi University of Mashad, located at Iran.

Table 1: Composition of treatments in the broiler diet

Treatments	Ingredient (per kg of diet)
$Zn_0D_0$	Control
$Zn_1D_0$	40 mg kg <sup>-1</sup> zinc sulfate
$Zn_0D_1$	50 mg kg <sup>-1</sup> dopamine
$Zn_1D_1$	50 mg kg <sup>-1</sup> dopamine and 40 mg kg <sup>-1</sup> zinc sulfate
$Zn_0D_2$	$100~{ m mg~kg^{-1}}$ dopamine
$Zn_1D_2$	100 mg kg <sup>-1</sup> dopamine and 40 mg kg <sup>-1</sup> zinc sulfate

Chickens and diets: One hundred eighty 1-day-old Ross broiler males and females were used in this experiment. At the first day of trail, chicks were randomly distributed in 36 boxes at the same condition until day 15. After that, the treatments in low dosage were offered and from day 21 to 42 the main dose of them was administered. The ration used during the trail was regulated through NRC (1994) with 21.8% CP and 3150 kcal kg<sup>-1</sup> ME. The level of feed intake was recorded daily and the chicks were weighed weekly.

**Blood sampling:** A volume of 5 mL blood sample was collected into an ethylenediminetetraacetic acid-coated tube from the brachial vein of each chick. The samples were centrifuged at 700 x g for 15 min at 20°C. Plasma was kept at -80°C until measurement. Enzymatic method by ELISA Reader was used to measure of GH concentration, 3-glyceride and cholesterol.

Experimental design and statistical analysis: The collected data were statistically analyzed using General Linear Models procedure (GLM) of SAS software (1995). A randomized complete block design was utilized with a 3×2×2 factorial arrangement of treatments with three levels of dopamine (0, 50 and 100 mg kg<sup>-1</sup>) and two levels of zinc (0 and 40 mg kg<sup>-1</sup>) in each sex (Table 1). The independent variables were treatment. All other variables, such as body weight, feed intake, feed conversion, GH concentration, 3-glyceride, cholesterol and Abdominal fat were considered as dependent variables.

### RESULTS AND DISCUSSION

Dopamine and zinc had a significant effect on cholesterol concentration of plasma (p<0.05). However, no difference was found among the effect of treatments on 3-glyceride and abdominal fat. The interaction effects of sex×Zn, Zn×DA and sex×Zn×DA on cholesterol concentration of plasma were significant (p<0.05). This result shows that different levels of Zn in each sex and separate levels of DA have different responses (Table 2 and 3). The maximum concentration of cholesterol in plasma was observed in male chicks and related to diet contained  $Zn_1D_0$  and the minimum was related to females administered  $Zn_0D_1$ . The least abdominal fat was observed

Table 2: ANOVA of effects of treatments on physiological parameters

	Growth			
Source of	hormone	Cholesterol	3-glyceride	Abdominal
variation	$(ng mL^{-1})$	(mg 100 mL <sup>-1</sup> )	(mg 100 mL <sup>-1</sup> )	fat (%)
Sex	4.218 ns	239.686ns	7.823ns	$0.450 \mathrm{ns}$
Zn	$0.649 \mathrm{ns}$	1587.003*	127.379ns	0.046ns
DA	2.415 ns	984.946*	352.300ns	$0.147 \mathrm{ns}$
Sex×Zn	2.445 ns	1711.135**	46.931 ns	0.056ns
$Sex \times DA$	2.043ns	420.346ns	188.281ns	0.648ns
$Zn\times DA$	3.000 ns	1630.480**	114.218ns	$0.019 \mathrm{ns}$
$Sex \times Zn \times DA$	0.339 ns	948.126*	375.585ns	$0.420 \mathrm{ns}$
Block	$0.430 \mathrm{ns}$	94.410ns	138.900ns	1.386 ns
R-square	0.434	0.641	0.438	0.451

<sup>\*</sup>Significant at p<0.05, \*\*Significant at p<0.01, ns: Non significant

Table 3: The effects of DA and Zn supplementations on the physiological parameter (Mean±SEM)

