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Research Article Effect of Transplanted Bone Marrow on Kidney Tissue of γ-Irradiated Pregnant Rats and Their Fetuses

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Abstract

Background and Objectives: The damaging effects of ionizing radiation lead to cell death. The present study was performed to assess the possible ameliorating effects of bone marrow transplantation (BMT) on the histopathological and histochemical changes in the kidney tissue of γ -irradiated pregnant rats and their fetuses. **Materials and Methods:** Pregnant rats were divided into 5 sets (6 females in each set): Group C (untreated pregnant rats), group R7 (pregnant rats exposed to 2Gy of γ -rays on the 7th day of pregnancy), group R7+BM (pregnant rats exposed to 2Gy of γ -rays on the 7th day of pregnancy) then injected by freshly BMT (75 × 10⁶±5 cells) intra peritoneally after 1 h of irradiation, group R14 (pregnant rats exposed to 2Gy of γ -rays on the 14th day of pregnancy), group R14+BM (pregnant rats exposed to 2Gy γ -rays on the 14th day of pregnancy and after 1 h received 1 dose of BMT). All pregnant rats were sacrificed on the 20th day of pregnancy and kidney samples of pregnant rats and their fetuses were removed for histopathological and histochemical studies. **Results:** Gamma rays caused many histological and histochemical deviations in the kidney tissue of mothers and their fetuses on day 7 or 14 of gestation, but bone marrow transplantation highly improved the damage were occurred due to γ -rays. **Conclusion:** Bone marrow transplantation has the ability to decrease the injury of gamma rays.

Key words: lonizing radiation, bone marrow transplantation, gamma irradiated rats, gestation

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Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Prenatal exposure to ionizing radiation can overlap with embryonic and fetal development, depending on dose and gestational age. Radiation is usually explained as the emission and diffusion of energy in the form of waves or particles through space or substance¹. It interrelates with matter by straight and indirect methods. Both methods cause molecular injury and that translated to biological destruction². Exposure to ionizing radiation whether working or during radiotherapy cause severe systemic damage to numerous cellular and sub cellular structures³. Ionizing radiation forms radicals in the DNA and in the adjacent water particles of the hydration shell of the DNA which destroy DNA⁴. High doses of ionizing radiation may be deadly to the cell by damaging both DNA strands⁵. The occurrence of injured nuclear DNA in cells can cause genomic instability and numerous cell divisions⁶.

lonizing radiation embodied a possible teratogen for the fetus, but this risk has been found to be reliant on the dose and the belongings creatable to the gestation time at exposure⁷. Emergent mammalian embryo is more sensitive to ionizing radiation than adult. Radiation persuaded deviations in mammals are closely connected to the period of growth of embryos at which radiation is assumed⁸. Gama rays (2Gy) caused embryonic loss and deformities in mice⁹.

Mutagenic belongings and organ-specific changes were induced by radiation exposure¹⁰. Kidney is the greater influential route for the absorption of adventurous materials conflict in the environment. The histopathological deviations reported due to radiation exposure were linked to a decline in kidney functions¹¹. Cloudy cellular swelling, congestion and thick cellular penetration were occurred by gamma radiation on mice kidney tissue¹². The kidney of gravid rats exposed at the dose level of 3Gy on days 7, 13 or 18 of pregnancy showed degeneration and provocative penetration within many convoluted tubules¹³.

Bone marrow is a complex tissue consists of 2 sections, haematopoietic and stromal one. The stromal section is the structural basis of the haematopoietic microenvironment which is a complex tissue that contains a subset of cells called mesenchymal stem cells (MSCs)¹⁴. The MSCs maintain stem cell features, are used to recover the healing of hurt tissues. It are also used to indulgence dysfunction of other tissues, such as tissues of the nervous system, the cardiac system and lunate bone¹⁵. Bone marrow transplantation boosts the antioxidant level and defends from oxidative stress in irradiated rats¹⁶. Bone marrow derived mesenchymal stem cells (BMSCs) have shown great capacity for ischemic tissue reparation^{17,18}. Human bone marrow-derived mesenchymal

stem cell transplantation may have clinical requests for the treatment and repair of tissue injury made by radiation¹⁰. The present study was done to assess the possible ameliorating effects of BMT on the histopathological and histochemical changes in the kidney tissue of γ -irradiated pregnant rats and their fetuses.

