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Leptin Gene Polymorphism in Iranian Native Golpayegani and Taleshi Cows

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Abstract: The objective of the present investigation was to study genetic variations in the exon II of leptin gene in Iranian Golpayegani and Taleshi cows. Blood samples were collected from 76 Golpayegani and 64 Taleshi cows randomly selected from Delijan city (Esfahan province) and Talesh city (Guilan province) in Iran, respectively. Genomic DNA was extracted from 100 micro litters of blood using guanidium thiocyanate-silica gel method. A 94 bp fragment from exon II of the bovine leptin gene was amplified using the polymerase chain reaction. Digestion of PCR products with *Bsp13I* restriction enzyme differentiated C and T alleles. The T allele frequencies were 0.29 and 0.45 in Golpayegani and Taleshi breeds, respectively. The frequencies of the genotypes CC, CT and TT were 0.42, 0.58 and 0 in Golpayegani and 0.36, 0.36 and 0.27 in Taleshi cows. The average heterozygosities were 0.41 and 0.50 in Golpayegani and Taleshi breeds, respectively. The χ^2 test did not confirm the Hardy-Weinberg equilibrium in both populations.

Key words: Leptin, polymorphism, PCR-RFLP, Golpayegani, Taleshi

INTRODUCTION

Economical traits are among quantitative traits that are controlled by many genes each having a small effect (Gelderman, 1997). The major gene model suggests that only a few genes may account for relatively large proportion of the genetic variation (Lande, 1981), such major genes being the genes usually involved in the biology of a trait and are the candidate genes for marker identification. There is also the possibility that major genes may be linked with some Quantitative Trait Loci (QTL) contributing to a major part of the variation in traits (Fruhbeck *et al.*, 1998). One of the candidate genes for marker assistant selection is leptin gene. Leptin is a 16-kDa protein that is synthesized by adipose tissue and is involved in regulation of feed intake, energy balance, fertility and immune functions (Fruhbeck *et al.*, 1998). Plasma leptin levels in cattle and sheep increase linearly with increased body fat mass and with increased energy balance (Blache *et al.*, 2002; Ehrhardt *et al.*, 2000). Leptin gene expressed in a variety of tissues including adipose tissue, placenta, mammary glands, skeletal muscles, gastric mucosa, brain and pituitary glands. It seems that leptin has a large effect in coordinating whole body energy metabolism and may be classified as a metabolism modifier (Houseknecht *et al.*, 1998). It has been shown that leptin gene influences milk performance in cattle (Liefers *et al.*, 2002) and reproduction in beef cattle (Almeida *et al.*, 2003).

In cattle, the leptin gene is located on chromosome 4 and consists of three exons (Pomp *et al.*, 1997). Genetic differences in the leptin gene were first observed in mice ob/ob mice lack functional leptin and are hyperphagic, obese and infertile (Hamann and Matthaei, 1996). Several polymorphisms in this gene have been found in cattle. Restriction fragment length polymorphism of the bovine leptin gene was reported for first time by Lein *et al.* (1997).

Two RFLP were identified in exon 3: *NruI* (a C/T substitution resulting in a change from valine to alanine) (Lagonigro *et al.*, 2003) and *HphI* (a C/T substitution resulting in a change from alanine to valine) (Haegeman *et al.*, 2000). In exon 2, three RFLP were described: *Clal* (an A/T substitution resulting in an amino acid change from tyrosine to phenylalanine) (Lagonigro *et al.*, 2003), *Kpn2I* (*Bsp13I*) (a C/T substitution resulting in an amino acid change from arginine to cysteine) (Buchanan *et al.*, 2002) and *Sau3AI* (Pomp *et al.*, 1997). The cysteine/arginine change in exon 2 (*Kpn2I*) is a non-conserved substitution and is more likely to alter the functioning of the leptin hormone (Buchanan *et al.*, 2002). In livestock, such polymorphisms may be associated with, or linked to, economic traits, which are controlled by many genes each having a small effect (Gelderman, 1997). Leptin is related to both energy metabolism and reproduction and it was shown that leptin polymorphisms had significant effect on calving interval and weight at first calving in beef cows (Almeida *et al.*, 2003). These observations suggest that leptin may be a

candidate gene for some of the economically important production traits in dairy cattle. Therefore, the objective of the present investigation was to study the genetic variations in the exon 2 of leptin gene of Iranian native Golpayegani and Taleshi cows.

