

<http://www.pjbs.org>

PJBS

ISSN 1028-8880

**Pakistan
Journal of Biological Sciences**

ANSI*net*

Asian Network for Scientific Information
308 Lasani Town, Sargodha Road, Faisalabad - Pakistan

Evaluation of Genetic Diversity in *Aegilops geniculata* Roth Accessions using Morphological and RAPD Markers

^{1,2}A. Mahjoub, ³Mohamed Salah El Gharbi, ¹K. Mguis,
²Mohamed El Gazzah and ¹N.B. Brahim

¹Laboratory of Botanic of Inrat, Hedi Karray Street, 2049 Ariana, Tunisia

²Genetic and Bio-Resources Unit, University of Sciences of Tunis, Tunis, Tunisia

³Laboratory of Cereals Genetic of Inrat, Hedi Karray Street, 2049 Ariana, Tunisia

Abstract: Thirteen *Aegilops geniculata* Roth (geniculate goat grass) accessions from collection of the North and Central Tunisian (Cap-Bon, Mogodsés, Kroumiry and the Dorsal areas) were used to assess its genetic diversity by morphological and Random Amplified Polymorphic DNA (RAPD) data and to evaluate relationship between morphological and RAPD markers. Nineteen morphological traits were analyzed on all accessions using Principal Analysis Component (PCA) and clusters were constrained based on median joining distances. Nineteen arbitrary universal primers were used for the amplification of random DNA sequences and generated 212 bands ranging from 0.5 to 3 kb with 71.27% polymorphism across the 13 accessions. Both RAPD and morphological data classified accessions in two main groups. Both methods were used to compare how morphological traits and RAPD molecular markers described accessions relationship and showed a high degree of variation among analyzed accessions, indicating an important source of genetic diversity that can be used in future breeding programs. Morphological PCA traits and cluster indicated climatic stage. In fact, they grouped *Ae. geniculata* accessions according to genetic criteria such as earliness and high kernel yield. Comparison of morphological and molecular data using the Mantel test indicated a non significant correlation ($r = -0.268$). Nevertheless, RAPD and selected morphological characters appear as useful and complementary techniques for evaluation of genetic diversity in *Ae. geniculata*.

Key words: *Aegilops geniculata*, correlation, cluster analysis, genetic diversity, morphological traits, RAPD markers

INTRODUCTION

Aegilops geniculata Roth (*Ae. ovata* auct.) is an annual self-fertile plant, allo-tetraploid species ($2n = 4x = 28$) with MU genomes (Van Slageren, 1994), belonging to tribe *Triticeae* Dumort, subtribe *Triticinae* Griseb. It has a wide distribution in Asia and around the Mediterranean Sea region, characterized by a dry summer season with high temperature and high irradiance. Three annual species of *Aegilops* were reported in Tunisia (Cuénod *et al.*, 1954): *Ae. geniculata* Roth, *Ae. triuncialis* L. and *Ae. ventricosa* Taush. *Ae. geniculata* Roth is widely distributed while the last two species are quite rare. Its geographical distribution would indicate a large distribution of climatic regions: cold and humid mountains, hot and dry valley. Habitats of *Ae. geniculata* Roth in the fertile crescent differ widely in the humid areas receiving around 800 mm and the lower arid area

with less than 150 mm (Ben Brahim *et al.*, 2002). *Aegilops*, is equally adapted to areas with altitude ranging from 10 to 900 m.

Several factors are contributed to the disappearance of the native flora, in particular, those of numerous local varieties and species. Consequently, loss of diversity is observed for all cultivated species and especially for gramineous species such as wild wheat and its cultivated derives. Efficient strategy to solve loss of plant diversity consists of exploiting wild germplasm genomes of wheat species, which preserve a good part of their adaptive factor, diseases tolerance and their genetic richness (Zaharieva *et al.*, 2001). The introgression of a particular gene constituted the main objective of the improvement of wheat (Wang *et al.*, 2000; Nevo *et al.*, 2002). Therefore, it is necessary to have a thorough knowledge of the genetic relationship, ecological distribution and identification of desirable genes in the alien species to be

used in breeding wheat varieties (Chan and Sun, 1997; Sun *et al.*, 1999). Traditionally, evaluation of the genetic diversity in wheat has been based on the differences in agro-morphological traits or pedigree information (Sneller *et al.*, 1997; Bernard *et al.*, 1998). However, molecular markers are becoming essential tools in wheat breeding since they offer alternative solutions to many breeding problems resulting from the traditional phenotypic markers that are difficult and/or time-consuming to select by plant breeders (Najimi *et al.*, 2003). They have been especially used for studying the genetic diversity among a number of species of the tribe Triticeae (Chalmers *et al.*, 2001; Bai *et al.*, 2003; Sun *et al.*, 2003). In this context, several authors have already demonstrated the usefulness of RAPD at specific level in the genus *Aegilops* (Zaharieva *et al.*, 2001; Migdadi, 2006).

In order to assess, conserve and update the genetic diversity of the Tunisian *Ae. geniculata* Roth, we initiated a research program. It aims to collect the seeds of local *Ae. geniculata* species from different sites and evaluate their agro-morphological and molecular traits of breeding

interest. Morphological distances clustering and phenologic RAPD clustering were compared to estimate correlation between morphological traits and RAPD marker.

