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Association of CTLA-4 Gene 49A/G Polymorphism with the Incidence of Type 1 Diabetes Mellitus in the Iranian Kurdish Population

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Abstract: Cytotoxic T-lymphocyte-associated antigen-4 (CTLA-4) has an inhibitory function on T cells and is critical for the induction of peripheral tolerance. CTLA-4+49 G allele affects the CTLA-4 function and has been reported to be correlated with a higher risk of various autoimmune diseases including type 1 diabetes (T1D). The present study was conducted to investigate the association between the polymorphism of the CTLA-4 exon 1+49 A/G and susceptibility to TID and type 2 diabetes (T2D) in Kurds living in Iranian Kurdistan. The+49 A/G polymorphism was analyzed in 60 patients with T1D, 56 patients with T2D and 107 control subjects using PCR Single-strand Conformation Polymorphism (SSCP) and restriction fragment length polymorphism methods. All studied populations (T1D, T2D and Controls) were in Hardy-Weinberg equilibrium (p, 0.39, 0.94 and 0.89, respectively). Both+49 G allele (p = 0.015, OR = 1.86) and +49 A/G genotype frequencies (p = 0.012, OR = 2.31) were significantly higher in T1D patients than control. There was significant over-representation of the G allele in female T1D patients. No significant differences in +49 G allele and +49 A/G genotype frequencies were found between T2D and control subjects. SSCP analysis did not show new mutation in the amplified segment. The results of this study indicate that CTLA-4+49 A/G gene polymorphism confers genetic susceptibility to T1D but not T2D in the Kurdish population living in Iranian Kurdistan and women carrying the +49 G allele are at greater risk of getting T1D than men having the G allele.

Key words: Type 1 diabetes, CTLA-4, polymorphism, susceptibility, loci

INTRODUCTION

Type 1 diabetes (T1D) is a chronic disease in which pancreatic islet β cells will be targeted for destruction by the immune system (Atkinson and Eisenbarth, 2001). Genetic and environmental factors are involved in the fast growing incidence of the disease (Gale, 2002). High concordance rates in monozygotic and dizygotic twins indicate the importance of genetic factors in the etiology of T1D (Hyttinen et al., 2003). Based on genetic traits, those at high genetic risk can be diagnosed at birth or before islet autoantibodies are detectable in plasma. The best time for treatment of T1D in susceptible subjects is the period between progressing islet beta-cell destruction and the appearance of the symptoms of T1D, when there are enough functional beta cells remaining to be rescued through various intervention methods (Van der Auwera et al., 2002; Van Belle et al., 2011).

Human Leukocyte Antigen (HLA) is the most important genetic determinant of T1D. It has been

guessed that HLA-mediated susceptibility can be accounted for about half of the genetic risk of T1D (Mehers and Gillespie, 2008). Genome-wide association studies have resulted in the identification of a lot of non-HLA susceptibility loci for T1D including IDDM12 located on chromosome 2q33, that encodes for key lymphocyte co-receptor genes, including cytotoxic T lymphocyte antigen-4 (CTLA-4) and CD28 (Grant and Hakonarson, 2009). CTLA-4, also known as CD152, is a cell-surface transmembrane glycoprotein which is expressed mainly on CD4+ and CD8+ T cell subsets and plays a key role in the fine tuning of T-cell immunity by providing a negative signal for T-cell activation (Carreno et al., 2000; Wells et al., 2001). CTLA-4 indirectly, via competing with CD28 for CD80/86 binding attenuates stimulatory signal to T-cells.

There are contradictory results regarding the connection between the CTLA-4 gene polymorphisms and genetic susceptibility to some autoimmune diseases (Kristiansen *et al.*, 2000). Among CTLA-4 gene

polymorphisms, an adenine to guanine transition at position 49 of exon 1, rs231775, also known as +49A/G leads to an alanine to threonine amino acid substitution at codon 17 of the leader peptide (Nistico *et al.*, 1996). +49A/G leads to less efficient glycosylation and reduced expression of membrane CTLA4 protein (Anjos *et al.*, 2002). Reduced CTLA-4 expression and/or change in CTLA-4 activity result in uncontrolled T-cell-associated autoimmunity disease (Waterhouse *et al.*, 1995; Rudd and Schneider, 2003).

Some studies found an association between +49A/G and susceptibility to T1D (Marron et al., 1997; Nistico et al., 1996), whereas others found no evidence for such association (Owerbach et al., 1997). Inconsistency in results illustrates the importance of further studies in various ethnicities. The Iranian Kurds, primarily living in the west of Iran, constitute approximately 7% of Iran's overall population. To our knowledge there is no existing data regarding the association between +49A/G and susceptibility to T1D or T2D in Iranian Kurdish population. The present study was conducted to investigate the association of CTLA-4 SNP +49A/G with a predisposition to T1D and T2D in Iranian Kurdish population.