MATERIAL AND METHODS

Experimental animals: Mature albino rats (*Rattus albinus*), their body weight from 120-150 g were kept in cages. The males were separated from females up to reproducing. One male were caged with two females of proestrus or estrous periods. In the next morning the gestation was assured by the occurrence of vaginal plug or presence of sperms in smears of vaginal content and that day was considered the 1st day of gestation.

Study area: The present study was performed in National Center for Radiation Research and Technology, Atomic Energy Authority, Cairo, Egypt from January to March, 2013.

Irradiation: Irradiation was done in the National Center for Radiation Research and Technology, Atomic Energy Authority, Cairo, Egypt by Gamma-cell 40 (137 Cesium).

Bone marrow transplantation: Donors and recipients of bone marrow transplantation were selected of the similar strain. The donors were sacrificed and femur bones were cleaned and both ends were fragmented by bone nibbling forceps. The marrow was driven of the femur into saline solution under sterilized conditions bounded by ice cubes and mixed by drawing and expelling it several times from the syringe without needle in order to avoid mechanical damage to the cells. Total viable of cells about $75 \times 10^6 \pm 5$ were injected one hour post irradiation¹⁹.

Experimental groups: The pregnant rats were separated into 5 groups (6 females in each group), Group C (untreated pregnant rats), group R7 (Pregnant rats exposed to 2Gy of gamma rays on the 7th day of pregnancy), group R7+BM (pregnant rats exposed to 2Gy of gamma rays on the 7th day of pregnancy and received freshly BMT ($75 \times 10^6 \pm 5$ cells) by intra peritoneal admission 1 h post-irradiation, group of R14 (pregnant rats insecure to 2Gy of gamma rays on the 14th day of pregnancy), group of R14+BM (pregnant rats exposed to 2Gy gamma rays on the 14th day of pregnancy), group of R14+BM (pregnant rats exposed to 2Gy gamma rays on the 14th day of pregnancy and after 1 h received one dose of BMT. All pregnant rats were sacrificed on the 20th day of pregnancy.

Histological and histochemical studies: After sacrifation the pregnant animals on the 20th day of pregnancy, small pieces of kidney tissues of mothers and their fetuses were quickly removed and fixed in 10% neutral buffer formol and Carnoy's fluid for histological and histochemical studies. Sections were stained by haematoxylin and eosin stain agreeing to the technique of Carleton *et al.*²⁰, collagen fibers were demonstrated by Mallory's trichrome stain²¹, polysaccharides were demonstrated by periodic acid Schiff's technique²¹, total protein were detected by mercuric bromophenol blue²².

Statical analysis: The data analyzed using one way analysis of variance followed by Duncan's test. Differences between groups were considered significant at $p \le 0.05$.

RESULTS

Histopathological and histochemical studies in the kidney tissue of pregnant rats

Histopathological results: Control kidney tissue consists of Malpighian s corpuscles with Bowman's capsules, Bowman's spaces, glomeruli, distal and proximal convoluted tubules (Fig. 1a). Kidney of pregnant rats of group R7 showed faintly stained cells and nuclei of the convoluted tubules, some nuclei appeared deeply stained (pyknotic), wide lumens in the distal convoluted tubules, ruptured brush borders of the proximal convoluted tubules, haemolysed RBCs in the glomeruli, hemorrhagic areas (Fig. 1b). Bone marrow treatment improved the kidney architecture in the pregnant rats of groups R7+BM (Fig. 1c). Highly distorted malpighian capsules, most nuclei of the proximal convoluted tubules cells were hypertrophied and pyknotic, others were karyolytic, large and small degenerated areas, atrophied glomeruli with debris of degenerated tubules were noted in R14 group (Fig. 1d, e). Improved the kidney architecture were noted in R14+BM (Fig. 1f).

Normal distribution of collagen fibers in the kidney tissue of control group (Fig. 2a). Thin collagen fibers are supporting walls of the blood vessels, distal convoluted tubules, proximal convoluted tubules, blood vessels and Bowman's capsules in kidney tissue of group R7 (Fig. 2b). A slight increase in the collagen fibers in walls of the distal and proximal convoluted tubules and in between and around Bowman's capsules in group R7+BM (Fig. 2c). Increased collagen fibers in the distal and proximal convoluted tubules and some glomeruli, but the remnant glomeruli are poorly stained were noted in R14 group (Fig. 2d). Increased collagen fibers in between and around Bowman's capsules were found in R14+BM group (Fig. 2e).

