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The Histopathalogical Effects of Retinoic Acid on the Tissues

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Abstract: The aim of this study is to sum up the important information that has emerged from the last 10 years of experimental investigations over the effects of retinoic acid (RA) on embryonic structure and adult tissues. Administration of exogenous RA can affect the connective tissues including enhancement of myeloid compartment and suppression of erythroid cells and conversion of hematopoietic stem cells to erythroid progenitors. Also, it is able to induce osteogenic differentiation of stem cells derived from adipose tissues and etc. Examining the neural tissue highlighted that disruption of RA signaling in the adult leads to degeneration of motor neurons and development of some diseases. In vitro administration of All-Trans Retinoic Acid (ATRA) increased dendritic growth and synaptophysin puncta intensity and increased expressions of neuronal nuclei, neuron specific enolase, synaptophysin. RA also promotes expression of a marker of mature astrocytes. On muscular tissue, it can inhibit proliferation of smooth muscle cells (SMC) while promoting differentiation of SMC in vitro instead. The ATRA stimulates skeletal myogenesis while inhibiting cardiomyogenesis and hypertrophy and proliferation of cultured neonatal cardiomyocytes and cardiofibroblasts. In addition, differences in levels of embryonic RA may contribute to variability in great artery anomalies. In epithelial tissue, the squamous epithelium exposed to ATRA showed the columnar differentiation independent to proliferation. Also RA seems able to rescue the regeneration process of injured gut and revealing a better wound healing of the intestine undergone intra-operative radiotherapy. It can interrupt the process of progressive fibrosis, enhancements of the langerhans islets, exocrine pancreas, modulate the health of the mammary glands and repairs the lung cell. Thus, differences in levels of endogenous RA in embryonic and adult tissues may contribute to anomalies and pathogenesis of disease, furthermore RA has paradoxical effects on the parts forming the connective and muscles tissue in equal conditions.

Key words: Histopathalogical, retinoic acid, tissues, embryo, effects

INTRODUCTION

Animal tissues can be grouped into four basic types: connective, muscle, nervous and epithelial. Multiple tissue types comprise organs and body structures. While all animals can generally be considered to contain the four tissue types, the manifestation of these tissues can differ depending on the type of organism.

The normal development of most embryonic structures depends mainly on an intricate but highly orchestrated interplay between spatiotemporally and differentially expressed activators and their antagonistic molecules (Wang et al., 2004; Dean et al., 2009). Among the activators that support the normal development of most embryonic structures, retinoic acid (RA) is well

known as a very influential molecules that is interrelated in many developmental (Hoffman et al., Manolescu et al., 2010) and multiple biological processes. It is involved in regulation of growth, remodeling and metabolic responses in adult tissues (Nov. 2010) through regulation of a wide variety of pathways in cellular differentiation and homeostasis in various tissues and organs, such as head and teeth, by inducing a series of homeobox genes (Maden, 2006). Retinoic acid is essential for lymph node formation (Van de Payert et al., 2009). In addition, RA can also suppress carcinoma cell growth and is currently used in treatment of some cancers. Growth inhibition by RA may be exerted by induction of differentiation, cell cycle arrest, or apoptosis, or by a combination of these activities. Paradoxically, in the

context of some cells, RA not only fails to inhibit growth but, instead, enhances proliferation and survival (Noy, 2010). Unnaturally high quantities of RA, (hypervitaminosis A), as well as its insufficiency (hypovitaminosis A, which in fact is a worldwide problem of great magnitude) during embryonic development cause a great spectrum of congenital malformations (Nobakht *et al.*, 2006) and as well as in pathological consequences for adult (Penniston and Tanumihardjo, 2006).

However, in the light of specific experimental attempts, roles of the RA revealed to be expanded and whereas, most of previous reviews focus on anti-neoplastic effects or only on a special tissue, then a need to an overall and new review on the basis of the other recent studies was seriously felt.

Connective tissue and RA

Blood (Myeloid): Hematopoietic development during embryogenesis involves the interaction of extrinsic signaling pathways coupled to an intrinsic cell fate that is regulated by cell-specific transcription factors. The RA has been linked to stem cell self-renewal in adults and also participates in yolk sac blood island formation (De Jong *et al.*, 2010).

There are strong evidences that vitamin A (Kang et al., 2007) and the retinoids have immune regulatory functions (Kang et al., 2007; Chen et al., 2009) and improve the numbers of various immune cell types such as T helper- 2 (Iwata et al., 2003) and neutrophil (Yousefi and Samene, 2010). Exogenous all-trans retinoic acid (ATRA) enhances the proliferation and differentiation of myeloid compartment specially neutrophil during both late-yolk sac and fetal liver stages of hematopoiesis in rat embryo (Yousefi and Samene, 2010) and the patients who were treated with the ATRA (Leelasiri et al., 2005). Also, the findings of previous studies show that the ATRA enhances the hematopoietic progenitor cells in vitro (Collins et al., 2001; Yu et al., 2010) and directly enhances the number of T lymphocytes in the peripheral blood (Seguin-Devaux et al., 2005). A recent study shows that addition of the ATRA greatly enhances the generation of hematopoietic progenitors from human embryonic stem cells and the RA-treated cells highly express definitive hematopoietic genes that form large numbers of multilineage and myeloid colonies (Yu et al., 2010). The RA plays a key role in myeloid differentiation through their agonistic nuclear receptors (RAR alpha/RXR) to modulate the expression of target genes (Zhang et al., 2000). This receptor can bi-directionally modulate granulopoiesis

differentiation factor when liganded to the RA or as an inhibitor in absence of ligand (Kastner *et al.*, 2001).