MATERIALS AND METHODS

The study subjects comprised 60 unrelated T1D patients, 56 T2D patients and 107 healthy controls with no previous history of T1D or other autoimmune diseases. Diabetic patients were recruited from Kurdistan province diabetes center located in Tohid hospital, Sanandaj. The diagnosis of diabetes mellitus in the center was based on the American Diabetes Association diabetes diagnostic criteria. The research protocol was approved by the ethics committee at the Kurdistan University of Medical Sciences. All subjects enrolled in the study signed consent and institutional ethical requirements were met.

DNA Extraction and CTLA-4 Genotyping: Genomic DNA was extracted from frozen whole blood using the Promega wizard DNA Purification Kit (Cat. #A1120). CTLA-4+49A/G polymorphism was defined using a polymerase chain reaction-restricted fragment length polymorphism (PCR-RFLP) method as described by Dilmec *et al.* (2008). The PCR was performed using DreamTaq PCR Master Mix (#K1071) in a 50 μL reaction mixture containing Dream*Taq* PCR Master Mix (2X) 25 μL, Forward primer 1.0 μM, Reverse primer 1.0 μM, Template DNA 50 ng, Water, nuclease-free to 50 μL.

PCR profile for CTLA-4 +49A/G polymorphism: The PCR conditions comprised an initial denaturation at 94°C for 3 min, followed by 30 cycles of denaturation at 94°C

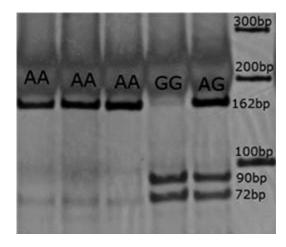


Fig. 1: Genotyping of CTLA-4+49A/G polymorphism by Bbv I RFLP, From right: Lane 1: 100-300 bp DNA ladder, Lane 2: Heterozygous (AG, 162, 90 and 72 bp), Lane 3: Homozygous (GG, 90 and 72 bp) and Lanes 4, 5 and 6: Wild type (AA genotype, 162 bp)

for 45 sec, annealing at 58° C for 45 sec and extension at 72° C for 30 sec, with a final extension at 72° C for 6 min. 5 μ L PCR product (162 bp) was digested using 1.5 U of Bbv I (Bse XI, 100u #ER1451) in a 20 μ L volume overnight at 65°C. Digested A allele yielded a fragment of a 162 bp and G allele yielded 90 and 72 bp fragments (Fig. 1). The primers were 5'-GCT CTA CTT CCT GAA GAC CT-3' (forward) and 5'-AGT CTC ACT CAC CTT TGC AG-3' (reverse).

Single-strand conformational polymorphism (SSCP): For confirmation of RFLP results and finding possible new mutations in the amplicon, SSCP was performed as described (Takahashi *et al.*, 1995). For this purpose 2μL aliquot of the product was diluted in 48 μL of denaturing loading buffer (95% formamide, 10 mM NaOH, 0.05% bromophenol blue and 0.05% xylene cyanol FF), heated at 95°C for 10 min and then immediately cooled on ice for 10 min. Aliquots of 7 μL were then loaded on a 12% polyacrylamide gel at 8 W constant power for 16-18 h at 4°C and finally silver stained.

Statistical analysis: The distribution of genotypes for 49 A/G SNP was assessed for deviation from the Hardy-weinberg Equilibrium (HWE) by chi square testing. Genotype and allele frequencies of the 49 A/G SNP analyzed in cases and controls with the De Finetti program (http://ihg.gsf.de/cgi-bin/hw/hwal.pl). The dominant/ recessive effect of single (AA, AG and GG) and combined (e.g. AG/GG vs. AA) effects of CTLA-4 genotypes on phenotype (disease) expression was examined in studied population using the Finetti program.

RESULTS

Genotype analysis: Samples from 60 patients with T1D, 56 patients with T2D and 107 healthy controls were successfully genotyped for CTLA-4 +49A/G polymorphism. The genotype frequency distribution of +49A/G was in Hardy-Weinberg equilibrium in T1D, T2D and control groups (p, 0.39, 0.94 and 0.89, respectively). Genotype and allele frequencies of +49A/G in T1D patients were significantly different from both T2D and healthy controls (Table 1). T1D patients had a higher frequency of heterozygous +49A/G (Odds Ratio (OR) = 2.308, C.I. = 1.18-4.48, p = 0.01), G allele frequency difference, (OR = 1.86, C.I. = 1.12-3.08, p = 0.01), allele positivity (AA versus AG+GG), (OR = 2.34, C.I. = 1.23-4.47, p = 0.01) versus both control and T2D groups. The Cochran-armitage test for trend results showed the presence of strong association between G allele with T1D (common OR = 1.85, p = 0.01) but not with T2D (OR = 1.10, p = 0.85). CTLA-4 49 G allele frequency was not increased in diabetic patients compared to controls (21.4 vs. 20.6%, not significant). The distribution of GG, AG and AA CTLA-4 genotypes was similar in T2D and control groups.

