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New Records of Eriophyoid Mites (Acari: Prostigmata: Eriophyoidea) from Saudi Arabia

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Abstract: A field study was conducted to investigate eriophyoid mites associated with some fruit trees in Riyadh, Saudi Arabia. The survey was carried out in four localities (El-Waseel, Al-Beer, Al-Haiyer and El-Deriya) and included seven species of fruit trees, namely olives (Olea europea), fig (Ficus carica), grapes (Vitis vinifera), apple (Malus domestica), citrus (Citrus spp.), pomegranate (Punica granatum) and pear (Pyrus communis). Seven new records of eriophyoid species (Aceria benghalensis Soliman and Abou-Awad, A. olivi Zaher and Abou-Awad, Caleptrimerus baileyi K., Colomerus oculivitis (Attiah), Oxycenus niloticus (Zaher and Abou-Awad), Rhynchaphytoptus ficifolia (Keifer) and Tegolophus hassani (Keifer)), belonging two families, Eriophyidae and Diptilomiopidae, were collected from four species of fruit crops covering four different production localities in Riyadh. An illustrated identification key for these mites is provided. The present study is the first scientific study on Saudi eriophyoid mites.

Key words: Eriophyoid mites, fruit crops, fauna of Saudi Arabia, olive, new record

INTRODUCTION

Eriophyoid mites have a worldwide distribution whereas more than 3,690 species are known (Amrine *et al.*, 2003). Members of Eriophyidae and Diptilomiopidae families live on trees, shrubs and herbaceous plants causing heavy injuries to leaves, buds and fruits (Zaher, 1986; Ripka, 2008). In addition, some of these mites are able to transmit some virus diseases, causing complete deterioration to the trees (Kozlowski, 2000; Stenger *et al.*, 2005; Smales *et al.*, 1996).

Mite fauna of Saudi Arabia (SA) has received a very little consideration and the groundwork for essential taxonomic, biological and ecological studies of agricultural mites is extremely rare. Despite the economic importance of many different fruit crops grown in SA, such as date palm, olives and citrus trees, a few numbers of eriophyoid mite species have been reported (Martin, 1972).

Fruit crops like fig (Ficus carica), grapevine (Vitis vinifera), citrus (Citrus spp.), apple (Malus domestica) and olive (Olea europea) are extremely important for local consumption. However, olive occupies the most advanced position in terms of acreage and fruit production for both local consumption and exportation. There is only one eriophyoid specis (Aceria oleae Nalepa) that has been recorded on olive in SA (Martin, 1972). However, previous studies recorded 11 eriophyid mite species on olive trees in different countries, namely,

Aceria oleae Nalepa, Oxycenus maxwelli Keifer, Ditrymacus athiasella Keifer, Aculus olearius (Castagnoli), Shevtchenkella oleae (Natcheff), Aceria Olive (Zaher and Abou-Awad), Oxycenus niloticus (Zaher and Abou-Awad), Aculops benakii (Hatzinikolis), Aceria cretica (Hatzinikolis), Shevtchenkella bicarinata (Meyer) and Tegolophus Hassani (Keifer) (Snjezana et al., 2009).

The objectives of this study were to (1) investigate eriophyoid mites associated with some fruit trees in Riyadh and (2) provide an identification key for new recorded mites species found in this study. The mite identification key presented in this study will serve to pave the way for further taxonomical investigations for the super family Eriophyiodea in the kingdom.

MATERIALS AND METHODS

The survey was conducted in four localities surrounding Riyadh city (El-Waseel, Al-Beer, Al-Haiyer and El-Deriya) and included seven species of fruit trees, namely olives (*Olea europea*), fig (*Ficus carica*), grapes (*Vitis vinifera*), apple (*Malus domestica*), citrus (*Citrus* spp.), pomegranate (*Punica granatum*) and pear (*Pyrus communis*). At each locality, sampling was carried out irregularly from October 2008 to September 2009 including plant foliages, branches and buds. Samples were individually bagged in tightly-closed plastic bags and transported the same day to the Acarology Laboratory,

Department of Plant Protection, College of Food and Agriculture Sciences, King Saud University for mite extraction.

Mites were extracted from leaves and buds samples by using a fine hair brush under a high quality Olympus stereo-microscope (SZX-10) at magnifications 100-200×. Subsequently, mites were mounted onto micro-slides with Keifer medium and later dried in a 40°C-oven for one week.