Histochemical results: Table 1 showed reduced mean optical transparency (MOT) of polysaccharides in the atrophied glomeruli and tubules of the kidney of groups R7 and R14 compared to the control group. The percentage of change in MOT was clearly approach to the normal values in the kidney tissue of the pregnant rats of groups R7+BM and R14+BM. Significant decrease in MOT value of total protein content in kidney tissue of pregnant rats in groups R7 and R14 were noted compared to the control group. Somewhat normal MOT of total protein was noticed in the glomeruli, distal and proximal convoluted tubules in groups R7+BM and R14+BM.

Histological and histochemical results of fetuses

Histological results: Normal fetal kidney tissue were noted in the control group (Fig. 3a). Radiation exposure on day 7 of gestation caused many dystrophic changes in the fetal kidney tissue maternally exposed to 2Gy γ -rays. These changes include: Degenerated convoluted tubules, some glomeruli are atrophied with dilated Bowman's spaces, numerous hemorrhagic areas, deeply stained pyknotic nuclei with debris of degenerated cells of the convoluted tubules (Fig. 3b). Bone marrow transplantation post-irradiation (2Gy) on day 7 of gestation showed normal appearance of the fetal kidney tissue (Fig. 3c). Highly distorted and degenerated cells of distal and proximal convoluted tubules which are surrounded by numerous fibrotic and hemorrhagic areas, glomeruli are atrophied, lobulated or hypertrophied with numerous pyknotic nuclei were noted in R14 group (Fig. 3d, e). Normal appearance of kidney tissue were found in R14+BM group (Fig. 3f).

Table 1: MOT and the percentage of change values of PAS+ve materials and total protein in the glomeruli and tubules of the pregnant rats

Groups	MOT values of PAS+ve materials				MOT values of total protein			
	Glomeruli Mean±SE	Change (%)	Tubules Mean±SE	Change (%)	Glomeruli Mean±SE	Change (%)	Tubules Mean±SE	Change (%)
Control	114.00±1.48		91.48±3.12		155.00±3.83		145.38±3.86	
R7	80.48±5.37*	29.40	74.03±3.25*	36.00	105.48±5.23*	31.95	102.33±3.12*	29.60
R7+BM	92.69±4.48*	18.70	54.33±2.95*	19.07	152.34±6.78	2.02	143.45±2.32	1.32
R14	89.00±3.53*	21.92	70.82±6.00*	22.58	110.89±3.36*	28.46	104.84±2.37*	27.88
R14+BM	91.89±6.02*	19.39	83.56±3.14*	8.65	147.89±3.57	4.58	138.78±3.74	4.53

*Values are considered significantly different (p<0.05), SE: Standard error 3.2



Fig. 1(a-f): Sections of kidney tissue of control and treated pregnant rats stained with Hx and E ×100, (a) Control group, malpighian corpuscles contain glomeruli (G), Bowman's capsules (Bc), Bowman's spaces (Bs), distal convoluted tubules (ds) and proximal convoluted tubules (pc), (b) R7 group: Faintly stained cells and nuclei of the proximal and distal convoluted tubules, some nuclei appear deeply stained (>), wide lumens in the distal convoluted tubules, ruptured brush borders of the proximal convoluted tubules, haemolysed RBCs can be detected in the glomeruli and hemorrhagic areas (1), (c) R7+BM group: Well developed kidney architecture, (d, e) R14 group: Highly distorted malpighian corpuscles, distal and proximal convoluted tubules and most of their nuclei were hypertrophied and pyknotic (p), others are karyolytic (k), large and small degenerated areas (de), wide lumens of distal convoluted tubules (dct), ruptured brush borders of proximal convoluted tubules (pct) with debris of degenerated tubules (de) and atrophied glomeruli (G) and (f) R14+BM group: Kidney tissue restore its normal architecture, but some pyknotic nuclei (p) are still detected

Normal distribution of collagen fibers in walls of the convoluted tubules, glomeruli and stroma of the fetal control kidney cortex was observed in Fig. 4a. Highly increased collagen fibers were detected in the inner cortical part, but fibrosis and hemorrhagic areas in the outer part of the cortex acquired red coloration which indicating fibrosis with absence