MATERIALS AND METHODS

Animals and DNA extraction: Blood samples were randomly collected from 76 Golpayegani and 64 Taleshi cattle breeds from Deligan city (Esfahan province) and Talesh city (Guilan province) in Iran, respectively. Genomic DNA was extracted from 100 μ L of blood using guanidium thiocyanate-silica gel method (Boom *et al.*, 1990). Quality and quantity of DNA were measured by spectrophotometer by taking the optical density at wavelength of 260 and 280 nm, respectively.

PCR reaction: One microliter of DNA was amplified in a total volume of 25 μ L PCR mix using the Biometra T Personal Ver: 1.11 thermocycler. The PCR mix contained: 2.5 μ L PCR buffer 10-X (200 mM $(\text{NH}_4)_2\text{SO}_4$, 0.1 mM Tween 20%, 750 mM Tris-HCl pH = 8.8), 2.5 mM MgCl_2 , 200 μ M dNTPs and 10 pM from each primer, 1 U *Taq* DNA polymerase and 11 μ L ddH_2O . The thermal cycling profile consist of an initial denaturation step of 5 min at 94°C, followed by 35 cycles 45sec at 94°C, 45 sec at 58°C, 45 sec at 72°C and final extension step of 10 min at 72°C. A 94 bp fragment from exon II of the bovine leptin gene was amplified using the following primers described by :

LeptF: 5'ATGCGCTGTGGACCCCTGTATC 3'
LeptR: 5'- TGGTGTTCATCCTGGACCTCC-3'

Products of amplification were recognized by electrophoresis on 1.5% agarose gel stained with ethidium bromide.

RFLP: Five micro liters of PCR products were incubated for 5 h at 50°C with three units of *Bsp*13I restriction enzyme. The C allele was cleaved into two fragments of 75 and 19 bp, while the T allele remained uncut at 94 bp (Buchanan *et al.*, 2002). Digestion products were separated by electrophoresis on non-denaturing 8% polyacrylamide gel stained with AgNO_3 .

Statistical analysis: The frequencies of genotypes, alleles, mean expected and observed heterozygosities and χ^2 test were calculated using PopGene32 (ver 1.31) program [<http://cc.oulu.fi/~jaspi/popgen/popdown.htm>].

RESULTS AND DISCUSSION

Iranian cattle (*Bos indicus* and *Bos taurus*) have unique features like adaptability to extreme climatic conditions, subsistence on poor feed and fodder and better resistance capabilities to withstand environmental stress and tropical disease. There are several diverse cattle breeds in Iran that are primarily being used for drought milk and meat. Although, cattle in Iranian is the most important livestock species and plays a major role in agricultural economy, yet population of some of the important cattle breeds is either declining or breed characters are being diluted under the present production system. To avoid further loss of important gene/gene-pool and preserve maximum amount of genetic diversity, an objective breed classification based on genetic uniqueness is of priority.

This study was done to investigate leptin polymorphism in two Iranian cattle breeds. Present results in leptin polymorphism showed two alleles (C and T) and three genotypes (CC, CT and TT) (Fig. 1) for leptin exon II gene. The CC and CT genotypes were observed in both Golpayegani and Taleshi breeds whereas the TT genotype was observed only in the Taleshi cattle breed. The higher observed frequency for CT genotype was 0.58 in Golpayegani breed (Table 1).

Although the TT genotype was not observed in Golpayegani breed, its frequency was relatively high (0.27) in Taleshi cattle breed. Similar results were reported by Choudhary *et al.* (2005), who did not detected TT genotype in the Haryana, Sahiwal, Gir and Nimari cattle breeds, but reported comparatively high TT genotype frequency (0.30) in Jersey cattle.