MATERIALS AND METHODS

Plant material and morphological traits: Collecting missions were conducted in North and Central Tunisia (Cap-Bon, Mogodses, Kroumiry and the Dorsal areas) (Fig 1), in spring during 2 years, 2000 and 2001. Thirteen accessions of *Ae. geniculata* have been collected. These wild materials were distributed in a wide range of climatic conditions (Table 1) with various site altitudes. Seeds of each accession were germinated in Jiffy pots placed in a greenhouse in early November 2002. Two weeks after emergence they were transplanted in the field of the National Institute of Agronomic Research of Tunisia (INRAT). The climate is typically Mediterranean, with mild winter and relatively hot summer. Plants were grown in natural conditions (under rain-fed conditions; no

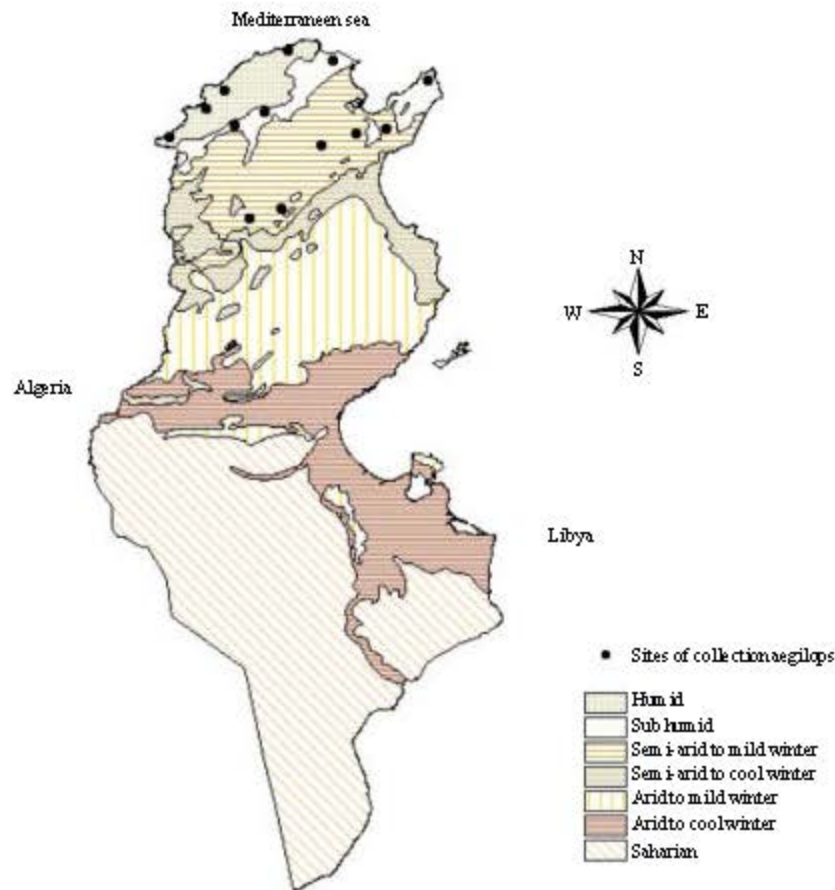


Fig. 1: Distribution sites of *Aegilops geniculata* in Tunisia

Table 1: Characteristics of the origin sites of *Ae. geniculata* accessions in Tunisia

Accession codes	Sites/province	Altitude (m)	Rainfall (mm)	Bioclimatic area
J	Souk jemaâ/Dorsal	870-900	350	Upper semi arid
G	Goussa/North-west	100	351	Sub humid
Abd	Djebel Abderahmen/Cap bon	400	380	Sub humid
B	Bizerte/North-east	300-500	400	Sub humid
Zg	Zaghuan/Dorsal	175	496	Upper semi arid
N	Nefza/Mogodsés	10	720	Sub humid
M	Mekna/Kroumery	500	780	Sub humid
O	Djebel Oues/Dorsal	400	453	Upper semi arid
Z	Ain Zana/Kroumery	641	780	Sub humid
S	Djebel Serj/Dorsal	500	400	Upper arid
Sb	Sbeitla/Center	670	328	Upper arid
T	Tabarka/Kroumery	170	800	Humid
R	Djebel Rexas/North-east	47	453	Upper semi arid

Table 2: Agro-morphological measured traits on *Ae. geniculata*

Abbreviations	Traits
DG	Days to germination
DEM	Days to emergence
DL ₁ T ₁	Days to full expansion of the first leaf
DL ₂ T ₁	Days to the second leaf emergence
DT ₁	Days to the first tiller
T ₁ Le	First tiller length (cm)
NLT ₁	Number of leaves on the first tiller
NT	Total number of tillers
Dfl	Days to flowering
D _{50%} H	Days to 50% of heading
DH	Days to full heading
SpN/E	Spikelet number per ear
E/P	Ears per plant
S/E	Seeds per ear
PHt	Plant height (cm)
ELe	Ear length (cm)
SLe	Seed length (cm)
SN/P	Seeds number per plant
SW/P	Seeds weight per plant (g)

pesticides and no fertilizer were applied; weeds were manually eliminated). Thirteen accessions were planted in completely randomized design with four replicates. Each replication consisted of a 10 plants row. Distance between plants was 70 cm. Rows were 80 cm apart. Heading was noticed 140 days after sowing and the plants were harvested at complete maturity (180 days after sowing). Quantitative characters were evaluated from leaves, ears and seeds from each plant. Morphological measurements of 19 traits were assigned in Table 2. Analysis of Principal Component (PCA), morphological distance clustering and phenologic RAPD markers tree were dealt.