	Growth			
	hormone	Cholesterol	3-glyceride	Abdominal
Treatments	$(ng mL^{-1})$	$(mg\ 100\ mL^{-1})$	$(mg\ 100\ mL^{-1})$	fat (%)
Male				
$Zn_0D_0$	$6.53\pm0.78^{a}$	$129.30\pm4.10^{bcd}$	117.00±10.60a	$0.83\pm0.03^{ab}$
$Zn_1D_0$	$6.27 \pm 0.84^a$	158.20±8.40a	100.50±19.60a	$1.17\pm0.20^{ab}$
$Zn_0D_1$	$6.90\pm1.20^a$	$115.90\pm0.80^{cd}$	97.10±5.40°	$1.07\pm0.22^{ab}$
$Zn_1D_1$	5.40±0.65a	138.80±7.40 <sup>abc</sup>	$81.30\pm8.00^a$	$0.50\pm0.21^{b}$
$Zn_0D_2$	5.77±0.48 <sup>a</sup>	$147.80\pm2.70^{ab}$	94.20±4.10 <sup>a</sup>	$0.63\pm0.12^{ab}$
$Zn_1D_2$	6.17±1.84°	$119.70\pm7.30^{cd}$	$80.80\pm6.90^a$	$1.13 \pm 0.18^{ab}$
Female				
$Zn_0D_0$	$5.03\pm0.34^{a}$	138.50±16.10 <sup>abc</sup>	105.30±14.50a	$1.10\pm0.25^{ab}$
$Zn_1D_0$	5.55±0.25a	$124.20\pm2.10^{bcd}$	89.40±15.70 <sup>a</sup>	$1.63\pm0.50^{ab}$
$Zn_0D_1$	$7.27 \pm 1.27^{a}$	$105.30\pm3.80^{d}$	94.80±13.70 <sup>a</sup>	$2.00\pm1.17^{ab}$
$Zn_1D_1$	7.70±1.41°	132.80±4.40 <sup>abcd</sup>	102.30±3.90°	$3.17\pm2.23^a$
$Zn_0D_2$	5.47±0.57a	128.80±17.80 <sup>bcd</sup>	102.80±9.80 <sup>a</sup>	$1.27\pm0.33^{ab}$
$Zn_1D_2$	7.77±1.95°	121.30±2.90 <sup>bcd</sup>	86.30±5.30 <sup>a</sup>	$0.97\pm0.03^{ab}$
Means with	in the same	column without	a common supe	erscript differ

Means within the same column without a common superscript differ  $(p \le 0.05)$ 

in Zn<sub>1</sub>D<sub>1</sub> treatment (males) which can be explained partly by lipolytic effect of catecholamines. These results are agreed with Catherine et al. (1984), Stiles et al. (1984), Buyse et al. (1990) and Rosebrough (1997). Some researches have shown that  $\beta$ -adrenergic complex such as DA can improve feed conversion ratio, decrease carcass fat, increase muscle weight and growth of chicks (Catherine et al., 1984; Colin, 1989; Adeola and Ball, 1992). Some medium composites (i.e., \beta-adrenergic receptors and cyclic adenosine mono phosphate) are necessary for activation of catecholamines (Goldberg and Haddox, 1977). β-adrenergic agonists inhibited the synthesize of fatty acids in chicks and stimulated lypolising of fat tissue (Catherine et al., 1984). Cincotta and Meier (1989) reported that administrations (injections and in feed) of bromocriptine, a DA agonist, reduced body fat stores, plasma total cholesterol and triglyceride concentrations in several rodent species. It was also shown that serum cholesterol levels significantly declined using DA (Ormiston *et al.*, 2004).

The main and interaction effects of treatments on GH concentration of chickens were not significant (Table 2 and 3). These results are different with some previous studies that indicated administration of nor-epinephrine,

epinephrine, DA and β-agonists increase the range of secretory pulses of GH. Also, releasing some factors from hypothalamus that control the secretion of GH is affected by adrenergic compounds through α-adrenergic receptors (Chester, 1986; Buyse *et al.*, 1990). The different result of present study may be due to not enough dosage of DA to affect GH concentration of plasma. However, Chang *et al.* (2003) found that DA had no effect on growth of broiler chicks at 6, 9 and 12 days of age. There is no demarcation between neurotransmitters and neuropeptides because they may coexist in the same neuron in different areas of the CNS and in mediobasal hypothalamus. So, both amine and amino acid neurotransmitters have roles in the control of GH release. In a series of studies, the increase in GH was shown to be induced by DA agonists (Muller, 1997).

