

<http://www.pjbs.org>

**PJBS**

ISSN 1028-8880

**Pakistan  
Journal of Biological Sciences**

**ANSI***net*

Asian Network for Scientific Information  
308 Lasani Town, Sargodha Road, Faisalabad - Pakistan



## Research Article

# Antioxidant Activity of Silymarin in Microcystin-LR Cardiac and Pulmonary Induced Injuries on Mice

Ayman Al-Hazmi

Faculty of Applied Medical Sciences, Taif University, Taif City, Saudi Arabia

## Abstract

**Background and Objectives:** Microcystin-LR has a toxic effect on several organs causing the destruction and fibrosis of these organs. This study was done to evaluate the antioxidant activity of silymarin on some oxidative stress parameters on heart and lung injuries induced by microcystin-LR in mice. **Materials and Methods:** Total 72 Balb/c male mice aged between five to seven weeks were grouped into 6; Group 1 contained twelve mice which were assigned as the healthy control group (C). Two microcystin-LR control groups assigned M6 and M12 contain 12 mice each. A fourth group contains twelve mice called the silymarin control group (S). The fifth and sixth groups contain twenty-four mice assigned as microcystin-LR silymarin groups SM6 and SM12. A blood sample was collected for estimation of serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST). The homogenates of heart and lungs were extracted for estimation of protein carbonyl content (CC), lipid peroxidation products (LPO), reduced glutathione (GSH), methylglyoxal (MG) and activity of protein phosphatase I (PPI). **Results:** Significant statistical differences in both ALT and AST were observed between all groups ( $P > 0.01$ ). In cardiac homogenate, a significant statistical difference was observed in PPI, LPO and CC between all groups ( $P < 0.05$ ). Furthermore, methylglyoxal showed a significant statistical difference between all groups ( $p < 0.01$ ). **Conclusion:** The findings suggested a potential therapeutic role of using silymarin as an antioxidant agent against cardiac and pulmonary injuries induced by MC-LR.

**Key words:** Microcystin-LR, silymarin, antioxidant, oxidative stress, heart, lungs, cyanobacteria

**Citation:** Ayman Al-Hazmi, 2020. Antioxidant activity of silymarin in microcystin-LR cardiac and pulmonary induced injuries on mice. Pak. J. Biol. Sci., 23: 1369-1373.

**Corresponding Author:** Ayman Al-Hazmi, Faculty of Applied Medical Sciences, Taif University, Taif City, Saudi Arabia

**Copyright:** © 2020 Ayman Al-Hazmi. This is an open access article distributed under the terms of the creative commons attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

**Competing Interest:** The authors have declared that no competing interest exists.

**Data Availability:** All relevant data are within the paper and its supporting information files.

## INTRODUCTION

Cyanobacteria is a blue-green alga that is present in different water supplies as dams, lakes and blooms<sup>1</sup>. Microcystin is a cyclic heptapeptide consisting of variable amino acids<sup>2</sup>. Microcystin-LR which contains two amino acids leucine (L) and arginine (R) is the most microcystin associated with the incident of toxicity in different countries<sup>3</sup>. Humans can be exposed to microcystin-LR mainly through drinking water contaminated with cyanobacteria. The liver is the main organ that detoxified microcystin-LR and its by-products can be seen in urine<sup>4</sup>. In the liver, microcystin-LR causes severe damage characterized by hepatocyte destruction, hepatic hemorrhage, leading to shock and death<sup>5</sup>. In Saudi Arabia, cyanobacteria were found in permanent and semipermanent rainwater ponds in Asir region<sup>6</sup>. A previous study conducted on male mice that were administered a high dose of microcystin-LR found that mice liver had chronic inflammation characterized by hepatocyte degeneration with significant elevation of serum ALT and AST, reduction in gamma-glutamyl transferase (GGT) and total protein and albumin. These injuries caused by the inhibitory effect of microcystin-LR on protein phosphatases which disturbs intracellular signaling and cytoskeleton of these tissue<sup>7</sup>. Microcystin-LR toxicity on myocytes led to damage and death<sup>8</sup>. In the same vein, a previous study conducted on mice showed that intraperitoneally injection of these mice with a toxic dose of microcystin-LR caused pulmonary thrombosis<sup>9</sup>. Moreover, two English cases were complicated with pneumonia after exposure to microcystin-LR<sup>10</sup>. Silymarin is a poorly water-soluble flavonoid that has anti-oxidant, anti-inflammatory and anti-cancer effects<sup>11</sup>. Extracts of both fruits and seeds are the main source of *Silymarin maianum*<sup>12</sup>. Silymarin induces anti-oxidant activity either through slowing calcium metabolism preventing initiation step in auto-oxidation or inhibiting lipid peroxidation by induction of superoxide dismutase and glutathione peroxidase<sup>13</sup>. In addition to its antioxidation activity, silymarin inhibits toxin absorption by the cell<sup>5</sup>. During inflammation, silymarin has an anti-inflammatory effect by the regulation of inflammatory mediators. It also inhibits neutrophils migration factor and prostaglandins synthesis<sup>6</sup>. In an animal study, silymarin protects cardiac tissue against ischemic heart diseases<sup>14</sup>. A previous study showed that treatment of animals injected with bleomycin with silymarin induced recovery of pulmonary injuries<sup>15</sup>. This study aimed to evaluate the antioxidant activity of silymarin on heart and lung injuries induced by microcystin-LR in Balb/c mice.