Gender analysis: The gender distribution of alleles and genotype frequencies of CTLA-4 +49A/G in T1D and control groups is shown in Table 2. T1D female patients had a higher frequency of heterozygous +49A/G (OR = 2. 47, C.I. = 1.07-5.69, p = 0.03) and allele positivity (OR = 2.35, C.I. = 1.04-5.29, p = 0.03) but not homozygous G allele than female control groups. G allele frequency

difference, (OR = 1.74, C.I. = 0.92-3.31, p = 0.08) was not significant among female T1D patients and female control groups. The Cochran-armitage test for trend results did not show an association between G allele with T1D (OR = 1.63, p = 0.07). Male T1D patients did not show significant differences for homozygous G allele (p = 0.07), heterozygous +49A/G (p = 0.18) and allele positivity (p = 0.11) with the male control group.

DISCUSSION

Type 1 diabetes is a chronic autoimmune disease in which destruction or damaging of the pancreatic islet beta-cells results in insulin deficiency and hyperglycemia (La Torre, 2012). There have been intensive efforts in recent years to reverse or ameliorate the diabetic state in T1D patients or animal models of T1D through various methods including, immune interventions (Lemmark and Larsson, 2013), restoration of the beta-cell mass (Cam et al., 1999; Ahmadi et al., 2010) or potentiation of peripheral insulin action (Dehghani et al., 1997). Those strategies are more effective in the earlier stages of the autoimmune attack, when there are enough remaining healthy islet beta cells. In this study the contributions of CTLA-4 +49 A/G genotype to T1D and T2D susceptibility was tested in Iranian Kurdish population. Statistically significant differences were found in both G allele and heterozygous A/G genotype distribution among T1D patients with both T2D patients and control subjects but not between T2D patients and controls. The T1D risk was not increased in the homozygous form of G allele probably due to relatively

Table 1: Allele and genotype frequencies of CTLA-4+49A/G in T1D, T2D and control groups

		No. (percent of total)	p-value			
Group		T1D $(n = 60)$	T2D (n = 56)	Control (n = 107)	T1D	T2D
Allele	A	81 (67.5)	98 (78.6)	170 (79.4)		
	G	39 (32.5)	24 (21.4)	44 (20.6)	0.01*	0.85
Genotype	A/A	25 (41.7)	35 (62.5)	67 (62.6)		
	A/G	32 (53.3)	18 (32.1)	36 (33.6)	0.01*	0.90
	G/G	3 (5.0)	3 (5.0)	4 (3.7)		
	(AG+GG)	35 (58.3)	21 (37.5)	40 (37.4)	0.01*	0.98

^{*}Significant difference from the control group p-value<0.05, T1D: Type 1 diabetes, T2D: Type 2 diabetes

Table 2: The gender distribution of alleles and genotype frequencies of CTLA-4+49A/G in T1D and control groups

		No. (percent of total)	No. (percent of total)						
		TID		Control		p-value			
Group									
Gender		Female (n = 40)	Male $(n = 20)$	Female (n = 63)	Male (n = 44)	Female	<u> Male</u>		
Allele	A	55 (69)	26 (65)	100 (79.4)	70 (79.5)				
	G	25 (31)	14 (35)	26 (20.6)	18(20.5)	0.08	0.08		
Genotype	A/A	17 (42.5)	8 (40)	40 (63.5)	27 (61.4)				
	A/G	21 (52.5)	10 (50)	20 (31.7)	16 (36.3)	0.03*	0.18		
	G/G	2 (5.0)	2 (10)	3 (4.8)	1 (2.3)				
	(AG+GG)	23 (57.5)	12 (60)	23 (36.5)	17 (37.4)	0.03*	0.11		

^{*}Significant difference from (female) Control group, T1D: Type 1 diabetes, T2D: Type 2 diabetes

low number of homozygous form of G allele in both cases and controls. The CTLA4 49 A/G genotype was found to be more frequently present in female T1D patients rather than male patients as compared to related control groups.