Fig. 2(a-e): Distribution of collagen fibers in the kidney tissue of control and treated pregnant rats stained with Mallory s trichrome stain (×100), (a) Control group: Thin collagen fibers supporting walls of the blood vessels, distal and proximal convoluted tubules, blood vessels and Bowman's capsules, (b) R7 group: Highly increased collagen fibers in walls of distal and proximal convoluted tubules and the brush borders of proximal convoluted tubules, (c) R7+BM group: A slight increase in the collagen fibers in walls of distal and proximal convoluted tubules and Bowman's capsules, (d) R14 group: Increased collagen fibers in the distal and proximal convoluted tubules and some glomeruli, but the remnant glomeruli are poorly stained (1) and (e) R14+BM group: Increased collagen fibers in between and around Bowman's capsules

of kidney medulla in the fetal kidney tissue of group R7 (Fig. 4b). Deeply stained collagen fibers and signs of fibrosis were still detected in the fetal kidney tissue of group R7+BM (Fig. 4c). The fetal kidney tissue of group R14 showed

decreased collagen fibers in the convoluted tubules and most glomeruli (Fig. 4d). Somewhat normal appearance of collagen fibers was detected in the fetal kidney tissue of group R14+BM (Fig. 4e).



Fig. 3(a-f): Sections of kidney tissue of fetuses of the control and treated pregnant rats stained with Hx and E, (a) Control group: Kidney tissue which contains glomeruli (G), proximal convoluted tubules with their brush borders (pc) and distal convoluted tubules with their wide lumens (dc) (×100), (b) R7 group: Degenerated convoluted tubules, some glomeruli are atrophied with dilated Bowman's spaces (1), numerous hemorrhagic areas (h), deeply stained pyknotic nuclei (1) with debris of degenerated cells of the convoluted tubules (>) (×100), (c) R7+BM group: Normal kidney tissue (×50), (d, e) R14 group: Highly distorted and degenerated cells of distal and proximal convoluted tubules which are surrounded by numerous fibrotic and hemorrhagic areas, glomeruli are atrophied (1), lobulated (1) or hypertrophied (>) with numerous pyknotic nuclei (p) and (f) R14+BM group: Normal appearance of kidney tissue

Histochemical results: Table 2 showed MOT values of PAS+ve materials in the fetal kidney tissue. The glomeruli and tubules of fetal kidney tissue were poorly stained in groups R7 and R14 compared to the control group. Somewhat normal MOT values of PAS+ve materials were detected in group R7+BM. Also in group R14+BM, some of distal and proximal convoluted tubules and glomeruli restored their normal content of polysaccharides. Also,

reduced MOT value of total protein were noted in the glomeruli and distal and proximal convoluted tubules in the fetal kidney of groups R7 and R14 compared to the control group. Bone marrow transplantation improved MOT value of total protein in the glomeruli and proximal convoluted tubules with less stain affinity in distal convoluted tubules was detected in the fetal kidney tissue of group R7+BM and R14+BM.



Fig. 4(a-e): Distribution of collagen fibers in the kidney tissue of fetuses of the control and treated pregnant rats stained with mallory s trichrome stain, (a) Control group: Normal distribution of collagen fibers in walls of the convoluted tubules, glomeruli and stroma of the kidney cortex (×50), (b) R7 group: Dense stain affinity of collagen fibers in the inner cortical part, but fibrotic and hemorrhagic areas in the outer part of the cortex acquire red coloration with absence of kidney medulla (×100), (c) R7+BM group: Deeply stained collagen fibers in the kidney cortex and signs of fibrosis are still detected (×100), (d) R14 group: Decreased collagen fibers in the convoluted tubules and most glomeruli (×100) and (e) R14+BM group: Somewhat normal appearance of collagen fibers, but signs of fibrosis were still detected (×50)