## MATERIALS AND METHODS

**Experimental methods:** This study was carried out in the Faculty of Applied Medical Sciences at Taif University between August 2019 to December of the same year. Seventy-two male Balb/c mice 5-7 weeks of age weighing about 30 grams were grouped into 6 groups; the first group was the control group (C), which consisted of twelve mice fed with a normal diet without any supplementation. Whereas, the second and the third groups contained twelve mice each called the microcystin-LR control group (M6 and M12). The mice in these groups were injected with microcystin-LR 19 mg kg<sup>-1</sup> b.wt. intraperitoneally. After six and twelve hours of injection, the mice in both M6 and M12 were killed. Twelve mice supplemented orally with 250 mg silymarin per kg body weight daily were classified as silymarin control group (S), then killed after ten days. The mice of the fifth and sixth groups were orally supplemented with 250 mg silymarin per kg body weight daily. Ten days later, each mouse in these groups was injected with 19 mg microcystin-LR per kg body weight intraperitoneally. Six hours later, the mice of the fifth group were killed (SM6), while the mice of the sixth group were killed after 12 h (SM12) of microcystin-LR injection. Immediately, a blood sample was collected into a plain tube from each mouse after death, then centrifuged. The serum was isolated and was stored at -20°C for ALT and AST estimation. After death, both heart and lungs were resected immediately from each mouse. In normal saline tubes containing heparin, both organs were perfused. Furthermore, it was homogenized with phosphate buffer saline with PH 7.2 using Ultra Turrax homogenizer, centrifuged at 3000 g for 30 min. Finally, the supernatant was removed and stored<sup>16</sup> at -20°C.

**Biochemical measurement:** The chemicals were purchased from (Sigma-Aldrich, USA). Estimation of biochemical parameters levels was done by using a spectrophotometer (UV-1800, Shimadzu). The serum transaminases (ALT and AST) of each mouse were measured. Moreover, reduced glutathione level and protein phosphatase I activity were estimated in cardiac and lungs homogenates. Finally, protein carbonyl content protein, products of lipid peroxidation and toxic methylglyoxal were estimated in both heart and lungs homogenates<sup>16</sup>.

**Statistical analysis:** All data were presented by using mean  $\pm$  or plus-minus standard deviation (SD). The one-way Analysis of Variance (ANOVA) was used in the comparison of each

biochemical parameter between all 6 groups. The level of significance was set at  $p < 0.05$ . SPSS software version 16 (SPSS Inc., Chicago, IL, USA) was used for statistical analysis.

## RESULTS

Data in Table 1 presented the serum ALT activity showed a highly statistical difference between six groups ( $p < 0.01$ ). The M12 had a higher ALT activity ( $1956.7 \pm 60.77$ ) compared with the control group ( $427.9 \pm 2.31$ ). Meanwhile, the SC had a lower activity ( $427.9 \pm 2.31$ ). Furthermore, a highly significant statistical difference was observed in protein phosphatase I activity between all groups ( $p < 0.01$ ). The M12 group had a higher AST activity ( $2.923 \pm 0.007$ ) compared with the control group ( $2.923 \pm 0.007$ ). The SC group had a lower activity ( $0.405 \pm 0.006$ ). Table 2 represents the PPI activity, LPO, GSH, CC and MG levels in the cardiac homogenate of 6 groups. A significant statistical difference in the PPI activity was discovered between all groups ( $p < 0.05$ ). The M12 group had a lower activity ( $3.11 \pm 0.077$ ), while the SC group had a higher activity ( $5.84 \pm 0.010$ ) compared with the control group ( $5.56 \pm 0.091$ ). In addition, a significant statistical difference appeared in the GSH level between all groups ( $p < 0.05$ ) with a higher level in the SC group ( $2.00 \pm 0.070$ ) while the lower level in the M12 group ( $0.775 \pm 0.017$ ) compared with the control group ( $1.70 \pm 0.070$ ). In the same homogenate, a significant statistical difference was found in the CC level between all