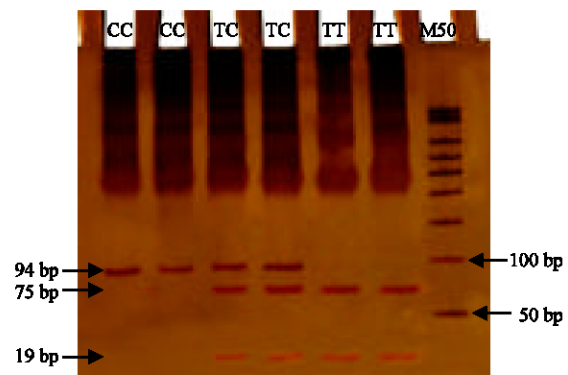


Fig. 1: Acrylamide electrophoresis of different leptin genotypes. Molecular marker is M50 (From top to bottom 500, 450, 400, 350, 300, 250, 200, 150, 100 and 50 bp)

Table 1: Observed, expected, average heterozygosities and χ^2 test and genotype frequencies of leptin exon 2 for the Iranian native cattle breed

Breed	Observed heterozygosity	Expected heterozygosity	Average heterozygosity	χ^2	Genotype frequencies		
					TT	TC	CC
Golpayegani	0.58	0.41	0.41	12.28*	0.00	0.58	0.42
Taleshi	0.36	0.50	0.50	4.97**	0.27	0.36	0.36

*Significant (p<0.01); **Significant (p<0.05)

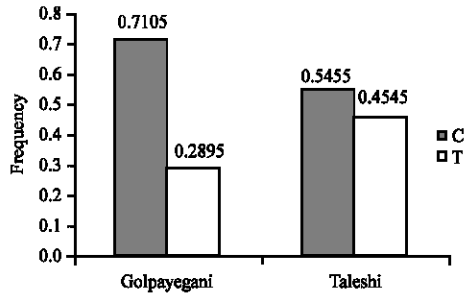


Fig. 2: Allelic frequencies of the leptin exon 2 for two Iranian native cattle breed

For the allelic frequencies, the frequency of T allele was lower than C allele in both cattle breeds (Fig. 2). Present results agree with Buchanan *et al.* (2002) who had reported C allele frequencies of 0.66 in charolias cattle and 0.68 in Simmental cattle and Madeja *et al.* (2004) who found frequency of 0.66 in Polish Black and White cattle. Buchanan *et al.* (2003) reported 0.55 for C allele frequency in Brown Swiss cattle. Present findings for Taleshi cattle (*B. Indicus*) breed is in contrast to Choudhary *et al.* (2005), who reported that the T allele found only in *B. Taurus* cattle breeds. The polymorphisms of leptin gene were reported by several studies. Heravi *et al.* (2006) found two genotypes, AA and AB, in Irania Holstein cows, which had the frequencies of 0.89 and 0.11, respectively. Madeja *et al.* (2004) investigated three restriction fragment length polymorphisms (RFLP): *HphI*, *Kpn2I* (*Bsp13I*) and *Sau3AI* in the leptin gene in Polish Black and White cattle. They reported the allelic frequencies of 0.54 (C) and 0.46 (T) for *Kpn2I*; 0.66 (C) and 0.34 (T) for *HphI* and 0.86 (A), 0.11 (B) and 0.03 (C) for *Sau3AI*. Konfortov *et al.* (1999) using sequence analysis detected 20 polymorphisms in both introns and exons of leptin gene in a diverse panel of cattle. Liefers *et al.* (2003) studied three microsatellite markers (R4C, RFLP1, A59V and BM1500) in leptin gene. Lagonigro *et al.* (2003) reported a new non-conservative mutation in exon II of the leptin gene. They used five different breeds and the frequency of the rare allele ranged from 0 to 15.2%. Pomp *et al.* (1997) using PCR-based polymorphism showed a rare polymorphism in the leptin gene. They reported that the *Sau3AI* restriction enzyme digested the PCR product four times instead of three times. These cattles are excellent sources of biological information for

studies on genetic characterization in Iran. These breeds showed a middle degree of genetic variability and deviation from H-W equilibrium for the leptin locus (Table 1); this might be explained by breeding method, which is used in these stations. With respect to low number of population, the inbreeding rate is high so heterozygosity and genetic variability is not high in the studied populations.