Table 2. MOT and the pe	preantage of change values of DA	S±vo matorials and total i	protoin in the fotal.	alomoruli and tubulor
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	MOT values of PAS+ve materials				MOT and percentage of change average values of total protein			
	Glomeruli	Tubules	Glomeruli	Tubules	Glomeruli	Tubules	Glomeruli	Tubules
Groups	Mean±SE	change (%)	Mean±SE	change (%)	Mean±SE	change (%)	Mean±SE	change (%)
Control	126.26±4.41		60.51±5.86		143.03±4.57		125.82±6.88	
R7	99.28±3.00*	21.36	45.32±6.78*	25.10	121.21±6.14*	15.25	75.31±6.37*	40.14
R7+BM	124.64±6.24	1.28	58.87±5.03	2.71	138.34±4.15	3.27	119.65±5.68	4.90
R14	93.18±3.03*	23.82	43.56±6.03*	28.01	113.28±4.15*	20.76	80.92±7.60*	35.69
R14+BM	122.26±7.40	4.00	56.34±6.03	6.88	135.87±6.52	5.00	115.53±s	8.17

*Values are considered significantly different (p<0.05), SE: Standard error

DISCUSSION

lonizing radiation caused many drastic alternations in the kidney tissue of pregnant rats and their fetuses. Gamma rays act either direct or indirect effect to produce biochemical injuries²³. Low-dose of ionizing radiation may induce specific transcriptional responses in human keratinocytes²⁴.

lonizing radiation induced late disruption of the genome in the progenies of living cells and that is called radiation induced genomic instability, is established by delayed induction of radiation effects, such as chromosomal deviations and mutation and cell loss²⁵. It induces oxidative stress and generation of reactive oxygen species which lead to membrane destruction and that allow the entry of extra calcium into cells with sequential biochemical and micro functional cellular degranulation and necrosis²⁶.

Free radicals formed due to irradiation can cause a diversity of membrane changes such as lipid peroxidation, hydrolysis of phospholipids head groups, lipid-lipid crosslinks, disulfide bridge formation and amino acid rest hurt in membrane proteins and lipid protein crosslinks^{27,28}. Radiation may be cancer-causing for a wide variety of tumors such as thyroid, breast and bone marrow cancers^{29,30}.

Gravidity should is a very important period which female go through several physical, hormonal and psychic alternations and that effect on embryos, this effect varies according to the ontogenetic stage and pregnancy stage and animal species³¹⁻³³. In the present work kidney of the pregnant of groups R7 and R14 displayed many severe changes. These changes contain: Faintly stained and hypertrophied cells and nuclei of the convoluted tubules, some nuclei were deeply stained, ruptured brush borders of the proximal convoluted tubules, numerous haemolysed RBCs in the glomeruli, hemorrhagic areas, highly distorted malpighian capsules, large and small degenerated areas, atrophied glomeruli with debris of degenerated tubules. These results agree with those of Hussein¹³, Augustine et al.³⁴. Morphologic studies of radiation nephropathy have found signs of damage to blood vessels, glomeruli, tubular epithelium and interstitium³⁵. These deviations may be due to degradation of tissue lipids as a result of lipid peroxidation²⁸. Vacuoles formed in animal cells due to the breakdown of lipoprotein centers in the affected cells or may be destroy mitochondria, golgi apparatus and lysosomes³⁶.

In this study bone marrow treatment improved the kidney architecture. Signs of improvement in several tissues of gravid rats exposed to diverse doses of gamma rays then treated with bone marrow were noticed by many authors³⁷⁻³⁹. Bone marrow mesenchymal stem cells decrease radiation-made artery hurt by suppressing oxidative stress and inflammation⁴⁰. Highly increased collagen fibers were noticed in the kidney tissue of the pregnant rats of groups R7 and R14. There is an indication that gamma rays injuries bone tissue via free radical attack on the collagen⁴¹. Increased collagen fibers post-irradiation in the various tissues were noticed by many authors⁴²⁻⁴⁴.

A slight increase in collagen fibers was noticed in the kidney tissue of the pregnant rats after bone marrow transplantation. These outcomes agree with those of Abdel Naby⁴⁵ and Emam *et al.*⁴⁶. Bone marrow-derived mesenchymal stem cells (BMSCs) have shown great promise for ischemic tissue repair⁴⁷.