groups ( $p < 0.05$ ). The M12 group had a higher level ( $0.468 \pm 0.027$ ) while the SC had the lower level ( $0.180 \pm 0.007$ ). Moreover, the MG level showed a significant statistical difference between the six groups ( $p < 0.01$ ). The M12 group had a higher level ( $4.130 \pm 0.021$ ), while the SC group had a lower level ( $0.960 \pm 0.007$ ). The activity of PPI and the levels of GSH, LPO, CC and MG of all groups in the lung homogenate are represented in Table 3. A significant statistical difference was found in the CC level between all groups with a higher level in the M12 ( $0.590 \pm 0.009$ ) while the lower level in the SC ( $0.180 \pm 0.007$ ) compared with the control group ( $0.260 \pm 0.005$ ) ( $p < 0.05$ ). Finally, the PPI activity showed a significant statistical difference with the higher activity in the SC group ( $5.245 \pm 0.173$ ) while the lower activity in the M12 group ( $2.900 \pm 0.056$ ) compared with the control group ( $4.230 \pm 0.056$ ) ( $p < 0.05$ ).

## DISCUSSION

Cardiac diseases are still the most common cause of death all over the world. Oxidative stress plays an important role in cardiac disorders. Microcystin-LR is a toxic heptapeptide that affects different organs in the human body. It induces cell injury mainly through inhibition of serine/threonine protein phosphatase<sup>14</sup>. Microcystin-LR affects cardiomyocyte by inducing oxidative stress<sup>17</sup>. This stress either occurs due to the increasing production of reactive oxygen species (ROS) or

Table 1: Activities of Serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST) levels in different groups

Parameters	C	M6	M12	SC	SM6	SM12	p-value
ALT (U L <sup>-1</sup> )	550.5±7.41	1258.7±5.65	1956.7±60.77	427.9±2.31	742.7±29.52	885.2±31.16	0.007**
AST (U L <sup>-1</sup> )	0.405±0.006	1.750±0.006	2.923±0.007	0.391±0.006	0.521±0.005	1.980±31.16	0.009**

\*\* $p < 0.01$ , C: Control group, M6: Microcystin 6 hours group, M12: Microcystin 12 h group, SC: Silymarin control group, SM12: Silymarin microcystin 6 h group and silymarin microcystin 12 h group

Table 2: Biochemical parameters of cardiac homogenate exposed to microcystin-LR and Silymarin compared with control homogenate

Parameters	C	M6	M12	SC	SM6	SM12	p-value
PPI (U mg <sup>-1</sup> )	5.56±0.091	3.56±0.042	3.11±0.077	5.84±0.010	4.68±0.031	4.38±0.056	0.047*
LPO (uM)	420.00±0.006	444.89±1.847	490.67±0.491	389.89±1.323	434.67±0.951	469.89±2.690	0.097
GSH (nM)	1.70±0.070	1.36±0.084	0.775±0.017	2.00±0.070	1.980±0.021	1.254±0.018	0.033*
CC (nmol)	0.260±0.005	0.309±0.008	0.468±0.027	0.180±0.007	0.270±0.090	0.200±0.007	0.044*
MG (uM)	1.010±0.007	2.320±0.014	4.130±0.021	0.960±0.014	1.350±0.014	1.870±0.049	0.008**

\* $p < 0.05$ , \*\* $p < 0.01$ , C: Control group, M6: Microcystin 6 h group, M12: Microcystin 12 h group, SC: Silymarin control group, SM12: Silymarin microcystin 6 h group and silymarin microcystin 12 h group

Table 3: Biochemical parameters of lung homogenate exposed to microcystin-LR and Silymarin compared with control homogenate