RAPD analysis: The genomic DNAs were extracted from young leaves of *Ae. geniculata* accessions by the cetyltrimethylammonium bromide (CTAB) method with minor modification (Murry and Thompson, 1980). After purification, the DNA concentration was spectrophotometrically estimated. DNA integrity was assessed by 0.8% agarose gel electrophoresis (Sambrook *et al.*, 1989). Twenty nanogram of the extracted genomic DNA was diluted and used for PCR amplifications. Some primers which generate variable

amplification gave poor amplification products or non-repeatable banding patterns were discarded. Only nineteen primers purchased from Operon Technologies inc. (Alameda, USA) that showed numerous bands, were used for the amplification of random DNA sequences.

PCR reactions were performed, in a 25 µL volume reaction mixture containing between 20- 40 ng of total cellular DNA, 5 µL of 5x Taq DNA polymerase buffer, 0.5 µL dNTP (200 µM), 0.2 µ Taq DNA polymerase (5U µL⁻¹), 1 µL de MgCl₂ (2.5 mM) and 25 µM of primer. The reaction mix was overlaid with a drop of mineral water. PCRs were performed using a Biometra UNO II thermal-cycler and involved an initial denaturation step (94°C, 5 min), 40 amplification cycles (each 94°C, 30 sec; 38°C, 1 min and 72°C, 1 min) and a final extension step (72°C, 10 min). Amplification products were analysed by electrophoresis in 2% agarose gels in 1x TAE (Tris Acetate EDTA) buffer and visualized under UV light after Ethidium bromide staining (Sambrook *et al.*, 1989). Amplification was performed and only reproducible products were taken into account for further data analysis.

Data analysis: Principal Component Analysis (PCA) was dealt and the genetic distances between the different accessions were calculated on the centred and standardized variates using measured data with MVSP 3.13 software (Kovach, 1993). To group the populations based on morphological similarity level, cluster analysis was conducted on the Euclidian distance matrix with linkage average method using SAS software. The relationship between the morphological distance matrix and the distances obtained with RAPD markers was analysed. The comparison between the two dendrograms was performed according to the approach developed by Mantel (1967) correlation tests using Mxcomp procedure from NTSYS program (Rohlf, 1993). The principle of this approach is to compare the observed Z-value or r-value with its permutational distribution according a null hypothesis, which is not difference between the distance

matrix, $Z = 0$. In this comparison, 5000 random permutations were made. The null hypothesis of no correlation is rejected when Mantel statistic falls outside the 0.05 confidence level. Legendre and Legendre (1998), state that Mantel's test is only reliable on datasets comprising more than four accessions.

RESULTS

RAPR markers: Analysis of the amplification patterns in *Ae. geniculata* showed a difference by position and number of generated fragments. A total of 212 DNA fragments were generated by 19 primers with an average of 11.15 fragments per primers and were scored as RAPD markers (Table 3). Of these amplified fragments, 153 were polymorphic (about 71.27%), with an average of 11.76 fragments per accession and 8.05 fragments per primers. For each primer, a considerable variation was present for the number of fragments produced (from 9 to 14) and the number of polymorphic fragments (from 3 to 14) that ranged in size from 0.5 to 3 kb. The number and sizes of the DNA fragments were strictly dependent upon primer sequence.

The number of DNA polymorphic fragments per accessions varied from 5.57 in Goussa to 7.26 in Djebel Oust (Table 4). Of the 13 studied accessions, Dj Oust, Tabarka and Ain Zana accession produced the maximum number of DNA polymorphic fragments. This level suggests an efficiency of tested primers. This result showed that the primers OPG02, OPG12, OPM12, OPA06, OPJ06, OPD18, OPG10, OPJ16 and OPA12 were more efficient to assess the genetic relationships,

estimate genetic diversity and to explore the DNA polymorphism of studied genotypes.

Phylogenetic diagram was drawn using the Unweighted Pair Group Method with the Arithmetic Averaging (UPGMA) algorithm and referring to the similarity rate of 70 dendrogram obtained revealed three main clusters (Fig. 2). The first cluster A is formed by two sub-cluster. The first sub-cluster consisted of Dj Ressaas, Ain Zana accessions, the second one is constituted by Bizerte, Dj Oust and Zaghouan accessions, their similarities percentage varied between 68.07 and 73.23%. The second cluster B is formed by two sub-clusters. The first sub-cluster is composed by Dj Serj and Sbeitla accessions, the second is constituted by Souk jemaa,

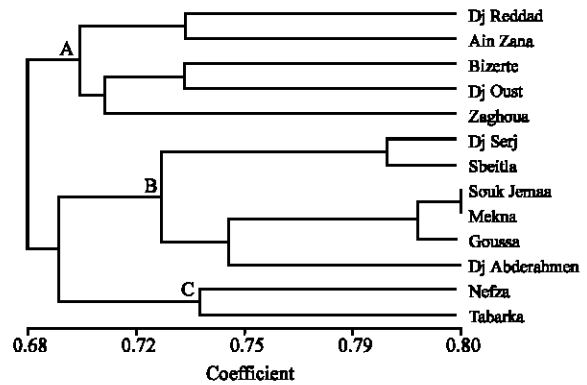


Fig. 2: Dendrogram constructed by UPGMA (weighted pair grouping of arithmetic means) and based on RAPD similarity values showing relationships among the *Aegilops geniculata* Roth

Table 3: Nucleotide sequence of primers with the number of amplified products and percentages of polymorphic fragments in *Ae. geniculata* with respect to the total number of amplification fragments (212) are given