Blood (Erythroid): Many in vitro studies have analyzed the effects of retinoic acid on hematopoietic progenitors and differentiation and in some cases, showed a positive role for clonal proliferation of progenitors (Ghatpande et al., 2002; Yu et al., 2010). In contrast, considerable evidence from other in vivo and in vitro studies revealed the suppressive effects of RA on proliferation and differentiation of human mast cell progenitors (Kinoshita et al., 2000), growth of bone marrow mesenchymal stem cells (Oliva et al., 2003), erythroid cell (Yu et al., 2010; Yousefi, 2009) and platelets in mouse embryo (Yousefi, 2009). Decreased numbers of both red blood cells and platelets could be that probably the primitive erythroid and megakaryocyte cells origin from previously unrecognized bipotential megakaryocyte/ primitive erythroid progenitors that both emerge from hemangioblast precursors during gastrulation. The megakaryocyte lineage is closely associated with erythroid lineages throughout ontogeny (Tober et al., 2007). The study of Kinoshita et al. (2000), indicated that the RA diminished the mean diameter of culture of mast cells, but our data shows that the RA dose not have any effect on the blood cell size (except NRBC) (Yousefi and Samene, 2010; Yousefi, 2009). The cyto-differentiating action of ATRA has led to its usage in the treatment of several malignancies, particularly acute promyelocytic leukemia (APL). There have been many reports regarding the cell biological effects of ATRA on human myeloma cells and a few clinical trials. Most of these reports have revealed growth inhibition by ATRA mediated by down-regulation of the IL-6/IL-6R auto/paracrine loop and upregulation of p21/Cip1 (Otsuki et al., 2003). However, it is concluded that this agent enhances the proliferation and differentiation of myeloid compartment while, inhibites erythroid cells, although the reported has shown that the ATRA were interfered with proliferation and differentiation of hematopoietic stem cells to the erythroid progenitors with 6×10⁻⁸ mol L⁻¹ in vitro and up-regulate the expression of hoxb2 and hoxb4 significantly (Du et al., 2009).

Bone and cartilage: The multi-step process of cartilage tissue formation is comprised of prechondrogenic mesenchymal cells compaction prior to differentiation into matrix-producing chondroblasts (Hoffman *et al.*, 2003). Also bone formation and remodeling require a meticulous orchestration of a number of signaling components (Ducy *et al.*, 2000; Teitelbaum, 2000).

The retinoids, predominantly the RA, are among the numerous signaling molecules that have been concerned in this process (Hoffman et al., 2003). The RA has been shown to influence differentiation and proliferation of osteoblasts (Kawaguchi et al., 2005; Skillington et al., 2002; Song et al., 2005) and Chondrocyte (Kirimoto et al., 2005). Recent studies have demonstrated that the RA is able to induce osteogenic differentiation of stem cells derived from adipose tissues (Malladi et al., 2006), dental pulp and periodontal ligament (Chadipiralla et al., 2010) in vitro. It is shown that the ATRA strongly induces collagenase-3 expression in cultures of embryonic metatarsal cartilage rudiments and in chondrocytic cells. This effect is dose and time dependent and requires the renewed synthesis of proteins and is mediated by RAR-RXR heterodimers. The above-mentioned effect acts through a signaling pathway involving p38 mitogen-activated protein kinase (p38MAPK). RA of chondrocytic cells also induces the production of matrix metalloproteinase MT1 and also a membrane-bound metalloproteinase essential for skeletal formation, which participates in a proteolytic cascade with collagenase-3. RA treatment also resulted in upregulation of Cbfa1, a transcription factor responsible for collagenase-3 and osteocalcin induction in osteoblastic cells (Jimenez et al., 2001). Also, it is reported that the RA can suppress the cell growth, cartilage nodule formation, accumulation of proteoglycan, alkaline phosphatase activity and mineralization while RA dose dependently upregulates the levels of type X collagen and matrix metalloproteinase-13 (MMP-13) mRNA which are marker proteins of hypertrophic chondrocytes Chondroprogenitor (ATDC5) cells in culture instead (Kirimoto et al., 2005).

Another study was performed to examine the influence of retinoid signaling on skeletal development, the primary limb mesenchymal cultures from the transgenic mice that overexpressed a weak, constitutively active retinoic acid receptor (retinoic acid receptor-α) in their developing limbs were compared with cultures from wild-type mice. The finding indicates that retinoids maintain cells within condensations in a prechondrogenic, mesenchymal cell state, which prevents the cells from differentiating into chondroblasts and the inhibition of receptor-mediated retinoid signaling induces expression of Sox9, a transcription factor that is considered a master switch for the differentiation of chondroblasts. These effects are largely mediated by the activation of the p38 MAPK signaling (Hoffman et al., 2003). So, the RA induces a regulatory cascade involving Cbfa1 and MMPs and is coupled to the development of a perichondrial invasion and osteogenic

differentiation process that occurs during endochondral ossification (Jimenez et al., 2001).

Neural tissue and RA: RA also is involved in the induction of neural differentiation, motor axon outgrowth and neural patterning. Like other developmental molecules, RA continues to play a role after development has been completed (Maden, 2007). It has recently come to light, however, that many of the same functions that RA directs in the embryo are involved in the regulation of neural plasticity, long-term potentiation in the hippocampus, regeneration in the adult brain (Mey, 2006), ATRA can be synthesized in discrete regions of the brain. Distribution of retinoid receptor proteins in the adult nervous system is different from what seen during development and suggests that retinoid signalling is likely to have a physiological role in adult cortex, amygdala, hypothalamus, hippocampus, striatum and associated brain regions (Lane and Bailey, 2005). Disruption of RA signalling in the adult leads to the degeneration of motor neurons (motor neuron disease), development of Alzheimer's disease and possibly, development of Parkinson's disease (Maden, 2007). The experiments on sciatic nerve lesions and spinal cord contusion injury demonstrate that the RA signaling cascade is activated by these traumatic events. Activation of RARbeta appears to be responsible for neurotrophic and neuritogenic effects of RA on dorsal root ganglia and embryonic spinal cord. While the physiological role of RA in the injured nervous system is still under investigation, three domains of functions are suggested: (1) neuroprotection and support of axonal growth, (2) modulation of the inflammatory reaction by microglia/macrophages and (3) regulation of glial differentiation (Mey and McCaffery, 2004). RA levels require precise regulation by controlled synthesis and catabolism and when RA concentrations deviate from normal, in either direction, abnormal growth and development occurs (McCaffery et al., 2003). In rodents, the control of neural patterning and differentiation would be disrupted when RA concentrations are lowered, whereas, inappropriately high concentrations of RA result in abnormal development of cerebellum and hindbrain nuclei. The latter parallels the malformations seen in the human embryo exposed to RA due to treatment of the mother with the acne drug Accutane (13-cis RA) and, in cases when the child survives beyond birth, a particular set of behavioural anomalies can be described (McCaffery et al., 2003). Investigations over the effects of vitamin A deficiency (VAD) on neurogenesis and memory and the ability of RA treatment in preventing VAD-induced impairments show that long-term VAD decreased neurogenesis and led to memory deficits.