As shown in the Table 4 and 5, the main effects of DA and Zn and interaction of sex×DA on feed conversion ratio from 35 to 42 days were significant (p<0.05). The minimum and maximum of feed conversion ratio was observed in male chicks used Zn<sub>1</sub>D<sub>2</sub> and females administered Zn<sub>1</sub>D<sub>1</sub>, respectively. These results can be caused by decreasing and increasing of feed intake by males and females, respectively. DA may play a role in gut-brain axis regulation (Sun and Reiner, 2000) and within the intestinal mucosal cells, act locally as a regulator of intestinal function (Lucas-Teixeira *et al.*, 2000). Disregulation of catecholamines has been associated with abnormal behaviours such as feed intake by the animal (Cheng *et al.*, 2001).

Gether et al. (2001) reported that Zn<sup>2+</sup> acts as a potent non-competetive inhibitor of DA uptake mediated by DA transporter. Zn2+ binding site is responsible for allosteric modulation of D2 dopamine receptor (Liu et al., 2006). The effect of Zn on chick weight, livability and humoral or cellular immunity was not significant in the study of Kidd et al. (1992). The level of Zn in the diet did not significantly influence broiler growth performance during the entire 7 week of the study of Bartlett and Smith (2003). However, Zn levels were apparently not low enough to affect feed intake or weight gain in that study. In the study of Chang et al. (2003), after peritoneal injection of DA to the chicks at day 6, there were no significant differences in body weight gain, feed consumption and feed conversion between 7 to 15 days. Also, Feeding diets supplemented with various levels of DA during 7 to 15 days had no significant effects on the weights of visceral organs, intestinal density, small intestinal and cecal weights and small intestinal and cecal lengths adjusted for feed-deprived body weight. These different results show that the effectiveness of DA depends on various levels of that and period of administration.

Table 4: ANOVA of effects of treatments on feed conversion ratio

	Days				
Source of variance	21 to 28				
Sex	0.0005ns	0.2607*	0.0000ns	0.3180	
Zn	0.0003ns 0.0108ns	0.2007 0.0069ns	0.0000ns 0.2204*	0.3180 0.0020ns	
	0.0100110	0.0000110	· · ·	0.0020110	
DA	0.0002ns	0.0963ns	0.2315*	0.1620ns	
Sex×Zn	0.0043ns	0.0000ns	0.1657ns	0.0130ns	
Sex×DA	0.0004ns	0.101 <i>6</i> ns	0.1764*	0.1480ns	
Zn×DA	0.1158 ns	0.0452ns	0.1015 ns	$0.0520 \mathrm{ns}$	
Sex×Zn×DA	0.0055ns	0.1023ns	0.1537 ns	$0.0520 \mathrm{ns}$	
Block	0.0021 ns	0.0923 ns	0.0291 ns	0.0980 ns	
R-square	0.8719	0.8083	0.6700	0.7520	

<sup>\*</sup> Significant at p<0.05, ns: Non significant

Table 5: The effects of DA and Zn supplementations on the feed conversion ratio (Mean±SEM)

	Days			
Treatments	21 to 28	28 to 35	35 to 42	42 to 49
Male				
$Zn_0D_0$	1.83±0.06 <sup>a</sup>	$2.28\pm0.40^{ab}$	$2.32\pm0.10^{ab}$	$2.18\pm0.15^{b}$
$Zn_1D_0$	1.83±0.10 <sup>a</sup>	$1.86\pm0.10^{b}$	$1.94\pm0.11^{bcde}$	$2.28\pm0.35^{b}$
$Zn_0D_1$	1.82±0.05a	$1.98\pm0.08^{b}$	$1.77\pm0.03^{de}$	$2.62\pm0.13^{ab}$
$Zn_1D_1$	1.84±0.09 <sup>a</sup>	$1.95\pm0.18^{b}$	$1.82\pm0.17^{\text{cde}}$	2.40±0.09 <sup>b</sup>
$Zn_0D_2$	1.94±0.07ª	$2.09\pm0.18^{ab}$	$2.09\pm0.16^{abcd}$	$2.63\pm0.25^{ab}$
$Zn_1D_2$	1.95±0.17a	$1.85\pm0.06^{b}$	1.68±0.17°	$2.28\pm0.23^{b}$
Female				
$Zn_0D_0$	1.96±0.09a	$2.17 \pm 0.17^{ab}$	$2.32\pm0.05^{ab}$	$2.72\pm0.15^{ab}$
$Zn_1D_0$	$1.88\pm0.06^a$	$2.14\pm0.11$ ab	$2.41\pm0.15^a$	$2.55\pm0.05^{ab}$
$Zn_0D_1$	1.95±0.13 <sup>a</sup>	$2.22\pm0.21$ ab	2.42±0.11 <sup>a</sup>	$2.54\pm0.06^{ab}$
$Zn_1D_1$	$2.01\pm0.04^a$	$2.76\pm0.40^a$	$2.06\pm0.15^{abcde}$	$2.64\pm0.03^{ab}$
$Zn_0D_2$	$2.03\pm0.08^a$	$2.58\pm0.09^{ab}$	$2.19\pm0.14^{abc}$	$2.83\pm0.05^{ab}$
$Zn_1D_2$	1.92±0.09ª	2.39±0.33ab	2.05±0.03abcde	3.13±0.36 <sup>a</sup>