Primers	Sequences (5'-3')	Total amplified fragments	Polymorphic fragments	Polymorphism (%)
OPM14	AGGGTCGTTTC	9	3	33.33
OPB13	TTCCCCGCT	12	4	33.33
OPJ18	TGGTCGCAGA	11	7	63.63
OPG02	GGCATGAGC	13	12	92.30
OPF10	GGAAGCTTGG	12	10	83.33
OPD10	GGTCTACACC	9	4	44.44
OPG10	AGGGCCGTCT	11	10	90.90
OPG12	CAGCTCACGA	14	14	100.00
OPM12	GGGACGTTGG	9	9	100.00
OPA06	GGTCCCTGAC	14	14	100.00
OPJ16	CTGCTTAGGG	11	9	81.81
OPJ06	TCGTTCCGCA	9	8	88.88
OPM16	GTAACAGCC	11	3	27.27
OPD20	ACCCGGTCAC	11	6	54.54
OPD18	GAGAGCCAAC	9	7	77.77
OPE14	TGCGGCTGAG	14	9	64.28
OPA12	TCGGCGATAG	9	7	77.77
OPB05	TGCGCCCTTC	13	10	76.92
OPJ04	CCGAACACGG	11	7	63.63
Total		212	153	71.27

Table 4: Number of DNA polymorphic fragments per accessions

Primers	R	S	Sb	J	Abd	Z	M	G	B	O	N	T	Zg
OPM14	7	4	5	4	6	6	4	5	6	6	4	6	6
OPB13	5	4	4	6	6	5	6	5	6	6	5	6	7
OPJ18	5	5	7	6	7	5	7	7	8	6	9	7	6
OPG02	8	7	5	8	7	8	5	6	5	5	7	9	7
OPF10	6	5	6	6	5	6	5	6	8	8	7	6	7
OPD10	6	3	4	5	6	3	4	4	5	6	7	6	7
OPG10	5	5	5	5	6	5	5	5	5	6	7	8	6
OPG12	6	4	8	6	7	8	4	7	6	7	4	6	7
OPM12	5	5	6	7	4	7	6	5	6	6	3	8	5
OPA06	9	10	9	8	7	6	12	9	11	8	5	5	6
OPJ16	5	3	5	1	5	5	1	2	3	6	2	6	5
OPJ06	4	4	4	1	3	2	0	2	6	2	6	5	6
OPM16	9	8	10	8	10	9	9	8	9	9	8	9	9
OPD20	5	6	7	8	7	8	8	8	8	7	5	9	8
OPD18	5	5	7	5	5	5	4	4	5	6	4	5	5
OPE14	9	8	9	8	5	9	8	6	8	12	6	11	11
OPA12	5	6	5	6	8	6	7	4	7	6	7	6	5
OPB05	8	5	7	8	5	9	7	6	8	12	4	6	10
OPJ04	10	7	7	7	6	10	7	7	9	9	9	9	10
Total	122	104	120	113	115	122	109	106	129	138	109	133	133
Average/primers	6.42	5.47	6.31	5.94	6.05	6.42	5.73	5.57	6.78	7.26	5.73	7	7

Table 5: PCA variable loadings

Morphological traits	PC 1	PC 2	PC 3
DG	0.249	0.008	0.234
DEM	0.212	0.141	0.421
DL ₁ T ₁	0.190	0.058	0.515
DL ₂ T ₁	0.280	-0.102	0.260
DT ₁	0.286	-0.184	0.168
T ₁ Le	-0.300	-0.036	0.156
NLT ₁	-0.198	-0.278	0.053
NT	-0.224	0.260	0.123
Dfl	0.004	0.426	-0.022
D _{50%} H	0.002	0.417	-0.006
DH	0.014	0.420	-0.007
SpN/E	0.216	-0.186	0.125
E/P	-0.239	0.231	0.182
S/E	-0.156	-0.347	-0.046
PHt	-0.313	-0.058	0.102
ELe	-0.265	-0.094	0.014
SLe	-0.212	-0.175	0.419
SN/P	-0.298	0.014	0.210
SW/P	-0.296	0.042	0.287

Mekna, Goussa and Dj Abderhamen. The estimated similarity coefficients ranged from 75.58 and 79.81%. A high similarity at the level of the DNA was showed between two combinations Mekna and Souk jema accessions with 82.16%. The third cluster C is formed by Nefza, Tabarka accessions showing a similarity of 73.70%. *Ae. geniculata* accessions exhibited a low level of intra-specific similarity that varied from 59.15 to 82.16%.

Morphology analysis: Based on eigen values of the order of 1.7 as was suggested by Tomassone *et al.* (1993), the PCA grouped variables into three components, which explained 83.98% of the total variation. The two first axes were considered as they elucidate the maximum simple variation (respectively 48.32 and 26.38% of the total variation) with the cumulative variation of 74.70%.

Loading variables and the PCA scores were also calculated (Table 5). Each principal component was interpreted by its correlation with the original variables. The PC1 accounted for 48.32% of the variation and showed the largest loading values with phenological, morphological and yield-related traits: Days to Germination (DG), days to the second leaf emergence (DL₂T₁), days to the first tiller (DT₁), first tiller length (T₁Le), spikelet number per spike (SpN/E), ear per plant (E/P), Plant height (PHt), ear length (ELe), seeds number per plant (SN/P), seeds weight per plant (SW/P), whereas, the PC2 accounted for 26.38% of the variation shared the largest loading values with phenological, morphological and yield-related traits: Number of leaves on the first tiller (NLT₁), total number of tillers (NT), days to flowering (Dfl), days to 50% of heading (D_{50%}H), days to full heading (DH), Seeds per ear (S/E). Considering the plot defined by the PC1 and PC2 and taking in account their projection on the third plan (PC3), most variables were correlated negatively with the first principal component (PC1) (p<0.01). A clear separation of *Ae. geniculata* accessions was observed and five main groups can be distinguished (Fig. 3).