Reduced polysaccharides in the atrophied glomeruli, distal and proximal convoluted tubules of kidney of mothers of groups R7 and R14. Glomeruli and brush borders of proximal convoluted tubules of the kidney cortex of irradiated pregnant rats showed increased stain affinity of PAS+ve materials reaction⁴⁸. Also, decreased total protein in the glomeruli, Malpighian's corpuscles, walls of the convoluted tubules with negatively stained degenerated areas were noticed in the kidney cortex of mothers of R7 and R14, but treatment with bone marrow post-irradiation showed normal appearance of total protein content. Increase in stain affinity of total protein may be due to increased RBCs, increased collagen fibers or presence of fibrous tissue, but reduced stain affinity of total protein may be due to injured protein molecules by irradiation or reduced ability of tissue to produce proteins⁴⁹. Histological examination showed a significant lessening of tissue damage with less inflammatory and apoptotic cells in bone marrow mesenchymal stromal cells treated animals compared to the control group⁵⁰.

Radiation exposure of mothers on day 7 or 14 of gestation caused many dystrophic changes in the fetal kidney. The fetal kidney tissue is more sensitive to γ -rays than the mothers. This sensitivity was conversed by many authors^{51,52}. Highly affected glomeruli, distal and proximal convoluted tubules were more marked in embryos than their mothers⁵³.

Normal appearance of the fetal kidney tissue were noted in groups R7+BM and R14+BM. Bone marrow transplantation in irradiated pregnant rats has improved the developing fetus and it s placenta after radiation^{38,46}.

Results of the present study showed highly increased collagen fibers in fetal kidney. Decreased production of collagenolytic enzymes might contribute to additional accretion of collagen⁵⁴. The progress observed in the fetal ileum tissue their mothers exposed to γ -rays and treated with bone marrow that may be due to the ability of bone marrow cells to differentiate to mature non-haematopoietic cells of multiple tissues^{55,56}.

In the present study, fetal kidney tissue of groups R7 and R14 indicated poorly stained polysaccharides. Highly reduced polysaccharides in fetal kidney cortex post-irradiation may be due to the degeneration and vacuolation detected in most cells of proximal and distal convoluted and renal Malpighian corpuscles or may be due to failure of Golgi apparatus to produce polysaccharides⁴⁸. Normal distribution of polysaccharides were noted in groups of R7+BM and R14+BM. Bone marrow derived stem cells have cellular plasticity which provoked many detectives to use of these cells in the renewal of nonhematopoietic tissues⁵⁷.

The present study reduced stain affinity of total protein in the distal and proximal convoluted tubules and glomeruli was detected in the fetal kidney cortex of groups R7 and R14. Decreased protein in the fetal and maternal distal and proximal convoluted tubules and Malpighian corpuscles were also detected by Hanley and Knutson⁵⁸.

Normal distribution of total protein was observed in the glomeruli and proximal convoluted tubules with less stain affinity in the distal convoluted tubules of fetal kidney tissue of group R7+BM with normal distribution of total protein in the fetal kidney tissue of group R14+BM. Bone marrow transplantation is an effective strategy to decrease the harmful effects of total body irradiation in the colitis treatments⁵⁹. Stem cells can be transplanted to replace non-functional stem cells in tissues to accelerate tissue therapeutic and return the original function⁶⁰. The regenerative possibility of stem cells was studied by many authors⁶¹⁻⁶³. This study included the rats during pregnancy, effect of irradiation on their fetuses and possible protective role of bone marrow transplantation to modify radiation injury during pregnancy. The study recommended that the pregnant mothers should avoid exposure to any type of radiation and numerous experiments should be hold to use of stem cells in radiation prevention and treatment of certain disease.

CONCLUSION

According to the present results gamma irradiation (2Gy) during day 7 and 14 of pregnancy caused many histopathological and histochemical changes in the kidney tissue of pregnant rats and their fetuses. Bone marrow transplantation post irradiation could decrease radiation injury in the kidney tissue of pregnant rats and their fetuses.

SIGNIFICANCE STATEMENT

Results from this present study indicated that bone marrow transplantation has the ability to decrease the injury of gamma rays. This study will help the researcher to uncover the critical role of bone marrow transplantation toward reducing the risk of radiation hazards during pregnancy.