REFERENCES

- Almeida, S.E., E.A. Almeida, J.C.F. Moraes and T. Weimer, 2003. Molecular marker in the LEP gene and reproductive performance of beef cattle. *J. Anim. Breed. Genet.*, 120: 106.
- Blache, D., R.L. Tellam, L.M. Chagas, M.A. Blackberry, P.E. Vercoc and G.B. Martin, 2000. Level of nutrition affects leptin concentrations in plasma and cerebrospinal fluid in sheep. *J. Endocrinol.*, 165: 625-637.
- Boom, R., C.J.A. Sol and M.M.M. Salimans, 1990. Rapid and simple method for purification of nucleic acids. *J. Clin. Microbiol.*, 28: 495-503.
- Buchanan, F.C., C.J. Fitzsimmons, A.G. Van Kessel, T.D. Thue, D.C. Winkelman-Sim and S.M. Schmutz, 2002. Association of a missense mutation in the bovine leptin gene with carcass fat content and leptin mRNA levels. *J. Genet. Sel. Evol.*, 34: 105-116.
- Buchanan, F.C., A.G. Van Kessel, C. Waldner, D.A. Christensen, B. Laarveld and S.M. Schmutz, 2003. An association between a leptin single nucleotide polymorphism and milk and protein yield. *J. Dairy. Sci.*, 86: 3164-3166.
- Choudhary, V., P. Kumar, T.K. Bhattacharya, B. Bhushan and A. Sharma, 2005. DNA polymorphism of leptin gene in *Bos indicus* and *Bos taurus* cattle. *J. Genet. Mol. Biol.*, 28: 740-742.
- Ehrhardt, R.A., R.M. Slepatis, J. Siegal-Willott, M.E. Van Amburgh, A.W. Bell and Y.R. Boisclair, 2000. Development of a specific radioimmunoassay to measure physiological changes of circulating leptin in cattle and sheep. *J. Endocrinol.*, 166: 519-528.
- Fruhbeck, G., S.A. Jebb and A.M. Prentice, 1998. Leptin: Physiology and pathophysiology. *J. Clin. Physiol.*, 18: 399-419.

- Gelderman, H., 1997. Investigations on inheritance of quantitative characters in animals by gene markers. *Methods Theor. Applied Genet.*, 46: 319-330.
- Haegeman, A., A. Van Zeveren and L.J. Peelman, 2000. New mutation in exon 2 of bovine leptin gene. *J. Anim. Genet.*, 3: 79.
- Hamann, A. and S. Matthaei, 1996. Regulation of energy balance by leptin. *Exp. Clin. Endocrinol. Diabetes*, 104: 293-300.
- Heravi, M., A.R.M. Ahouei, M. Nassiry and A. Javadmanesh, 2006. Association of leptin polymorphism with production, reproduction and plasma glucose level in Iranian holstein cows. *Asian-Aust. J. Anim. Sci.*, 19: 627.
- Houseknecht, K.L., C.A. Baile, R.L. Matteri and M.E. Spurlock, 1998. The biology of leptin: A review. *J. Anim. Sci.*, 76: 1405-1420.
- Konfortov, B.A., V.E. Licence and J.R. Miller, 1999. Re-sequencing of DNA from a diverse panel of cattle reveals a high level of polymorphism in both intron and exon. *Mamm. Genome*, 10: 1142-1145.
- Lagonigro, R., P. Wiener, F. Pilla, J.A. Woolliams and J. William, 2003. A new mutation in the coding region of the bovine leptin gene associated with feed intake. *J. Anim. Genet.*, 34: 371-374.
- Lande, R., 1981. The minimum number of gene contributing to qualitative variation between and within populations. *Gene*, 99: 541-553.
- Liefers, S.C., R.F. Veerkamp and T.V. Lene, 2002. Association between leptin gene polymorphism and production, live weight energy balance, feed intake and fertility in Holstein heifers. *J. Dairy Sci.*, 85: 1633-1638.
- Liefers, S.C., R.F. Veerkamp, M.F.W. Te Pas, C. Delavaud, Y. Chilliard and T. Van der lende, 2003. Leptin concentrations in relation to energy balance, milk yield, intake, live weight and estrus in dairy cows. *J. Dairy Sci.*, 86: 799-807.
- Lien, S., H. Sundvold, H. Klungland and D.I. Vage, 1997. Two novel polymorphisms in the bovine obesity gene (OBS). *J. Anim. Genet.*, 28: 245.
- Madeja, Z., T. Adamowicz, A. Chmurzynska, T. Jankowski and J. Melonek, 2004. Effect of leptin gene polymorphisms on breeding value for milk production traits. *J. Dairy Sci.*, 87: 3925-3927.
- Pomp, D., T. Zou, A.C. Clutter and W. Barendse, 1997. Rapid communication mapping of leptin to bovine chromosome 4 by linkage analysis of a PCR-based polymorphism. *J. Anim. Sci.*, 75: 1427.