More importantly, these effects were reversed to what manifested in 4 weeks of RA treatment (Bonnet *et al.*, 2008).

Bonnet *et al.* (2008) have firstly demonstrated that the effect of vitamin A deficient diet on the level of hippoccampal neurogenesis would be reversible and that RA treatment is important for the maintenance of the hippocampal plasticity and function.

It is shown that activity blockade increases RA synthesis in neurons and that acute RA treatment enhanced synaptic transmission. The RA-induced increase in synaptic strength is occluded by activity blockade-induced synaptic scaling (Aoto et al., 2008). Also a study has shown that within 30 min ATRA increased dendritic growth approximately two folds and PSD-95 synaptophysin and puncta intensity approximately three folds, in cultured mouse hippocampal neurons, suggesting the increased synapse formation (Chen and Napoli, 2008). The differentiation property of RA was confirmed by showing an extensive outgrowth of neurites, increased expressions of neuronal nuclei, neuron specific enolase, synaptophysin and synaptic associated protein-97 and decreased expression of inhibitor of differentiation-1 (Cheung et al., 2009). The ATRA decrease cell viability but not neurite outgrowth (Radio et al., 2010). Showing that addition of exogenous ATRA to slice cultures can promote expression of a marker of mature astrocytes, glial fibrillary acidic protein (GFAP), while inhibitioning the endogenous RA synthesis reduces GFAP expression instead. The results suggest a role for retinoic acid in modulating glial differentiation (King et al., 2009).

Muscular tissue and RA: Evidences suggests that ATRA plays a key role in the development and differentiation of smooth muscle cells (SMC) in vitro (Su et al., 2010) and it's very important during normal embryonic development (Ross et al., 2000). The ATRA increased the expression of myocardin, caldesmon, 22 kDa smooth muscle cell-specific protein (SM22alpha) and SM-myosin heavy chains in Rabbit Bone Marrow-Derived Mesenchymal Stem cells (RBMSCs). Furthermore it significantly inhibits proliferation of RBMSCs in a concentration-dependent manner and up-regulates the expression of SMC specific proteins and promotes SMC differentiation (Su et al., 2010). In vitro ATRA treatment induces smooth muscle differentiation of P19 cells (Manabe and Owens, 2001). In addition, ATRA has been shown to inhibit the proliferation SMCs. by inhibiting the expression of cyclins and cyclin-dependent kinases (Kosaka et al., 2001). Also a quantitative morphometry of the pulmonary arteries has shown that ATRA treatment (30 mg kg⁻¹) has

significantly reduced the percentage of muscularized arteries in peripheral pulmonary arteries only with an external diameter between 15 and 50 μ m and also significantly reduced the medial wall thickness in small muscular arteries only with an external diameter between 50 and 100 μ m (Zhang *et al.*, 2010). As obtained conclusions, the ATRA showed to inhibit proliferation while increasing differentiation instead. By the way, types of ATRA behaviors under different doses still remain obscure and need more researches.

Recently, Kennedy et al. (2009) have reported that the low levels of RA stimulate skeletal myogenesis by accelerating and increasing the expression of Wnt3a, Pax3, Meox1 and MRFs. This early and enhanced activation of skeletal muscle is refractory to inhibitory signals from bone morphogenetic protein 4 (BMP4) but not from a dominant negative β-catenin. Furthermore, low levels of RA inhibit stem cell differentiation into the cardiac muscle lineage, as shown by the absence of GATA-4 expression. The inhibitory activity of RA on cardiomyogenesis can be abrogated by the presence of BMP4. Therefore, BMP4 and RA function antagonistically to regulate each other's inhibition of entry into skeletal and cardiac muscle lineages, respectively (Kennedy et al., 2009). In addition, it was demonstrated that embryonic RA synthesis is required for proper differentiation of ventricular myocytes in mice (Niederreither et al., 2001; Subbarayan et al., 2000). The ATRA inhibits hypertrophy and proliferation of cultured neonatal cardiomyocytes and cardiofibroblasts induced by angiotensin II (Wang et al., 2002). Lu et al. (2003), revealed an inhibitory effect of low-dose chronic ATRA treatment which prevented the hypertrophy of intramyocardial and intrarenal arteries and ventricular fibrosis in spontaneously hypertensive rats (SHR). These findings revealed that the ATRA can prevent medial thickening of intramyocardial and intrarenal arteries and perivascular and interstitial fibrosis in the heart without any change of the blood chemistry profile in any group and provides the first evidence for an inhibitory effect of the ATRA on intramyocardial and intrarenal arterial hypertrophy and ventricular fibrosis in vivo (Lu et al., 2003). The ATRA also has an inhibitory effect on the cardiac fibroblast in vitro (Wang et al., 2002). So, the ATRA stimulates skeletal myogenesis while inhibiting cardiomyogenesis and hypertrophy and proliferation of cultured neonatal cardiomyocytes and cardiofibroblasts at lower levels on the other hand the ATRA plays a paradoxical behavior about skeletal muscles compared with cardiac muscles. It is suggested that the differences in levels of embryonic RA may contribute to variability in great artery anomalies observed in DGS/VCFS patients, so that Loss of Tbx1

and decrease of RA synthesis result in DiGeorge/velocardiofacial syndrome (DGS/VCFS)-like phenotypes in mouse models including defects in septation of the outflow tract of the heart and anomalies of pharyngeal arch-derived structures including arteries of the head and neck, laryngeal-tracheal cartilage and thymus/parathyroid. Recent studies have shown that reduction of RA or loss of Tbx1 can alter the contribution of second heart field progenitor cells to the elongating heart tube (Ryckebusch *et al.*, 2010).