Parameters	C	M6	M12	SC	SM6	SM12	p-value
PP1 (U mg <sup>-1</sup> )	4.230±0.056	3.060±0.042	2.900±0.056	5.245±0.173	5.134±0.188	5.080±0.029	0.049*
LPO (uM)	420.00±0.001	444.89±0.003	490.67±0.001	389.89±0.001	434.67±0.001	469.89±0.001	0.084
GSH (nM)	1.100±0.141	1.010±0.035	0.987±0.106	1.170±0.106	1.101±0.091	1.000±0.070	0.092
CC (nmol)	0.370±0.004	0.540±0.002	0.590±0.009	0.200±0.007	0.290±0.005	0.380±0.010	0.041*
MG (uM)	1.110±0.063	1.200±0.007	1.210±0.005	1.100±0.070	1.150±0.035	1.170±0.014	0.138

\* $p < 0.05$ , C: Control group, M6: Microcystin 6 hours group, M12: Microcystin 12 h group, SC: Silymarin control group, SM12: Silymarin microcystin 6 h group and silymarin microcystin 12 h group

decreasing antioxidant production<sup>18</sup>. The increase of ROS is leading to the peroxidation of membrane lipids mainly polyunsaturated fatty acids<sup>19</sup>. In addition, microcystin-LR increases the level of toxic methylglyoxal by inhibiting glyoxalase I<sup>20</sup>. Previously, a histological study conducted on mice showed that long exposure of these mice to microcystin-LR induced enlargement of cardiomyocytes, loss of cells cross-striations, fibrosis and immune cells infiltration<sup>21</sup>. This study was carried out before on male Balb/c mice showed that injecting these mice with microcystin-LR induced methylglyoxal while it decreased reduced glutathione in the cardiac homogenate. Moreover, serum AST activity increased in these mice compared with those of the control group. Certainly, AST activity resulted from either hepatocyte or cardiomyocyte destruction. Silymarin is a polyphenolic flavonoid, isolated from *Silybum marianum*<sup>22</sup>. It performed a therapeutic effect in different tissues including cardiac, lungs and liver through its antioxidant activity<sup>23</sup>. Silymarin antioxidant activity was accomplished by its action as a scavenger of free radical, besides, it increased reduced glutathione level<sup>24</sup>. The results showed that silymarin dose decreased infarct size, lipid peroxide and transaminases level, however, it induced the activity of glutathione transferase and catalase<sup>25</sup>. In this study, silymarin decreased serum activities of AST and ALT nearly by two-fold in mice injected with microcystin-LR. The lipid peroxidation products level as a result of oxidative degradation of polyunsaturated fatty acids in cell membrane were reduced after silymarin administration<sup>26</sup>. Moreover, it induced the level of reduced glutathione which improved antioxidant activity against oxidative stress resulting from microcystin-LR toxic effect. In addition, protein carbonyl content and methylglyoxal resulting from microcystin-LR toxicity were reduced after the administration of silymarin. Also, it was found that silymarin induced protein phosphatase I activity involved in cell growth and differentiation<sup>26</sup>. Nearly, results of the present study may represent the protective effect of silymarin for cardiomyocytes either directly through induction of reduced glutathione level or indirectly through induction of protein phosphatase I involved in the growth of cardiomyocytes. As for lung tissue, a previous study found that microcystin-LR induced pulmonary injuries characterized by alveolar collapse, cell apoptosis, polymorphonuclear cell infiltration and inflammatory changes in rat. These changes were followed by the damaged lung tissues and impairment in its function<sup>26</sup>. In this study, intraperitoneal injection of Male Balb/c mice with microcystin-LR induced protein carbonyl content, however, it reduced protein phosphatase activity. Silymarin had a therapeutic effect on many pulmonary diseases including

small cell carcinoma and lung ischemic diseases<sup>26</sup>. It played a significant role against pulmonary disorders either by its anti-inflammatory or antioxidant activities. Natural factor kaba B (NF-kB) induced inflammatory reaction in the lung by induction of chemokines, expression of pro-inflammatory cytokines and immune cells infiltration<sup>27</sup>. Silymarin acts as anti-inflammatory cytokine by inhibiting the inflammatory process through inhibition of NF-kB, tumor necrosis<sup>28</sup> factor- $\alpha$  (TNF- $\alpha$ ) and IL-6. Silymarin maintains a normal oxidant level by inducing the synthesis of superoxide dismutase catalase and precursor of reduced glutathione<sup>29</sup>. In this study, silymarin reduced both lipid peroxidation products and protein carbonyl content in lung homogenate resulting from lung oxidative stress and tissue damage. Moreover, it induced protein phosphatase activity responsible for lung tissue repair and cure. On the other hand, silymarin induced the reduced glutathione level responsible for antioxidant activity.