An Olympus compound microscope (BX-51) with an attached drawing tube was used for examination and initial pencil drawing of mite diagnostic features at magnifications of 400-1200×. The oil lens was used to examine the featherclaws, microtubercles and male and female genitalia. The line drawings of mites were scanned and imported into Adobe Photoshop and used as templates for final illustrations in Adobe illustrator. The figure measurement lines were fixed at 25 micrometer (μ m).

The terminology and morphological characteristics used in the identification key presented in this study are based on Amrine *et al.* (1996) and Amrine *et al.* (2003).

All specimens were deposited in the King Saud University Museum of Arthropod (KSMA), College of Food and Agriculture Sciences, King Saud University.

RESULTS

Seven eriophyoid mite species, namely, Aceria benghalensis Soliman and Abou-Awad, A. olivi Zaher and Abou-Awad, Caleptrimerus baileyi K., Colomerus oculivitis (Attiah), Oxycenus niloticus Zaher and Abou-Awad, Rhynchaphytoptus ficifoliae Keifer and Tegolophus hassani (Keifer), belonging to two families (Eriophyidae and Diptilomiopidae) were collected from leaves, buds and branches of four fruit crop species (Ficus carica, Olea europea, Malus domestica and Vitis vinifera) in different localities in Riyadh (Table 1). All of

Table 1: List of eriophyoid mites associated with various parts of four fruit crops from different localities, Riyadh, SA.

Mites recorded	Host plant and part(-s) infested	Locality
Aceria benghaleusis	Ficus carica	1, 2, 4
	(Under scales of open buds)	
A. olive	Ole a europea	2, 3, 4
	(Lower surface of leaves)	
Caleptrimerus baileyi	Malus domestica	3
	(Upper surface of leaves)	
Colomerus oculivitis	Vītis vinifera	3
	(Buds and branches)	
Oxycenus niloticus	Ole a europe a	2
	(Upper surface and around the	
	midvine of leaves)	
Rhynchaphytoptus ficifoliae	Ficus carica	1, 2, 3, 4
	(Lower surface of leaves)	
Tegolophus hassani	Oleaeuropea	1, 4
	(Upper and lower surface of leaves)	

¹⁻ Al-Beer 2- Al-Deriya 3- Al-Haiyer 4- El- Waseel

these seven mites found in this study are new records in SA. Plant samples collected from the fruit crop species, *Citrus* spp., *Punica granatum* and *Pyrus communis*, were free from mites.

DISCUSSION

Eriophyoid mites are one of the most specialized groups of plant feeders and symptoms of their feeding run the gamut from simple rusting to complex gall formation and they can transmit plant viruses posing a great threat on the yield of the crops (Keifer *et al.*, 1982).

Martin (1972) reported only four eriophyoid species (Aceria oleae Nalepa, Eriophyes granati Can., Aceria sheldoni Ewing and Phyllocoptruta oleivora Ashm.) on olive, pomegranate and citrus, respectively, in different regions of SA. Interestingly, in this study the four mite species reported by Martin (1972) were not recovered from the tree plants representing seven different species investigated in our survey.

The two species (Aceria benghalensis and Rhynchaphytoptus ficifoliae) extracted from fig buds and leaves were previously recorded in Egypt (Zaher, 1986), India (Mondal and Chokrabatris, 1982) and Iraq (Al-Mallah and Mohammad, 1989). Aceria benghalensis causes discoloration of scales and partial blasting of buds while Rhynchaphytoptus ficifoliae does not induce obvious symptoms (Mondal and Chokrabatris, 1982; Zaher, 1986; Al-Mallah and Mohammad, 1989). Colomerus oculivitis, found on grapevine, was recorded in Egypt and in U.S. (Attiah, 1967; Zaher, 1986; Amrine and Stasny, 1994). It causes scarlet and brown coloration around edges of leaves spread inwards causing dryness of leaves. Caleptrimerus baileyi was previously recorded in U.S. causing slight browning to the leaf surfaces and the deutogynes hibernate around the buds just back of the terminal bud (Keifer, 1952).