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REFERENCES

- Abdel Hafez, H., 2008. Effect of cabbage *Brassica oleraceae* (var. capitata) seed extract and gamma rays on rat testis. M.Sc. Thesis, Faculty of Science, Al-Azhar University, Egypt.
- Pinon-Lataillade, G., M.C. Viguier-Martinez, A.M. Touzalin, J. Maas and B. Jegou, 1991. Effect of an acute exposure of rat testes to gamma rays on germ cells and on Sertoli and Leydig cell functions. Reprod. Nutr. Dev., 31: 617-629.
- Salama, S.F., O.M. Ashry and E.M. Hussein, 2007. Concomitant effect of ciprofloxacin and echinacea counteracting severity of radiation damage in rats. Egypt. J. Radiat. Sci. Applic., 20: 365-383.
- Kopjar, N., S. Miocic, S. Ramic, M. Millic and T. Viculin, 2006. Assessment of the radioprotective effects of a mifostine and melatonin of human lymphocytes irradiated with γ-rays *in vitro*. Arch. Hig. Rada. Toksikol., 57: 155-163.
- Burgio, E., P. Piscitelli and L. Migliore, 2018. Ionizing radiation and human health: Reviewing models of exposure and mechanisms of cellular damage. An epigenetic perspective. Int. J. Environ. Res. Public Health, 10: 15-21.
- Al Amri, O.D., A.B. Cundy, Y. Di, A.N. Jha and J.M. Rotchell, 2012. Ionizing radiation-induced DNA damage response identified in marine mussels, *Mytilus* sp. Environ. Pollut., 168: 107-112.
- De Santis, M., E. Di Gianantonio, G. Straface, A.F. Cavaliere and A. Caruso *et al.*, 2005. Ionizing radiations in pregnancy and teratogenesis: A review of literature. Reprod. Toxicol., 20: 323-329.
- Maganha, J., E.D.S. Rocha, M.A.F. Brandão, V.M. Peters and M.D.O. Guerra, 2006. Embryo development alteration in rats treated with lapachol. Braz. Arch. Biol. Technol., 49: 927-934.
- Al-Shaibani, E.A., H.I. Nadia, O.S. Eissa, M.I. Rady and T.Z. Zaki, 2009. Teratological effects of gamma-irradiation during three gestaional intervals in rats. Egypt. J. Hosp. Med., 36: 456-467.
- Zhou, W., J. Xu, K. Zhao, J. Xu, Y. Dong and S. Tong, 2019. Efficacy of human bone marrow mesenchymal stem cell transplantation in repair of radiation-induced damage to the immune system. Int. J. Clin. Exp. Med., 12: 2651-2658.
- Jaggi, J.S., S.V. Seshan, M.R. McDevitt, K. LaPerle, G. Sgouros and D.A. Scheinberg, 2005. Renal tubulointerstitial changes after internal irradiation with α-particle-emitting actinium daughters. J. Am. Soc. Nephrol., 16: 2677-2689.

- Karawya, M., F. Hashim, S.A. Wahab, K. El-Deeb and S. Soliman *et al.*, 1994. Essential oils and lipids of *Nigela sativa* seeds and their biological screening. Zagazig Farma. J., 3: 49-54.
- Hussein, E., 2004. Natural protection of bone marrow transplantation to gamma irradiated pregnant rats in view of better restoration of certain vital organ functions. Ph.D. Thesis, Faculty of Science, Ain Shams University, Egypt.
- 14. Caplan, A.I., 1991. Mesenchymal stem cells. J. Orthop. Res., 9: 641-650.
- 15. Berner, A., C. Pfaller, T. Dienstknecht, J. Zellner and M. Müller *et al.*, 2011. Arthroplasty of the lunate using bone marrow mesenchymal stromal cells. Int. Orthop., 35: 379-387.
- Ashry, O.M., E.M. Hussein and S.F. Salama, 2009. Boosting of antioxidant defence by interferon-Alfa in irradiated bone marrow transplanted rats. Egypt. J. Radiat. Sci. Applic., 22: 19-33.
- 17. Zeng, X., S.P. Yu, T. Taylor, M. Ogle and L. Wei, 2012. Protective effect of apelin on cultured rat bone marrow mesenchymal stem cells against apoptosis. Stem Cell Res., 8: 357-367.
- Kode, J.A., S. Mukherjee, M.V. Joglekar and A.A. Hardikar, 2009. Mesenchymal stem cells: Immunobiology and role in immunomodulation and tissue regeneration. Cytotherapy, 11: 377-391.
- Decleve, A., G.B. Gerber, A. Leonard, M. Lambiet-Collier, A. Sassen and J.R. Maisin, 1972. Regeneration of thymus, spleen and bone marrow in X-irradiated AKR mice. Radiat. Res., 51: 318-332.
- 20. Carleton, H.M., R.A.B. Drury and E.A. Wallington, 2005. Carleton's Histological Technique. Oxford University Press, New York, ISBN: 9780192613103, Pages: 520.
- 21. Pearse, A.G.E., 1953. Histochemistry: Theoretical and Applied. 3rd Edn., Vol. 1. Churchill Livingstone, London.
- 22. Mazia, D., P.A. Brewer and M. Alfert, 1953. The cytochemical staining and measurement of protein with mercuric bromphenol blue. Biol. Bull., 104: 57-67.
- Srinivasan, M., A.R. Sudheer, K.R. Pillai, P.R. Kumar, P.R. Sudhakaran and V.P. Menon, 2006. Influence of ferulic acid on γ-radiation induced DNA damage, lipid peroxidation and antioxidant status in primary culture of isolated rat hepatocytes. Toxicology, 228: 249-258.
- Franco, N., J. Lamartine, V. Frouin, P. Le Minter and C. Petat *et al.*, 2005. Low-dose exposure to γ rays induces specific gene regulations in normal human keratinocytes. Radiat. Res., 163: 623-635.
- Suzuki, K., S. Kodama and M. Watanabe, 2010. Role of Ku80-dependent end-joining in delayed genomic instability in mammalian cells surviving ionizing radiation. Mutat. Res./Fund. Mol. Mech. Mutagen. 683: 29-34.
- 26. Nunia, V., G. Sancheti and P.K. Goyal, 2007. Protection of Swiss albino mice against whole-body gamma irradiation by diltiazem. Br. J. Radiol., 80: 77-84.