## CONCLUSION

In conclusion, silymarin obtained a therapeutic role in cardiac and pulmonary injuries through its antioxidant property that may be resulted by the induction of reduced glutathione level or activation of protein phosphatase I. The discovery of an active substance in silymarin that is responsible for its antioxidant activity is recommended.

## ACKNOWLEDGMENT

The author would like to thank all the staff at the animal house of the Deanship of Scientific Research, Taif University, SA for helping their help in this project.

## SIGNIFICANCE STATEMENT

This study discovered the antioxidant activity of silymarin that can be beneficial for using this nontoxic compound to reduce the oxidative stress produced in some chronic diseases. This study will help the researchers to uncover the critical areas of medicinal plants that many researchers were not able to explore. Thus, a new theory on medicinal plants may be arrived at.

## REFERENCES

1. Dixon, M.B., Y. Richard, L. Ho, C.W.K. Chow, B.K. O'Neill and G. Newcombe, 2011. A coagulation-powdered activated carbon-ultrafiltration-Multiple barrier approach for removing toxins from two Australian cyanobacterial blooms. *J. Hazard. Mater.*, 186: 1553-1559.

2. Falconer, I.R., A.R. Humpage, 2006. Cyanobacterial (blue-green algal) toxins in water supplies: Cylindrospermopsins. *Environ. Toxicol.*, 21: 299-304.
3. Brooke, S., G. Newcombe, B. Nicholson and G. Klass, 2006. Decrease in toxicity of microcystins LA and LR in drinking water by ozonation. *Toxicol.*, 48: 1054-1059.
4. Ortelli, D., P. Edder, E. Cognard and P. Jan, 2008. Fast screening and quantitation of microcystins in microalgae dietary supplement products and water by liquid chromatography coupled to time of flight mass spectrometry. *Analytica Chimica Acta*, 617: 230-237.
5. Mohamed, Z.A., 2008. Toxic cyanobacteria and cyanotoxins in public hot springs in Saudi Arabia. *Toxicol.*, 51: 17-27.
6. Zakaria A. Mohamed, Z.A and A.M. Al Shehri, 2009. Microcystins in groundwater wells and their accumulation in vegetable plants irrigated with contaminated waters in Saudi Arabia. *J. Hazard. Mater.*, 172: 310-315.
7. Trinkle-Mulcahy, L. and A.I. Lamond, 2006. Mitotic phosphatases: no longer silent partners. *Curr. Opin. Cell Biol.*, 18: 623-631.
8. Jin, Y., X. Zhao, H. Zhang, Q. Li, G. Lu and X. Zhao, 2016. Modulatory effect of silymarin on pulmonary vascular dysfunction through HIF-1 $\alpha$ -iNOS following rat lung ischemia-reperfusion injury. *Exp. Ther. Med.*, 12: 1135-1140.
9. Slatkin, D., R. Stoner, W. Adams, J. Kycia and H. Siegelman, 2006. Atypical pulmonary thrombosis caused by a toxic cyanobacterial peptide. *Sci.*, 220: 1383-1385.
10. Turner, P.C., A.J. Gammie, K. Hollinrake and G.A. Codd, 1990. Pneumonia associated with contact with cyanobacteria. *Br. Med. J.*, 300: 1440-1441.
11. Frascini, F., G. Demartini and D. Espostii, 2002. Pharmacology of silymarin. *Clin. Drug Invest.*, 22: 51-65.
12. Karimi, G., M. Vahabzadeh, P. Lari, M. Rashedinia and M. Moshiri, 2011. "Silymarin", a promising pharmacological agent for treatment of diseases. *Iran. J. Basic Med. Sci.*, 14: 308-317.
13. Kwon, D.Y., Y.S. Jung, S.J. Kim, Y.S. Kim, D.W. Choi and Y.C. Kim, 2013. Alterations in sulfur amino acid metabolism in mice treated with silymarin: A novel mechanism of its action involved in enhancement of the antioxidant defense in liver. *Planta. Med.*, 79: 997-1002.