Means within the same column without a common superscript differ  $(p{<}0.05)$ 

# CONCLUSION

Administration of DA to broiler chicks decrease carcass fat and improve carcass quality. The use of 100 mg DA with 40 mg Zn per kg of diet improves performance of male chicks. However, the effects of these treatments are different on males and females. Whereas, Zn supplementation can improve immune system and under heat stress condition the immunity of chicks may decrease, so more studies on the effect of different levels of Zn in various physiological conditions of chicks are necessary.

### ACKNOWLEDGMENTS

The authors would like to thank the Animal Science Department of Agricultural College of Mashad University for rendering technical and financial support.

# REFERENCES

Adeola, O. and R.O. Ball, 1992. Hypothalamic neurotransmitter concentrations and quality in stressed pigs offered excess dietary tryptophan and tyrosine. J. Anim. Sci., 70: 1888-1894.

- Bartlett, J.R. and M.O. Smith, 2003. Effects of different levels of zinc on the performance and immunocompetence of broilers under heat stress. Poult. Sci., 82: 1580-1588.
- Buyse, J., E. Decuypere, G. Huyghebaert and M. Herremans, 1990. The effect of clenbuterol supplementation on growth performance and on plasma hormone and metabolite levels of broilers. Poult. Sci., 70: 993-1002.
- Catherine, A.R., H. Ricks, P. Dalrimple, K. Baker and D.L. Ingle, 1984. Use of β-agonist to alter fat and muscle deposition in steers. J. Anim. Sci., 59: 1247-1255.
- Chaiseha, Y. and M.E. El Halawani, 2005. Neuroendocrinology of the female turkey reproductive cycle. J. Poult. Sci., 42: 87-100.
- Chandra, R.K. and D.H. Dayton, 1982. Trace element regulation of immunity and infection. Nutr. Res., 2: 721-733.
- Cheng, H.W., G. Dillworth, P. Singleton, Y. Chen and W.M. Muirt, 2001. Effects of group selection for productivity and longevity on blood concentration of serotonin, catecholamines and corticosterone of laying hens. Poult. Sci., 80: 1278-1285.
- Chang, S.C., M.J. Lin, J. Croom and Y.K. Fan, 2003. Administration of triiodothyronine and dopamine to broiler chicks increases growth, feed conversion and visceral organ mass. Poult. Sci., 82: 285-293.
- Chester, J., 1986. Fundamental of Comparative Vertebrate Endocrinology. Plenum, New York.
- Cincotta, A.H. and A.H. Meier, 1989. Reductions of body fat stores and total plasma cholesterol and trigelyceride concentrations in several species by bromocriptine treatment. Life Sci., 45: 2247-2254.
- Colin, G.S., 1989. Research note: Influence of β-agonist on plasma concentration of growth hormone in broiler chickens on low plane of nutrition. Poult. Sci., 68: 1015-1018.
- Dardenne, M., W. Savino, S. Borrih and J.F. Bach, 1985.
  A zinc dependent epitope of the molecule of thymulin, a thymic hormone. Proc. Natl. Acad. Sci. USA., 82: 7035.
- Dive, A., F. Foret, J. Jamart, P. Bulpa and E. Installe, 2000. Effect of dopamine on gastrointestinal motility during critical illness. Intensive Care Med., 26: 901-907.
- Gether, U., L. Norrequad and C.J. Loland, 2001. Delineating structure-function relationships in the dopamine transporter from natural and engineered Zn<sup>2+</sup> binding site. Life Sci., 68: 2187-2198.
- Goldberg, N.D. and M.K. Haddox, 1977. Cyclic GMP metabolism and involvement in biological regulation. Ann. Rev. Physiol., 46: 823-896.