The first group positively correlated to the two axes and gathered Ain Zana and Mekna accessions. They are originated from the sub humid area (mild and cool winter) at an altitude of 600 m. Moreover, Dj Abderhamen, Goussa, Tabarka and Dj Rerras accessions, are originated from humid and sub humid microclimate, formed the second group. This set is positively correlated to the PC 2 and negatively correlated to the PC 1. The third group is composed by Nefza accessions (from the sub humid area) and Dj Serj accessions (from the upper arid area). They are positively correlated to the PC 2 and negatively correlated

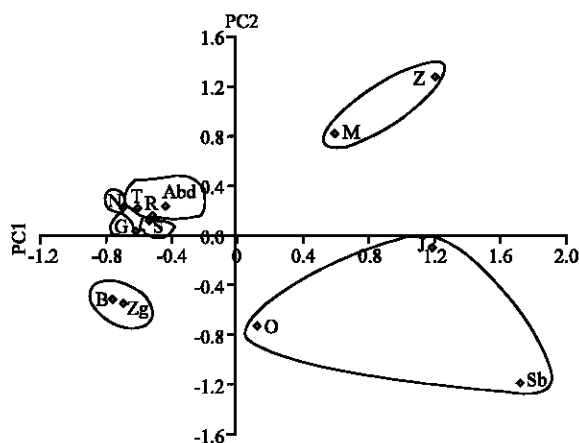


Fig. 3: Principal component analysis of *Aegilops geniculata* accessions

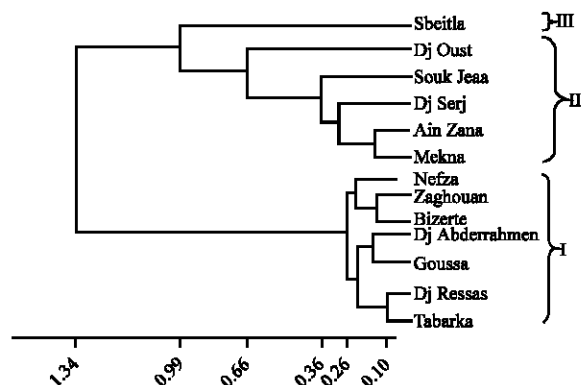


Fig. 4: Dendrogram of *Aegilops geniculata* accessions clustered with average linkage method

to the PC1. Bizerte and Zaghuan accessions are correlated negatively to the two axes and formed the fourth group. They are also belonging to the sub humid area. The fifth group is composed of Sbeitla, Souk jema and Dj Oust accessions which are negatively correlated to PC2 and positively correlated to PC1. They are collected from high altitudes (>400 m) of the upper semi arid with mild winter.

We used the Euclidian distance matrix with linkage average method to draw morphological distances. The dendrogram was constructed as indicated in Fig. 4. It shows three main Clusters (I, II and III) according to climatic regions. In fact, the first cluster contains the greatest number of accessions. It is composed by two sub-clusters (Tabarka, Dj Ressay, Goussa and Dj Abderrahmen) accessions and (Bizerte, Zaghuan and Nefza) accessions originated from humid and sub humid microclimate. The average distance between accessions of the first sub-cluster varies between 0.10 and 0.16; but that

of the second sub-cluster was ranged between 0.14 and 0.23. Cluster II is constituted by five accessions. Mekna and Ain Zana accessions originated from the sub humid area (mild and cool winter) at an altitude of 600 m, formed a sub-cluster. They are grouped at the distance of 0.15. Dj Serj, Souk jema and Dj Oust accessions joined the sub-cluster respectively at the distance of 0.29; 0.36; 0.66. They are collected from high altitudes (>400 m) of the upper semi arid with mild winter. Finally, cluster III represented by Sbeitla accession originated from the upper arid area and growing on an altitude of 670 m, is distinguished by a distance of 0.93.

Despite that the PCA of accessions showed five groups and the dendrogram clustering gave three groups, the most accessions gathered in the same group by PCA are also assembled in the same cluster. In fact, Ain Zana and Mekna accessions belong to the same group according to the two methods. The same remark is given for Zaghuan and Bizerte accessions. Also, we remark that Souk Jema and Dj Oust accessions are included in the same group. The 2 methods also showed Dj Abderrahmen, Goussa, Tabarka and Dj Ressay accessions are associated in the same group.

Comparison between RAPD and morphology: To provide an objective comparison matrices, generated from RAPD and morphological data, were compared using Mentel test. Not significant and quite low correlation between the dendrograms was obtained ($r = -0.268$, $p = 0.0198$) with MxComp procedure from NTSYS programs. In fact, calculated distances between accessions by both methods are different.

DISCUSSION

The estimation of genetic similarities between genotypes gives useful information to address breeding program and germoplasm resource management (Roldan-Ruiz *et al.*, 2001). In this study, morphological data analyses of 13 accessions of *Ae. geniculata* were coupled to molecular analysis (RAPD) to investigate the genetic diversity of *Ae. geniculata*. There has been an interest in differentiating between these accessions and there was low and non significant correlation between RAPD and morphological data.

Agro-morphological diversity among the *Aegilops* populations is further substantiated by principal component analysis, which indicated that the total variation was fairly distributed across all traits. Information obtained throughout principal component analysis may assist plant breeders to identify the number of highly differentiated population for use in crossing and

selection programs (Louati-Namouchi *et al.*, 2000). The PCA of agro-morphological traits classify populations according to climate regions. Several authors have shown that the geographic origin of the collected material was sufficient to obtain a reasonable grouping structure (Julier *et al.*, 1995).