14. Zholobenko, A. and M. Modriansky, 2014. Silymarin and its constituents in cardiac preconditioning. *Fitoterapia*, 97: 122-132.
15. Razavi-Azarkhiavi, K., M. Ali-Omrani, R. Solgi, P. Bagheri, M. Haji-Noormohammadi, N. Amani and M.R. Sepand, 2014. Silymarin alleviates bleomycin-induced pulmonary toxicity and lipid peroxidation in mice. *Pharm. Biol.*, 52: 1267-1271.
16. Mountfort, D.O., P. Holland and J. Sprosen, 2005. Method for detecting classes of microcystins by combination of protein phosphatase inhibition assay and ELISA: comparison with LC-MS. *Toxicol.*, 45: 199-206.
17. Gupta, O.P., S. Sing, N. Bani, B.D. Malhotra, S.K. Gupta and B. Handa, 2000. Anti-inflammatory and anti-arthritis activities of silymarin acting through inhibition of 5-lipoxygenase. *Phytomed.*, 7: 21-24.
18. Jayaraj, R., T. Anand and P.V.L. Rao, 2006. Activity and gene expression profile of certain antioxidant enzymes to microcystin-LR induced oxidative stress in mice. *Toxicol.*, 220: 136-146.
19. Joshi, G., R. Sultana, J. Tangpong, M.P. Cole and Daretre *et al.*, 2005. Free radical mediated oxidative stress and toxic side effects in brain induced by the anti cancer drug adriamycin: Insight into chemobrain. *Free Radical Res.*, 39: 1147-1154.
20. Kalapos, M.P., 2008. The tandem of free radicals and methylglyoxal. *Chem. Biol. Interact.*, 171: 251-271.
21. Lankoff, A., W.W. Carmichael, K.A. Grasman and M. Yuan, 2004. The uptake kinetics and immunotoxic effects of microcystin-LR in human and chicken peripheral blood lymphocytes *in vitro*. *Toxicol.*, 204: 23-40.
22. Di Meo, F., V. Lemaury, J. Cornil, R. Lazzaroni, J.L. Duroux, Y. Olivier and P. Trouillas, 2013. Free radical scavenging by natural polyphenols: atom versus electron transfer. *J. Phys. Chem. A*, 117: 2082-2092.
23. Morishima, C., M.C. Shuhart, C.C. Wang, D.M. Paschal and M.C. Apodaca *et al.*, 2010. Silymarin inhibits *in vitro* T-cell proliferation and cytokine production in hepatitis C virus infection. *Gastroenterol.*, 138: 671-681.
24. Gehringer, M.M., E.G. Shephard, T.G. Downing, C. Wiegand and B.A. Neilan, 2004. An investigation into the detoxification of microcystin-LR by the glutathione pathway in Balb/c mice. *Int. J. Biochem. Cell Biol.*, 36: 931-941.
25. Di Meo, F., V. Lemaury, J. Cornil, R. Lazzaroni, J.L. Duroux, Y. Olivier and P. Trouillas, 2013. Free radical scavenging by natural polyphenols: atom versus electron transfer. *J. Phys. Chem. A*, 117: 2082-2092.
26. Polyak, S.J., C. Morishima, M.C. Shuhart, C.C. Wang, Y. Liu and D.Y.W. Lee, 2007. Inhibition of T-cell inflammatory cytokines, hepatocyte NF- $\kappa$ B signaling and HCV infection by standardized silymarin. *Gastroenterology*, 132: 1925-1936.
27. Gharagozloo, M., E. Velardi, S. Bruscoli, M. Agostini and M.D. Sante *et al.*, 2010. Silymarin suppress CD4+ T cell activation and proliferation: Effects on NF- $\kappa$ B activity and IL-2 production. *Pharm. Res.*, 61: 405-409.
28. Kim, B.R., H.S. Seo, J.M. Ku, G.J. Kim and C.Y. Jeon *et al.*, 2013. Silibinin inhibits the production of pro-inflammatory cytokines through inhibition of NF- $\kappa$ B signaling pathway in HMC-1 human mast cells. *Inflamm. Res.*, 62: 941-950.
29. Clichici, S., D. Olteanu, A.L. Nagy, A. Oros, A. Filip and P.A. Mircea, 2015. Silymarin inhibits the progression of fibrosis in the early stages of liver injury in CCl<sub>4</sub>-treated rats. *J. Med. Food*, 18: 290-298.