Epithelial tissue and RA: Some epithelial cells cover the outside of organism; these often form multiple layers, such as the skin. Other epithelial cells form monolayers that line internal organs (Zegers et al., 2003), while others form internal three-dimensional organs consisting of tubules and cysts (Affolter et al., 2003) that infiltrate the whole organism, carrying liquids and gases containing nutrients, waste and other materials. These tubes can form elaborate networks in the lung, kidney, reproductive passages and vasculature tree, as well as many glands branching from the digestive system such as the liver, pancreas and salivary glands (Zegers et al., 2003).

Retinoids are essential for epithelial tissue growth and differentiation (Rexer et al., 2001) and regulate these functions through activation of specific nuclear RA receptors (RARs) (Stoppie et al., 2000).

Oral administration of R115866 (2.5 mg kg⁻¹) to rats induced a marked and transient increases of endogenous RA levels in the plasma, skin, fat, kidney and testis consistent with its ability of enhancing endogenous RA content in tissues, so it was found to exert retinoidal activities (Stoppie *et al.*, 2000; Thomas *et al.*, 2005) furthermore, it is shown that R115866 can also potentiate the effects of RA on epidermal keratinocytes when RA appears at low concentrations (Giltaire *et al.*, 2009). It is also manifested that RA homeostasis could be controlled and stabilized by cytochrome P26-induced oxidation (Verfaille *et al.*, 2008).

In Barrett's oesophagus, the level of RA biosynthesis was increased and morphological and molecular analysis of the squamous epithelium exposed to ATRA showed the columnar differentiation independent to proliferation (Chang *et al.*, 2007). In HBE1 cells, a respiratory tract cell line, DUOX2 mRNA was increased six folds after ATRA treatment. Similarly, the ATRA induced a 19-fold increase in DUOX2 mRNA expression in primary tracheobronchial epithelialcells with parallel increases in DUOX protein and DUOX-mediated H₂O₂ production as Well. The data extracted from the study done with Linderholm *et al.* (2010), affirmed that the ATRA is important to regulate DUOX2 expression, function and

rhinovirus-mediated DUOX2 induction in the respiratory epithelial cells. The results of other studies showed that RA is required for gut regeneration while inhibitors of RA synthesis could suppress formation of the gut and use of 13-cis RA seemed able to rescue the regeneration process from the inhibitor-induced hypoplasia (Kaneko et al., 2010). In order to clarify the reparative effects of pre- and post-treatment of the vitamin A on the jejunum, methotrexate firstly used to make the damages such as villus shortening, fusion, epithelial atrophy, crypt loss, inflammatory infiltrate in the lamina propria and goblet cell depletion of the gut and thereafter administration of vitamin A showed to diminish the severity of the uttered damages as well (Yuncu et al., 2004). Balkan et al. (2006), tried to assess preventive role of the RA over side effects of intra-operative radiotherapy on intestinal wound healing compared to the non-retinoic acid conditioned group in which the RA conditioned group statistically had a significantly higher tensile strength and lower intestinal elongation values, revealing better wound healing.

Natarajan et al. (2009), also performed a study over intestinal epithelium of cirrhotic rats indicating that altered retinoid metabolism might have an influence on changes in intestinal epithelial cell differentiation, seen in liver cirrhosis (Natarajan et al., 2009). Also RA treatment can interrupt some of the steps toward the progressive fibrosis in monocrotaline-induced Lung disease (Baybutt et al., 2007).

Furthermore, in regard to effects of retinoids on the alveolar cells the study showed that a vitamin A deficient diet can lead to a decreased number of alveoli and emphysema in rats as result (Nabeyrat *et al.*, 2001). The RA has also been shown to protect against injury-induced cell-cycle arrest of the type II pneumocytes by preserving cyclin activities and down-regulating the cyclin-dependent kinase inhibitor p21^{cip1}.

There are many expanded reviews done over the anticarcinogenic effects of RA on different cells but we tried to bring those specially indicating such effects on epithelial cells as following:

The KYSE30 cancerous cells were cultured on the nanoporous membranes to examine the effects of RA treatment (0.01 M) the RA showed to inhibit cancerous cell-substrate adhesion and motility and also induce morphological and functional terminal differentiation of the cell line. This experiment testified that the impedance amplitude decrease was really due to the cell morphology change after the treatment of RA (Yu et al., 2009). The RA is an important mediator that directs lung cell repair by stimulation of the type II pneumocyte proliferation in the case of type I pneumocyte injury. In addition, RA inhibits proliferation of the pneumocytes that are in contact with

each other thereby it is protective against abnormal proliferation as in carcinogenesis (Baybutt *et al.*, 2007).

Examination of RA treatment impact (10 µM 9- and 13-cis-retinoic acid) on the intestinal epithelial cells, Caco-2 cells an intestitual cell line derived from a human colorectal carcinoma that spontaneously differentiates under standard culture conditions, a well-characterized marker for functional differentiation of enterocytes) indicated that the RA acts as a differentiation factor since, at 60% preconfluence and at 0-day confluence, it (1) lowered labeled thymidine incorporation; (2) modulated D-type cyclins (cell cycle regulatory proteins) by down-regulating the mitogen-sensitive cyclin D1 expression (thereby blocking DNA synthesis and cell proliferation) and up-regulating cyclin D3 expression (thereby inducing cell differentiation) and (3) showed a trend of increase in (p38 MAPK), which triggers caudal-related homeobox 2 (CDX2), a central protein in cell differentiation (Grenier et al., 2007).

Glands (pancreas) and RA: The pancreas is composed of two different types of glandular tissues in intimate association with each other. The main mass is exocrine, embedded in which are pancreatic islets of endocrine cells or langerhans (Murtaugh and Melton, 2003). Previous studies have revealed that exogenous ATRA inhibits the bud morphogenesis and differentiation of Exocrine pancreas (acinus), while in same culture enhances endocrine cells formation, specially beta cells before normal times in vitro (Shen et al., 2007). Moreover, Colakoglu et al. (2005), have shown that a high dose of RA can decrease the exocrine pancreas (acinus) during pancreatic organogenesis. While our (Yousefi et al., 2010) data indicated that the ATRA has positive effects on proliferation, differentiation and maturation of pancreatic acinus and pancreatic islets of langerhans. Also, the mean diameter of pancreatic acinus and nuclei of acinar cells were decreased in rat's embryo on GD 18, when exposed to the ATRA in utero on GD 10. Colakoglu et al. (2005), found that the ATRA administration on the 8th day of gestation caused inhibition of acinar TGF-b2 immune reactivity while on GD 12, acinar expression of TGF-b2 in acid was similar to what found in the controls. Shen et al. (2007), reported that exocrine differentiation was found to be suppressed at all concentrations of the ATRA compared to controls in vitro. Also it was reported that exogenously added ATRA in tissue culture experiments stimulated differentiation of endocrine and duct cells and promoted apoptotic cell death of acinar tissue and inhibited acinar differentiation (Colakoglu et al., 2005; Tulachan et al., 2003). As result, it is not easy to determine a cause to account for such a high diversity.