The first group is constructed by Ain Zana and Mekna accessions and originated from the sub humid area with mild and cool winter and located at an altitude of 600 m. They are defined by a late heading and flowering (an average of 180 days), a late germination, a weak kernel yield and high biomass production. Jaradat and Humeid (1990) also observed that genotypes from cooler sites and located at high elevations (900 m altitude) were characterized by late heading and longer filling periods. Moreover, Dj Abderhamen, Goussa, Tabarka and Dj Ressay accessions, formed the second group, originated from humid and sub humid microclimate. Tabarka accession collected from humid microclimate high land providing a sufficient argument for its belonging to this group. The presence in this second environmental pool of the accession Dj Ressay collected from upper semi arid climatic suggested that this accession is situated in the limit of the sub humid area with an annual rainfall of 453 mm. The last group is characterized by early germination, high grain yield and biomass production. Seed yield as well as biomass should be considered when evaluating wild accessions. So, plants of Dj Abderhamen, Goussa, Tabarka and Dj Ressay, exhibited high kernel yield (an average of 660 seeds per plant). In this context, Zaharieva *et al.* (2001) reported an average seed number of 1400 in *Ae. geniculata* plants grown under Mediterranean conditions. She concludes that *Aegilops* plants produce a considerably greater number of spikes and seeds than wheat. The third group is composed by two accessions (Nefza from the sub humid area and Dj Serj belonging to the upper arid area). In fact, these accessions were found in the same group because they belong to regions with rainfall >400 mm. They exhibited early germination, high kernel yield, late heading and flowering, high biomass production.

Bizerte and Zaghuan accessions formed the fourth group. They are also belonging to the sub humid area, characterized by weak biomass production and high kernel yield. They exhibited early germination, heading and flowering (flowering after 160 days). Van Slageren (1994) reported that the flowering time of *Aegilops* in Europe is from April-May until June-July, depending on the species and their ecogeographical location. This trait enables wild species to escape environmental stress during flowering and ensure seed production. Bizerte, Zaghuan and Dj Abderhamen, Goussa, Tabarka and Dj

Ressay genotypes exhibited high plant height (PHt), high number of leaves on the first tiller (NLT₁) and high Seed length (SLe). The last character could be useful in selecting populations with high seedling viability as seed size is directly correlated with the vigour of the seedling (Bullita *et al.*, 1994).

The fifth group is composed of Sbeitla, Souk jemaâ and Dj Oust accessions, collected from upper semi arid with mild winter and high altitudes (>400 m). They are defined by late germination, early heading and flowering and weak kernel yield and biomass production. These populations could be selected for their earliness. Dib Ali *et al.* (1992) reported that in Mediterranean conditions, early heading could represent an important trait favouring plant survival and reproduction under drought and heat stress in the end of the life cycle on seeds per ear (S/E). This result was also mentioned by Chibani (1991) on barley (*Hordeum vulgare* L.). Zaharieva *et al.* (2003) noticed that earliness was the trait with highest contribution of the inter-population variation to the total variation. In fact, elongation and development events are the most sensitive to climatic factors variation (Regan *et al.*, 1992) and strong effect of climatic factors on development rate was noted in several crops (Whan *et al.*, 1991). Concerning the variability of wild Jordanian emmer, Jaradat and Humeid (1990) found that accessions from a dry location are characterized by their high number of spikelets and high productive tillering capacity value. These accessions might harbour genes for drought tolerance and possibility to higher photosynthetic ability. In addition, the restricted geographical area of accessions, with a slight climatic difference between the sub humid and the semi arid area, could explain the proximity of most accessions. Furthermore, accessions originating from sub humid area have higher vigour compared to those originating from dry environment.

The use of RAPD for identification of cultivars through DNA profiling is the current method of choice in measuring genetic variation within germplasm collections (Hernandez *et al.*, 1999). Consequently, the performance of RAPD markers was evaluated using various parameters such as percentage of polymorphism and clusters formed in the dendrogram. In this study, the high level of polymorphism (71.27%) observed in *Ae. geniculata* accessions was obtained by Tao *et al.* (1993) in *Sorghum bicolor* (L.) Moench. Moreover, this result was also reported by Zaharieva *et al.* (2001). The high level of polymorphism was probably due to the large number of primer used in this experiment. Guadagnuolo *et al.* (2001) affirmed that RAPD amplifications provided a largest set of polymorphic markers. Also, the UPGMA dendrogram for RAPD data showed that the structuration group is not

depending on the geographical origin. This result is not in agreement with that found using PCA accession loading and the dendrogram of morphological traits. In this study, representative dendrograms for the accessions based on RAPD markers and morphological traits showed that the overall correspondence between the similarity matrices is low and the correlation between these two dendrograms is not significant ($r = -0.268$). This result is in agreement with those of Roldan-Ruiz *et al.* (2001) working with 16 ryegrass varieties and reporting very low correlation ($r = -0.06$) between AFLP and morphological characters. Similar result was found by Campos *et al.* (2005) in 63 Mandarin (*Citrus* sp.) cultivars, noticing a non significant correlation ($r = 0.31$) between morphological and molecular data. Moreover, our result corroborated with several studies such as European barley varieties (Schut *et al.*, 1997), synthetic hexaploid wheats and their parents (Lage *et al.*, 2003) and Squash germplasm (Ferriol *et al.*, 2004). Two reasons have been mentioned by Semagn (2002) for this low relationship, molecular markers cover a larger proportion of the genome, including coding and noncoding regions, that the morphology and molecular markers are not subjected to artificial selection compared to morphology. For these reasons, RAPD and phenotypic data approaches will not necessarily yield closely matching results. The correlation between them could be improved if there was more morphological markers analyzed or more primer combinations of RAPD were used.