The incidence and prevalence of T1D are rapidly increasing worldwide (Maahs et al., 2010). Both genetic and environmental factors contribute to the risk of T1D (Gan et al., 2012). Gene polymorphisms in at-risk populations tilt the balance in their immune system toward self-destruction (Jahromi and Eisenbarth, 2007). Detection of the potential T1D candidates in an early stage of diabetes is required for on-time and effective therapeutic interventions. The most important genetic susceptibility locus for T1D is attributed to the HLA class II region but polymorphisms of individual genes outside the MHC also contribute to diabetes risk (Grant and Hakonarson, 2009). CTLA-4 is a co-stimulatory receptor and a vital negative regulator of T cell activation. CTLA-4 gene is a potential candidate for T cell-mediated autoimmune disorders (Benmansour et al., 2010; Chen et al., 2013). There are several polymorphic regions within the CTLA-4 gene, including a CTLA-4-318C/T, CTLA-449A/G and CTLA-4 3' (AT)n. Adenine to Guanine substitution at position +49 in exon 1 and resulting threonine to alanine substitution in the CTLA-4 molecule leads to incomplete glycosylation (Anjos et al., 2002). Such changes in CTLA-4 molecule may influence immunoregulation and may be used as an explanation of the predisposition to autoimmune diseases in +49 G allele carriers (Vaidya and Pearce, 2004).

Inconsistent results have been reported regarding the association of CTLA4 +49A/G and susceptibility to T1D in different studies. Some studies conclude that the G allele of +49A/G is associated with a predisposition to T1D (Takara et al., 2000; Tang et al., 2012; Mosaad et al., 2012, Douroudis et al., 2009; Kavvoura and Ioannidis, 2005; Ueda et al., 2003; Chen et al., 2013) while others did not find such association (Lemos et al., 2009; Fajardy et al., 2002; Cinek et al., 2002). The current data showed that AG genotype accounted for 53.3% of T1D patients in comparison to 33.6% in controls. An odds ratio of 1.85 indicates a strong association between G allele and T1D disease in Iranian Kurdish population. A similar finding has been reported by other researchers in different ethnicities (Baniasadi et al., 2006; Turpeinen et al., 2003; Cosentino et al., 2002; Krokowski et al., 1998; Genc et al., 2004). Frequency of 49 G/G genotype was not significantly different between T1D patients and controls (5.0 vs. 3.7%), in this study, probably due to the small number of this genotype (n = 3 vs. n = 4, respectively). Mojtahedi et al. (2005) reported that +49A/G is associated with susceptibility to T1D in southern Iranian population. They found higher frequency of AG genotype in both T1D patients and controls (71.5 vs. 45%) in comparison to what was found in this study (53.3 vs. 33.6%). Statistically speaking results were similar, in both studies significant difference was found in frequency of G allele between T1D patients and controls.

Even though, type 2 diabetes mellitus (T2D) has been considered to be a metabolic disease, new findings including latent autoimmune diabetes in adult patients (Pettersen et al., 2010; Badaru and Pihoker, 2012) and the role of T cell modulation in the development of obesityassociated insulin resistance (Winer et al., 2011) makes it difficult to distinguish the borders between T1D and T2D. To find out if +49 A/G polymorphism is associated with T2D, the distribution of alleles as well as the genotypic frequencies were analyzed in T2D patients. No significant association was found between G allele/genotype and T2D disease, indicating that +49 G allele/genotype is not a major risk factor for T2D. The result of this study is in agreement with what reported before in T2D patients in Estonian and German populations (Haller et al., 2004; Rau et al., 2001). It looks carrying G allele or GG genotype is not a risk factor for T2D, but it may affect other aspects of T2D, like age at disease manifestation (Rau et al., 2001).

Based on the current data, higher numbers of female T1D patient are carriers of +49 G allele/genotypes compared to male T1D patients. Even though gene polymorphisms are linked to susceptibility to T1D, autoimmunity trigger factors appears to be specific for each affected individual (Peng and Hagopian, 2006). There may be epigenetic mechanisms that make females more susceptible to higher risk of carrying G alleles. Even though association of CTLA-4 +49 A/G with T1D was more profound in females, relatively small size of the studied population makes it difficult to get a firm conclusion. To confirm the gender dependence of the high risk CTLA-4 49A/G SNP in T1D patients and to avoid erroneous conclusion, more studies in a bigger sample size is needed. If the similar results can be repeated in larger population sizes, CTLA-4 49A/G SNP will find a more positive predictive value in risk-assessment of T1D in women.

In summary, the result of this study, supported by significant increase in frequency of +49 A/G genotype in T1D patients, show that CTLA-4 49 A/G is a genetic risk for T1D, but not T2D, in Iranian Kurdish population. Gender analyzes of data support the idea that women carrying+49 G allele are more likely to develop T1D but this proposal needs to be confirmed in a larger data set in follow-up studies. In conclusion, result of this study is in support of the engagement of CTLA-4+49 A/G in T1D pathogenesis and its potential application for screening potential T1D patients.

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