Although, it's suggested that the RA can suppress exocrine differentiation by a combination of two mechanisms (1) up-regulation of the extra-cellular matrix component laminin and (2) enhancement of apoptosis. It's also suggested that loss of exocrine cells following application of exogenous RA correlates with an inhibition of Notch signaling activity and can be rescued by FGF-10 (Chen et al., 2004) However, much more studies are needed to clarify this issue. The results extracted from in vivo and in vitro studies have shown that exogenous ATRA causes differentiation of endocrine cell (Yamaguchi and Igarashi, 2006; Kang et al., 2004; Erasmus et al., 2003; Tulachan et al., 2003). Also, it expands Islets of langerhans in the pancreatic organogenesis, when administered by oral intubations in the morning of GD 10 (Yousefi et al., 2010) These observations are to in contrast the results obtained from the study done by Colakoglu et al. (2005), who reported that if it is administered before neurulation, the development of endocrine component will be disrupted, while its administration after neurulation has had less adverse affects on pancreatic organogenesis. The RA also increased insulin-secreting cells by stimulation of extra progenitor cells production (Erasmus et al., 2003). It is reported that pdx1 genes (Li et al., 2004) have had important and main roles to increase proliferation of insulin-secreting cells, which is regulated by the RA in vitro. Recent evidence indicates that the enhancing effect of RA on formation of enlarged islets may be resulted from up-regulation of extra-cellular laminin (Shen et al., 2007). It also inhibits the inflammatory response or on the other hand, modulates the health of mammary glands by suppression of TLR4/NF-xB expression both in vivo and in vitro (Gu et al., 2010). It can be concluded that retinoic acid may help in development of new therapeutic methods to increase the Langerhans islets, exocrine pancreas and modulate the health of the mammary glands in mammals.

CONCLUSION

As this study shows, the endogenous RA plays an important role in normal embryogenesis and adult tissue health and any change over the normal level of RA can develop many types of anomalies and pathogenesis of different tissues. The exogenous RA is probably accepted to compensate the endogenous RA deficit, conclusively treatment by the RA may be beneficial on the normal and cancerous tissues but it's unfortunately associated to adverse effects *in vivo* that many of them have been revealed and some are still obscure that needs much more

studies, thus it's recommended that more tests to be done on the animal models with artificially created favorite diseases. Furthermore, the RA may have some paradoxical effects on the parts forming the connective and muscular tissues in equal conditions and can also be used as a differentiation-proliferation-provoking agent on the epithelial cells especially in the pancreas.

REFERENCES

- Affolter, M., S. Bellusci, N. Itoh, B. Shilo, J.P. Thiery and Z. Werb, 2003. Tube or not tube: Remodeling epithelial tissues by branching morphogenesis. Dev. Cell, 4: 8-11.
- Aoto, J., C.I. Nam, M.M. Poon, P. Ting and L. Chen, 2008. Synaptic signaling by all-trans retinoic acid in homeostatic synaptic plasticity. Neuron, 60: 308-320.
- Balkan, M., M. Beyzadeoglu, K. Oysul, B. Dirican, S. Surenkok and T. Tufan, 2006. Retinoic acid and intestinal wound healing in intra-operatively irradiated rat. Acta Chir. Belg., 106: 73-76.
- Baybutt, R.C., B.L. Herndon, J. Umbehr, J. Mein and Y. Xue et al., 2007. Effects on cytokines and histology by treatment with the ACE inhibitor captopril and the antioxidant retinoic acid in the monocrotaline model of experimentally induced lung fibrosis. Curr. Pharm., 13: 1327-1333.
- Bonnet, E., K. Touyarot, S. Alfos, V. Pallet, P. Higueret and D.N. Abrous, 2008. Retinoic acid restores adult hippocampal neurogenesis and reverses spatial memory deficit in vitamin A deprived rats. PLoS. One, 3: e3487.
- Chadipiralla, K., J.M. Yochim, B. Bahuleyan, C.Y. Huang, F. Garcia-Godoy, P.E. Murray and E.J. Stelnicki, 2010. Osteogenic differentiation of stem cells derived from human periodontal ligaments and pulp of human exfoliated deciduous teeth. Cell Tissue Res., 340: 323-333.
- Chang, C.L., P. Lao-Sirieix, V. Save, G. De La Cueva Mendez, R. Laskey and R.C. Fitzgerald, 2007. Retinoic acid-induced glandular differentiation of the oesophagus. Gut, 56: 906-917.
- Chen, N. and J.L. Napoli, 2008. All-trans-retinoic acid stimulates translation and induces spine formation in hippocampal neurons through a membrane-associated RARalpha. FASEB. J., 22: 236-245.
- Chen, X., R.S. Welner and P.W. Kincade, 2009. A possible contribution of retinoids to regulation of fetal B lymphopoiesis. Eur. J. Immunol., 39: 2515-2524.
- Chen, Y., F.C. Pan, N. Brandes, S. Afelik, M. Solter and T. Pieler, 2004. Retinoic acid signaling is essential for pancreas development and promotes endocrine at the expense of exocrine cell differentiation in Xenopus. Dev. Biol., 271: 144-160.