PCA and dendrogram analysis of both morphological traits and RAPD markers showed an important genetic diversity of *Ae. geniculata* accessions. In fact, they were distributed in different groups belonging to different bioclimatic location and on some genetic characteristics as earliness and high kernel yield.

CONCLUSION

Any one of these methods could be used to study diversity and group genotypes, but none would be fully interchangeable in use. The choice of genetic diversity estimate will depend largely upon the tools available to the researcher and how they fit into the breeding scheme. Therefore, phenotypic traits are relatively less reliable and inefficient for precise discrimination of closely related genotypes and analysis of their genetic similarities. However, phenotypic traits, are useful for preliminary, fast, simple and inexpensive varieties identification and can be used as a general approach for assessing genetic diversity among phenotypically distinguishable cultivars, although they are inefficient on account of the time and cost involved (Martinez *et al.*, 2005). The utility of

combining genetic (RAPD) and morphologic characteristics reveals combinations of variation among the *Ae. geniculata* accessions that would not be apparent with any single measurement and could provide a more complete understanding of the germplasm collections diversity.

REFERENCES

- Bai, G., P. Guo and F.L. Kolb, 2003. Genetic relationships among head blight resistant cultivars of wheat assessed on the basis of molecular markers. *Crop Sci.*, 43: 498-507.
- Ben Brahim, N., H. Sebei and M.S. Gharbi, 2002. Collecte des Populations *D'Aegilops ovata* Auct. et évaluation de la diversité génétique pour l'amélioration du blé. 13th Edn., Journées Biologiques, Hammamet.
- Bernard, R.L., C.R. Cremeens, R.L. Cooper, F.L. Collins and O.A. Krober *et al.*, 1998. Evaluation of the USDA Soybean Germplasm Collection: Maturity Groups 000 to IV (FC01.547-PI266.807). Technical Bulletin-United States, Department of Agriculture, USA.
- Bullita, S., R. Floris, M.D. Hayward, A. Loi, C. Porqueddu and F. Veronesi, 1994. Morphological and biochemical variation in Sardinian populations of *M. polymorpha* L. suitable for rainfed mediterranean conditions. *Euphytica*, 77: 263-268.
- Campos, E.T., M.A.G. Spinosa, M.L. Warburton, A.S. Varella and A.V. Monter, 2005. Characterization of Mandarin using Morphological and AFLP Marker. Vol. 30, Interclencia, Venezuela, pp: 687-693.
- Chalmers, J.K., A.W. Campbell, J. Krestschmer, A. Karakousis and P.H. Henschke *et al.*, 2001. Construction of three linkage maps in bread wheat, *Triticum aestivum* L. *Aust. J. Agri. Res.*, 52: 1089-1119.
- Chan, K.F. and M. Sun, 1997. Genetic diversity and relationships detected by isozyme and RAPD analysis of crop and wild species of *Amaranthus*. *Theor. Applied Genet.*, 95: 865-873.
- Chibani, F., 1991. Analyse de la Variabilité Morphologique et Biochimique des Cultivas d'orge (*Hordeum vulgare* L.) Prospectes en Tunisie. Doctorat, Fac. Soc., Tunis.
- Cuénod, A., 1954. Flore analytique et synoptique de la Tunisie Cryptogames vasculaires, Gymnospermes et monocotylédones. *Tunis Imp S, E, F, A, N.* pp: 287.
- Dib Ali, T., P. Monneveux and J.L. Araus, 1992. Adaptation à la sécheresse et notion d'écotype chez le blé dur. II. Caractères physiologiques d'adaptation. *Agronomie*, 12: 371-379.