- Kidd, M.T., N.B. Anthony, Z. Johnson and S. Lee, 1992.
  Effect of zinc methionine supplementation on the performance of mature broiler breeders. J. Applied Poult. Res., 1: 207-211.
- Kimura, T., M. Hasegawa and O. Takano, 2002. The effect of dopamine on serum lipid concentration after propofol administration. Masui, 51: 286-288.
- Liu, G., L. Robillard, B. Banihashemi and P.R. Albert, 2002. Growth hormone-induced diacylglycerol and ceramide formation via Galpha i3 and Gbeta gamma in GH4 pituitary cells. Potentiation by dopamine-D2 receptor activation. J. Biol. Chem., 277: 48427-48433.
- Liu, Y., M.M. Teeter, C.J. DuRand and K.A. Neve, 2006. Identification of a Zn<sup>2+</sup> binding site on the dopamine D2 receptor. Biochem. Biophys. Res. Commun., 339: 873-879.
- Lot, T.Y., 1993. Effects of dopamine on the gastrointestinal tract of chicks. J. Pharm. Pharmacol., 45: 892-895.
- Lucas-Teixeira, V., M.A. Vieira-Coelho and P. Soares-da-Silva, 2000. Food intake abolishes the response of rat jejunal Na<sup>+</sup>, K<sup>+</sup>-ATPase to dopamine. J. Nutr., 130: 877-881.
- Miller, E.R., R.W. Leucke, D.E. Ullrey, B.U. Baltzer, B.L. Bradley and J.A. Holfer, 1968. Biochemical, skeletal and allometric changes due to zinc deficiency in the baby pig. J. Nutr., 95: 278-286.
- Muller, E.E., 1997. Cholinergic function and neural control of GH secretion. A critical re-appraisal. Eur. J. Endocrinol., 137: 338-342.
- National Research Council (NRC), 1994. Nutrient Requirements of Poultry. 9th Edn., National Academy Press, Washington, DC.
- Ormiston, T., O.M. Wolkowitz, V.I. Reus, R. Johnson and F. Manfredi, 2004. Hormonal changes with cholesterol reduction: A double-blind pilot study. J. Clin. Pharm. Ther., 29: 71-73.
- Patricia, S.B. and P. Pandolf, 2005. Antioxidant defenses and lipid peroxidation in the Cerebral, cortex and Malathion and/or zinc chloride. Toxicology, 207: 283-291.
- Reiner, A., D.J. Karle, K.D. Adderson and L. Medina, 1994. Catecholaminergic Perikarya and Fibers in the Avian Nervous System. In: Phylogeny and Development of Catecholamine Systems in the CNS of Vertebrates. Smeets, W.J. and A.J. Reiner (Eds.), Cmbridge University Press. Cambridge. England, pp: 135-181.
- Rosebrough, R.W., 1997. Nutritional effects on neurotransmitter in the broiler chicken. Comp. Biochem. Physiol., 107: 573-580.
- SAS, 1995. Procedures Guide, Version 6. 3rd Edn., SAS Institute Inc., Cary, NC.

- Sherman, A.R., 1992. Zinc, copper and iron nutrition and immunity. J. Nutr., 122: 604-609.
- Stiles, G.L., M.C. Caron and R.J. Lfkowitz, 1984. β-adrenergic receptors: Biochemical mechanism of physiological regulation. Physiol. Rev., 64: 661-743.
- Sun, Z. and A. Reiner, 2000. Localization of dopamine D1A and D1B receptor mRNAs in the forebrain and midbrain of the domestic chick. J. Chem. Neuroanat., 19: 211-224.
- Swinkels, J.W., E.T. Kornegay and M.W. Verstegen, 1994. Biology of zinc and biological value of dietary organic zinc complexes and chelates. Nutr. Res. Rev., 7: 129-149.
- Yamasaki, I., K. Sashihara, T. Takagi, T. Nakanishi, M.D. Denbow and M. Furuse, 2003. Comparison of hypothalamic monoamine contents of broiler and layer-type chickens at prehatch and posthatch. J. Poult. Sci., 40: 282-289.