- Cheung, Y.T., W.K. Lau, M.S. Yu, C.S. Lai and S.C. Yeung et al., 2009. Effects of all-trans-retinoic acid on human SH-SY5Y neuroblastoma as in vitro model in neurotoxicity research. Neurotoxicology, 30: 127-135.
- Colakoglu, N., A. Kukner, J. Oner, M.F. Sonmez, H. Oner and E. Ozan, 2005. Effects of high dose retinoic acid on TGF-beta2 expression during pancreatic organogenesis. J. Mol. Histol., 36: 413-418.
- Collins, S.J., J. Ulmer, L.E. Purton and G. Darlington, 2001. Multipotent hematopoietic cell lines derived from C/EBPalpha(-/-) knockout mice display granulocyte macrophagus-colony-stimulating factor, granulocytecolony-stimulating factor and retinoic acid-induced granulocytic differentiation. Blood, 98: 2382-2388.
- DU, C.Q., M.X. Huang and W.J. Liu, 2009. Effect of ATRA on the expression of genes Hoxb2 and Hoxb4 in cord blood erythroid progenitors. Zhongguo. Shi Yan Xue Ye Xue Za Zhi, 176: 1516-1521.
- De Jong, J.L.., A.J. Davidson, Y. Wang, J. Palis and P. Opara *et al.*, 2010. Interaction of retinoic acid and scl controls primitive blood development. Blood, 116: 201-209.
- Dean, D.B., J.T. Watson, B.R. Moed and Z. Zhang, 2009. Role of bone morphogenetic proteins and their antagonists in healing of bone fracture. Front Biosci., 14: 2878-2888.
- Ducy, P., T. Schinke and G. Karsenty, 2000. The osteoblast: A sophisticated fibroblast under central surveillance. Science, 289: 1501-1504.
- Erasmus, C., C. Penny and B. Kramer, 2003. Retinoic acid increases the length and volume density of ducts in the rat embryonic pancreas. Dev. Growth Differ, 45: 199-207.
- Ghatpande, S., A. Ghatpande, J. Sher, M.H. Zile and T. Evans, 2002. Retinoid signaling regulates primitive (yolk sac) hematopoiesis. Blood, 99: 2379-2386.
- Giltaire, S., F. Herphelin, A. Frankart, M. Herin, P. Stoppie and Y. Poumay, 2009. The CYP26 inhibitor R115866 potentiates the effects of all-trans retinoic acid on cultured human epidermal keratinocytes. Br. J. Dermatol., 160: 505-513.
- Grenier, E., F.S. Maupas, J.F. Beaulieu, E. Seidman and E. Delvin et al., 2007. Effect of retinoic acid on cell proliferation and differentiation as well as on lipid synthesis, lipoprotein secretion and apolipoprotein biogenesis. Am. J. Physiol. Gastrointest. Liver Physiol., 293: G1178-1189.
- Gu, B., J. Miao, Y. Fa, J. Lu and S. Zou, 2010. Retinoic acid attenuates lipopolysaccharide-induced inflammatory responses by suppressing TLR4/NF-kappaB expression in rat mammary tissue. Int. Immunopharmacol., 10: 799-805.

- Hoffman, L.M., A.D. Weston and T.M. Underhill, 2003.
 Molecular mechanisms regulating chondroblast differentiation. J. Bone Joint Surg. Am., 85: 124-132.
- Hoffman, L.M., K. Garcha, K. Karamboulas, M.F. Cowan and L.M. Drysdale *et al.*, 2006. BMP action in skeletogenesis involves attenuation of retinoid signaling. J. Cell Biol., 74: 101-113.
- Iwata, M., Y. Eshima and H. Kagechika, 2003. Retinoic acids exert direct effects on T cells to suppress Th1 development and enhance Th2 development via retinoic acid receptors. Int. Immunol., 15: 1017-1025.
- Jimenez, M.J., M. Balbin, J. Alvarez, T. Komori and P. Bianco et al., 2001. A regulatory cascade involving retinoic acid, Cbfa1, and matrix metalloproteinases is coupled to the development of a process of perichondrial invasion and osteogenic differentiation during bone formation. J. Cell Biol., 155: 1333-1344.
- Kaneko, N., Y. Katsuyama, K. Kawamura and S. Fujiwara, 2010. Regeneration of the gut requires retinoic acid in the budding ascidian Polyandrocarpa misakiensis. Dev. Growth Differ, 52: 457-468.
- Kang, M.K., Y.E. Yoon, J.Y. Yang, K.B. Kwon, J.W. Park and E.C. Jhee, 2004. Protective effect of retinoic acid on interleukin-1 beta-induced cytotoxicity of pancreatic beta-cells. Mech. Ageing. Dev., 125: 483-490.
- Kang, S.G., H.W. Lim, O.M. Andrisani, H.E. Broxmeyer and C.H. Kim, 2007. Vitamin A metabolites induce gut-homing FoxP3+ regulatory T cells. J. Immunol., 179: 3724-3733.
- Kastner, P., H.J. Lawrence, C. Waltzinger, N.B. Ghyselinck, P. Chambon and S. Chan, 2001. Positive and negative regulation of granulopoiesis by endogenous RARalpha. Blood, 97: 1314-1320.
- Kawaguchi, J., P.J. Mee and A.G. Smith, 2005. Osteogenic and chondrogenic differentiation of embryonic stem cells in response to specific growth factors. Bone, 36: 758-769.
- Kennedy, K.A., T. Porter, V. Mehta, S.D. Ryan, F. Price et al., 2009. Retinoic acid enhances skeletal muscle progenitor formation and bypasses inhibition by bone morphogenetic protein 4 but not dominant negative beta-catenin. BMC. Biol., 7: 67.
- King, L.A., N.B. Schwartz and M.S. Domowicz, 2009. Glial migratory streams in the developing hindbrain: A slice culture approach. J. Neurosci. Methods, 177: 30-43.
- Kinoshita, T., K. Koike, H.H. Mwamtemi, S. Ito and S. Ishida *et al.*, 2000. Retinoic acid is a negative regulator for the differentiation of cord blood-derived human mast cell progenitors. Blood, 95: 2821-2828.