- Ferriol, M., B. Picó, C.P. de Fernandez and F. Nuez, 2004. Molecular diversity of germplasm collection of squash (*Cucurbita moschata*) determined by SRAP and AFLP markers. *Crop Sci.*, 44: 653-664.
- Guadagnuolo, R., D.S. Bianchi and F. Felder, 2001. Specific genetic markers for wheat, spelt and four wild relatives: Comparison of isozymes, RAPDs and wheat microsatellites. *Genome*, 44: 610-621.
- Hernandez, P., A. Martin and G. Dorado, 1999. Development of SCARs by direct sequencing of RAPD products: A practical tool for the introgression and marker assisted selection of wheat. *Mol. Breed.*, 5: 245-253.
- Jaradat, A.A. and B.O. Humeid, 1990. Morphological variation in *Triticum dicoccoides* from Jordan. In: *Wheat Genetic Resources*, Srivastava, J.P. and A.B. Damania (Eds.). John Wiley and Sons, Chichester, UK.
- Julier, B., A. Porcheron, C. Ecalte and P. Guy, 1995. Genetic variability for morphology, growth and forage yield among perennial diploid and tetraploid Lucern population (*Medicago sativa* L.). *Agronomie*, 5: 295-304.
- Kovach, W.L., 1993. MVSP (Multivariate statistical Package). Kovach PLC.
- Lage, J., M.L. Warburton, J. Crossa, B. Skovmand and S.B. Andersen, 2003. Assessment of genetic diversity in synthetic hexaploid wheats and their *Triticum dicoccon* and *Aegilops tauschii* parents using AFLPs and agronomic traits. *Euphytica*, 134: 305-317.
- Legendre, P. and L. Legendre, 1998. *Numerical Ecology*. 2nd Edn., Elsevier Science BV., Amsterdam.
- Louati-Namouchi, I., M. Louati and A. Chriki, 2000. A quantitative study of some agronomic characters in Sulla (*Hedysarum coronarium* L.). *Agronomie*, 20: 223-231.
- Mantel, N.A., 1967. The detection of disease clustering and a generalized regression approach. *Cancer Res.*, 27: 209-220.
- Martinez, L., P. Cavagnaro and R. Masuelli, 2005. Evaluation of diversity among argentine grapevine (*Vitis vinifera* L.) varieties using morphological data and AFLP markers. *Elect. J. Biotechnol.*, 6: 37-45.
- Migdadi, H.M., A.M. Tell and S. Masoud, 2006. Genetic diversity in some *Aegilops* species in Jordan revealed using RAPD. *PGR Newslett.*, 139: 47-52.
- Murray, M.G. and W.F. Thompson, 1980. Rapid isolation of high molecular weight DNA. *Nucleic Acids Res.*, 8: 4321-4325.
- Najimi, B., S. El Jaafari, M. Jlibène and J.M. Jacquemin, 2003. Applications des marqueurs moléculaires dans l'amélioration du blé tendre pour la résistance aux maladies et aux insectes. *Biotechnol. Agron. Soc. Environ.*, 7: 17-35.
- Nevo, E., A.B. Korol, A. Beiles and T. Fahima, 2002. Evolution of Wild Emmer and Wheat Improvement. *Population Genetics, Genetic Resources and Genome Organization of Wheats Progenitor, Triticum dicoccoides*. Springer, Heidelberg.
- Regan, K.L., K.H.M. Siddique, N.C. Turner and B.R. Whan, 1992. Potential for increasing early vigour and total biomass in spring wheat. II. Characteristics associated with early vigour. *Aust. J. Agric. Res.*, 43: 541-553.
- Rohlf, F.J., 1993. NTSYS-pc: Numerical Taxonomy and Multivariate Analysis System. Ver. 2.20, Exeter Software, New York, ISBN: 0-925031-30-5, pp: 1-38.
- Roldan-Ruiz, I., F.A. van Eeuwijk, T.J. Gilliland, P. Dubreuil and C. Dillmann *et al.*, 2001. A comparative study of molecular and morphological methods of describing relationships between perennial ryegrass (*Lolium perenne* L.) varieties. *Theor. Applied Genet.*, 103: 1138-1150.
- Sambrook, J., E.F. Fritish and T. Maniatis, 1989. *Molecular Cloning A Laboratory Manual*. 2nd Edn., Clod Spring Harbor Laboratory, New York.
- Schut, J.W., X. Qi and P. Stam, 1997. Association between relationships measures based on AFLP markers, pedigree data and morphological traits in barley. *Theor. Applied Genet.*, 95: 1161-1168.
- Semagn, K., 2002. Genetic relationships among ten endod types as revealed by a combination of morphological RAPD and AFLP markers. *Hereditas*, 137: 149-156.
- Sneller, C.H., J.W. Miles and J.M. Hoyt, 1997. Agronomic performance of soybean plant introductions and their genetic similarity to elite genotypes. *Crop Sci.*, 37: 1595-1600.
- Sun, G.L., O. Diaz, B. Salomon and R. VonBothmer, 1999. Genetic diversity in *Elymus caninus* as revealed by isozyme, RAPD and microsatellite markers. *Genome*, 42: 420-431.
- Sun, G., M. Bond, H. Nass, R. Martin and Z. Dong, 2003. RAPD polymorphisms in spring wheat cultivars and lines with different level of *Fusarium* resistance. *Theor. Applied Genet.*, 106: 1059-1067.
- Tao, Y., M.M. Manners, M.M. Ludlow and R.F. Henzell, 1993. DNA polymorphism in grain sorghum (*Sorghum bicolor* L. Moench). *Theor. Applied Genet.*, 86: 679-688.

- Thomassone, R., C. Dervin and J.P. Masson, 1993. Biometrie: Modelisation des Phenomenes Biologiques. Masson, Paris, France, ISBN: 2-225-84074-1, pp: 553.
- Van Slageren, M.W., 1994. Wild Wheat: A Monograph of *Aegilops* L. and *Amblyopyrum* (Jaub and Spash) Eig (Poaceae). Agricultural University, Aleppo.
- Wang, Z.N., A. Hang, J. Hansen, C. Burton, C.A. Mallory-Smith and R.S. Zemetra, 2000. Visualization of A- and B-genome chromosomes in wheat (*Triticum aestivum* L.) x jointed goatgrass (*Aegilops cylindrical* Host) backcross progenies. *Genome*, 43: 1038-1044.
- Whan, B.R., W.K. Anderson, R.F. Gilmour, K.L. Regan and N.C. Turner, 1991. A Role For Physiology in Breeding for Improved Wheat Yield Under Drought Stress. In: *Physiology Breeding of Winter Cereals for Stressed Mediterranean Environments*, Acevedo, E., A.P. Conesa, P. Monneveux and J.P. Srivastava (Eds.). I.N.R.A, les Colloques, Montpellier, France, pp: 179-191.
- Zaharieva, M., E. Gaulin, M. Havaux, E. Acevedo and P. Monneveux, 2001. Drought and heat responses in the wild wheat relative *Aegilops geniculata* Roth: Potential interest for wheat improvement. *Crop Sci.*, 41: 1321-1329.
- Zaharieva, M., A. Dimov, P. Stankova, J. David and P. Monneveux, 2003. Morphological diversity and potential interest for wheat improvement of three *Aegilops* L. species from Bulgaria. *Genet. Resour. Crop Evol.*, 50: 507-517.