Olive is a strategic crop in SA and about 13 million trees are grown in Tabouk, El-gouf and Haiyl regions (Personal communication, Plant Production Department, Agriculture College, King Saud University). In this study, three mite species (Aceria olivi, Oxycenus niloticus and Tegolophus hassani) were extracted from olive plants. Members of these mites usually live underside terminal olive leaves where they insert themselves under stellate hairs, causing these stellate structures to drop off and making yellow leaf spots and various forms of leaf deformation or defoliation. Oxycenus niloticus is normally found on upper surface of terminal olive leaves and pitting produce some 1eaf and deformation (Hatzinikolis et al., 1989; Shahinia et al., 2009). These mites were previously recorded in different countries as follows: A. olive recorded in Egypt (Zaher, 1986; Hatzinikolis, 1989); O. niloticus recorded in Iran and

Egypt (Mahisani et al., 2004; Zaher, 1986) and T. hassani recorded in Egypt and Albania (Shahinia et al., 2009; Zaher, 1986). This study sheds the light on the importance of these mite species and more attention is needed regarding their biology, ecology and control.

The identification key presented in this study should encourage other researchers for more investigations in identifying eriophyoid mites in SA.

Identification key for adult stages of eriophyoid mites associated with olive, Olea europea; fig, Ficus carica; grapevine, Vitis vinifera and apple, Malus domestica in Riyadh region.

 Rostrum usually small, not abruptly bent down near bases; oral stylet short; dorsal setae various present or absent Eriophyidae Nalepa

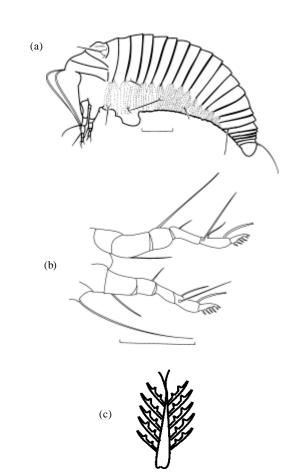


Fig. 1: Rhynchaphytoptus ficifoliae Keifer; (a) Side view; (b) Legs and (c) Featherclaw



Fig. 2: Genital coverflap of Colomerus oculivitis (Attiah)

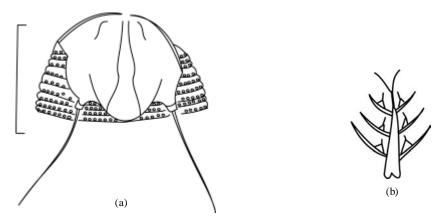


Fig. 3: Aceria olivi Zaher and Abou- Awad; (a) Dorsal shield and (b) Featherclaw

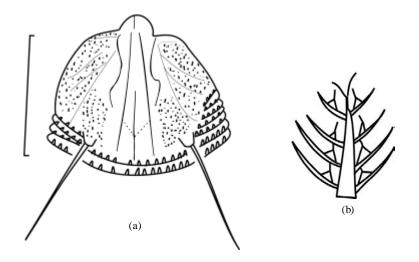


Fig. 4: Aceria benghalensis Soliman and Abou-Awad; (a) Dorsal shield and (b) Featherclaw

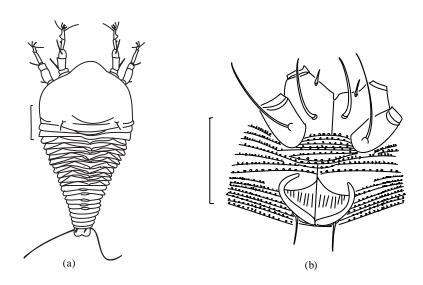


Fig. 5: Oxycenus niloticus Zaher and abou-Awad; (a) Dorsal view and (b) Genital coverflap

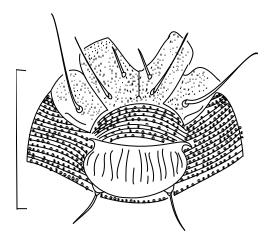


Fig. 6: Female genitalia and body anterior of *Tegolophus hassani* (Keifer)

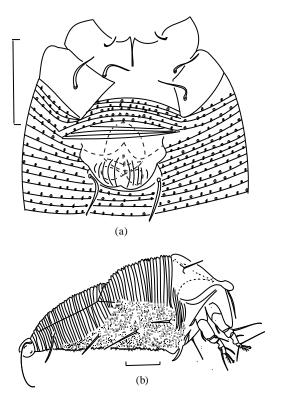


Fig. 7: Caleptrimerus bailyei Kiefer; (a) Female genitalia and body anterior and (b) Side view

 Posterior opithosoma without rear depression; scapular setae variable........6

Stage 6: Anterior coxae not separated *Tegolophus* Keifer anterior and posterior coxae with heavy lines of

granules curving around setiferous tubercles (Fig. 6) *Tegolophus hassani* (Keifer)

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