- Kirimoto, A., Y. Takagi, K. Ohya and H. Shimokawa, 2005.
 Effects of retinoic acid on the differentiation of chondrogenic progenitor cells, ATDC5. J. Med. Dent Sci., 52: 153-162.
- Kosaka, C., T. Sasaguri, Y. Komiyama and H. Takahashi, 2001. All-trans retinoic acid inhibits vascular smooth muscle cell proliferation targeting multiple genes for cyclins and cyclin-dependent kinases. Hypertens Res., 24: 579-588.
- Lane, M.A. and S.J. Bailey, 2005. Role of retinoid signalling in the adult brain. Prog. Neurobiol., 75: 275-293.
- Leelasiri, A., T. Numbenjapol, W. Prayoonwiwat, W. Mongkolsritrakul and C. Srisawat, 2005. Successful treatment of retinoic acid syndrome with dexamethasone: A case report. J. Med. Assoc. Thai., 88: 302-310.
- Li, Z., P. Manna, H. Kobayashi, T. Spilde and A. Bhatia et al., 2004. Multifaceted pancreatic mesenchymal control of epithelial lineage selection. Dev. Biol., 269: 252-263.
- Linderholm, A.L., J. Onitsuka, C. Xu, M. Chiu, W.M. Lee and R.W. Harper, 2010. All-trans retinoic acid mediates DUOX2 expression and function in respiratory tract epithelium. Am. J. Physiol. Lung. Cell. Mol. Physiol., 299: L215-221.
- Lu, L., T. Yao, Y.Z. Zhu, G.Y. Huang, Y.X. Cao and Y.C. Zhu, 2003. Chronic all-trans retinoic acid treatment prevents medial thickening of intramyocardial and intrarenal arteries in spontaneously hypertensive rats. Am. J. Physiol. Heart Circ. Physiol., 285: H1370-1377.
- Maden, M., 2006. Retinoids and spinal cord development. J. Neurobiol., 66: 726-738.
- Maden, M., 2007. Retinoic acid in the development, regeneration and maintenance of the nervous system. Nat. Rev. Neurosci., 8: 755-765.
- Malladi, P., Y. Xu, G.P. Yang and M.T. Longaker, 2006. Functions of vitamin D, retinoic acid, and dexamethasone in mouse adipose-derived mesenchymal cells. Tissue. Eng., 12: 2031-2040.
- Manabe, I. and G.K. Owens, 2001. Recruitment of serum response factor and hyperacetylation of histones at smooth muscle-specific regulatory regions during differentiation of a novel P19-derived in vitro smooth muscle differentiation system. Circ. Res., 88: 1127-1134.
- Manolescu, D.C., R. El-Kares, L. Lakhal-Chaieb, A. Montpetit, P.V. Bhat and P. Goodyer, 2010. Newborn serum retinoic acid level is associated with variants of genes in the retinol metabolism pathway. Pediatr. Res., 67: 598-602.

- McCaffery, P.J., J. Adams, M. Maden and E. Rosa-Molinar, 2003. Too much of a good thing: Retinoic acid as an endogenous regulator of neural differentiation and exogenous teratogen. Eur. J. Neurosci., 18: 457-472.
- Mey, J. and P. McCaffery, 2004. Retinoic acid signaling in the nervous system of adult vertebrates. Neuroscientist, 10: 409-421.
- Mey, J., 2006. New therapeutic target for CNS injury? The role of retinoic acid signaling after nerve lesions. J. Neurobiol., 66: 757-779.
- Murtaugh, L.C. and D.A. Melton, 2003. Genes, signals, and lineages in pancreas development. Annu. Rev. Cell Dev. Biol., 19: 71-89.
- Nabeyrat, E., S. Corroyer, V. Besnard, V. Cazals-Laville, J. Bourbon and A. Clement, 2001. Retinoic acid protects against hyperoxia-mediated cell-cycle arrest of lung alveolar epithelial cells by preserving late G1 cyclin activities. Am. J. Respir. Cell Mol. Biol., 25: 507-514.
- Natarajan, S.K., G.J. Amirtharaj, A. Ramachandran, A.B. Pulimood and K.A. Balasubramanian, 2009. Retinoid metabolism in the small intestine during development of liver cirrhosis. J. Gastroenterol. Hepatol., 24: 821-829.
- Niederreither, K., J. Vermot, N. Messaddeq, B. Schuhbaur, P.Chambon and P. Dolle, 2001. Embryonic retinoic acid synthesis is essential for heart morphogenesis in the mouse. Development, 128: 1019-1031.
- Nobakht, M., A. Zirak, M. Mehdizadeh and P.Tabatabaeei, 2006. Teratogenic effects of retinoic acid on neurulation in mice embryos. Pathophysiology, 13: 57-61.
- Noy, N., 2010. Between death and survival: Retinoic acid in regulation of apoptosis. Annu. Rev. Nutr., 30: 201-217.
- Oliva, A., A. Borriello, S. Zeppetelli, A. Di Feo and P. Cortellazzi et al., 2003. Retinoic acid inhibits the growth of bone marrow mesenchymal stem cells and induces p27Kipl and p161NK4A up-regulation. Mol. Cell. Biochem., 247: 55-60.
- Otsuki, T., H. Sakaguchi, T. Hatayama, P. Wu, A. Takata and F. Hyodoh, 2003. Effects of all-trans retinoic acid (ATRA) on human myeloma cells. Leuk Lymphoma, 44: 1651-1656.
- Penniston, K.L. and S.A. Tanumihardjo, 2006. The acute and chronic toxic effects of vitamin A. Am. J. Clin. Nutr., 83: 191-201.
- Radio, N.M., T.M. Freudenrich, B.L. Robinette, K.M. Crofton and W.R. Mundy, 2010. Comparison of PC12 and cerebellar granule cell cultures for evaluating neurite outgrowth using high content analysis. Neurotoxicol. Teratol., 32: 25-35.

- Rexer, B.N., W.L. Zheng and D.E. Ong, 2001. Retinoic acid biosynthesis by normal human breast epithelium is via aldehyde dehydrogenase 6,absent in MCF-7 cells. Cancer Res., 61: 7065-7070.
- Ross, S.A., P.J. McCaffery, U.C. Drager and L.M. de Luca, 2000. Retinoids in embryonal development. Physiological. Rev., 80: 1021-1054.
- Ryckebusch, L., N. Bertrand, K. Mesbah, F. Bajolle and K. Niederreither *et al.*, 2010. Decreased levels of embryonic retinoic acid synthesis accelerate recovery from arterial growth delay in a mouse model of DiGeorge syndrome. Circ. Res., 106: 686-694.
- Seguin-Devaux, C., D. Hanriot, M. Dailloux, V. Latger-Cannard and F. Zannad et al., 2005. Retinoic acid amplifies the host immune response to LPS through increased T lymphocytes number and LPS binding protein expression. Mol. Cell. Endocrinol., 245: 67-76.
- Shen, C.N., A. Marguerie, C.Y. Chien, C. Dickson, J.M. Slack and D. Tosh, 2007. All-trans retinoic acid suppresses exocrine differentiation and branching morphogenesis in the embryonic pancreas. Differentiation, 75: 62-74.
- Skillington, J., L. Choy and R. Derynck, 2002. Bone morphogenic protein and retinoic acid signaling cooperate to induce osteoblast differentiation of preadipocytes. J. Cell Biol., 159: 135-146.
- Song, H.M., R.P. Nacamuli, W. Xia, A.S. Bari, Y.Y. Shi, T.D. Fang and M.T. Longaker, 2005. High-dose retinoic acid modulates rat calvarial osteoblast biology. J. Cell Physiol., 202: 255-262.
- Stoppie, P., M. Borgers, P. Borghgraef, L. Dillen and J. Goossens et al., 2000. R115866 inhibits all-transretinoic acid metabolism and exerts retinoidal effects in rodents. J. Pharmacol. Exp. Ther., 293: 304-312.
- Su, Z.Y., Y. Li, X.L. Zhao and M. Zhang, 2010. All-trans retinoic acid promotes smooth muscle cell differentiation of rabbit bone marrow-derived mesenchymal stem cells. J. Zhejiang. Univ. Sci. B., 11: 489-496.
- Subbarayan, V., M. Mark, N. Messadeq, P. Rustin, P. Chambon and P. Kastner, 2000. RXR alpha overexpression in cardiomyocytes causes dilated cardiomyopathy but fails to rescue myocardial hypoplasia in RXR alpha -null fetuses. J. Clin. Invest., 105: 387-394.
- Teitelbaum, S.L., 2000. Osteoclasts, integrins and osteoporosis. J. Bone Miner. Metab., 18: 344-349.
- Thomas, S., R. Prabhu and K.A. Balasubramanian, 2005. Retinoid metabolism in the rat small intestine. Br. J. Nutr., 93: 59-63.

- Tober, J., A. Koniski, K.E. McGrath, R. Vemishetti and R. Emerson et al., 2007. The megakaryocyte lineage originates from hemangioblast precursors and is an integral component both of primitive and of definitive hematopoiesis. Blood, 109: 1433-1441.
- Tulachan, S.S., R. Doi, Y. Kawaguchi, S. Tsuji and S. Nakajima et al., 2003. All-trans retinoic acid induces differentiation of ducts and endocrine cells by mesenchymal/epithelial interactions in embryonic pancreas. Diabetes, 52: 76-84.
- Van de Pavert, S.A., B.J. Olivier, G. Goverse, M.F. Vondenhoff and M. Greuter et al., 2009. Chemokine CXCL13 is essential for lymph node initiation and is induced by retinoic acid and neuronal stimulation. Nat. Immunol., 10: 1193-1199.
- Verfaille, C.J., M. Borgers and M.A. van Steensel, 2008. Retinoic acid metabolism blocking agents (RAMBAs): A new paradigm in the treatment of hyperkeratotic disorders. J. Dtsch. Dermatol. Ges., 6: 355-364.
- Wang, H.J., Y.C. Zhu and T. Yao, 2002. Effects of all-trans retinoic acid on angiotensin II-induced myocyte hypertrophy. J. Applied Physiol., 92: 2162-2168.
- Wang, X.P., M. Suomalainen, C.J. Jorgez, M.M. Matzuk, S. Werner and I. Thesleff, 2004. Follistatin regulates enamel patterning in mouse incisors by asymmetrically inhibiting BMP signaling and ameloblast differentiation. Dev. Cell, 7: 719-730.
- Yamaguchi, Y. and R. Igarashi, 2006. Nanotechnology for therapy of type 2 diabetes. Nippon. Rinsho., 64: 295-300.
- Yousefi, B., 2009. The effects of all-trans retinoic acid on blood cells in rats embryo. Pak. J. Pharm. Sci., 22: 23-26.

- Yousefi, B. and H.R. Samene, 2010. The effects of all-trans retinoic acid on leukocytes in rats Embryo. Pak. J. Biol. Sci., 13: 185-189.
- Yousefi, B., N. Bromand and S. Haghighi, 2010. Effects of all-trans retinoic acid on the pancreas development in rat embryo. Iran J. Endocrinol. Metab., 11: 576-582.
- Yu, C., Y. Liu, Z. Miao, M. Yin and W. Lu et al., 2010. Retinoic acid enhances the generation of hematopoietic progenitors from human embryonic stem cell-derived hemato-vascular precursors. Blood, 10.1182/blood. DOI:-2010-01-263335
- Yu, J., Z. Liu, M. Yang and A. Mak, 2009. Nanoporous membrane-based cell chip for the study of anti-cancer drug effect of retinoic acid with impedance spectroscopy. Talanta, 80: 189-194.
- Yuncu, M., A. Eralp, M. Koruk, I. Sari, C. Bagei and S. Inaloz, 2004. Effect of vitamin A against methotrexate-induced damage to the small intestine in rats. Med. Princ. Pract., 13: 346-352.
- Zegers, M.M., L.E. O'Brien, W. Yu, A. Datta and K.E. Mostov, 2003. Epithelial polarity and tubulogenesis in vitro. Trends. Cell Biol., 13: 169-176.
- Zhang, E., B. Jiang, A. Yokochi, J. Maruyama and Y. Mitani *et al.*, 2010. Effect of all-trans-retinoic Acid on the development of chronic hypoxia-induced pulmonary hypertension. Circ. J., 74: 1696-1703.
- Zhang, J.W., J.Y. Wang, S.J. Chen and Z. Chen, 2000. Mechanisms of all-trans retinoic acid-induced differentiation of acute promyelocytic leukemia cells. J. Biosci., 25: 275-284.