- Manda, K. and A.L. Bhatia, 2003. Pre-administration of beta-carotene protects tissue glutathione and lipid peroxidation status following exposure to gamma radiation. J. Environ. Biol., 24: 369-372.
- Cakmak, G., L.M. Miller, F. Zorlu and F. Severcan, 2012. Amifostine, a radioprotectant agent, protects rat brain tissue lipids against ionizing radiation induced damage: An FTIR microspectroscopic imaging study. Arch. Biochem. Biophys., 520: 67-73.
- 29. Emam, N.M.M., 2012. The possible protective role of bone marrow transplantation against alternations induced by gamma radiations on fetal gastrointestinal tract of pregnant albino rats. Egypt. J. Hosp. Med., 49: 628-660.
- Bray, F.N., B.J. Simmons, A.H. Wolfson and K. Nouri, 2016. Acute and chronic cutaneous reactions to ionizing radiation therapy. Dermatol. Ther., 6: 185-206.
- 31. Groen, R.S., J.Y. Bae and K.J. Lim, 2012. Fear of the unknown: lonizing radiation exposure during pregnancy. Am. J. Obstetr. Gynecol., 206: 456-462.
- 32. Fushiki, S., 2013. Radiation hazards in children-Lessons from Chernobyl, Three Mile Island and Fukushima. Brain Dev., 35: 220-227.
- Soffritti, M. and L. Giuliani, 2019. The carcinogenic potential of non ionizing radiations: The cases of S 50 Hz MF and 1.8 GHz GSM radiofrequency radiation. Basic Clin. Pharmacol. Toxicol., 125: 58-69.
- Augustine, A.D., T. Gondré-Lewis, W. McBride, L. Miller, T.C. Pellmar and S. Rockwell, 2005. Animal models for radiation injury, protection and therapy. Radiat. Res., 164: 100-109.
- 35. Yurut-Caloglu, V., M. Caloglu, T. Deniz-Yalta, T. Aktoz and D. Nurlu *et al.*, 2015. Radiation-induced acute kidney toxicity: Protective effect of L-carnitine versus amifostine. Int. J. Radiat. Res., 13: 317-324.
- Venditti, P., R. De Rosa and S. Di Meo, 2004. Effect of cold-induced hyperthyroidism on H₂O₂ production and susceptibility to stress conditions of rat liver mitochondria. Free Radic. Biol. Med., 36: 348-358.
- 37. Hasan, H., 2007. Biological studies of the effect of a venom fraction isolated from the scorpion, *Androctonus amoreuxi* on irradiated rats. M.Sc. Thesis, Faculty of Science, Ain Shams University, Egypt.
- 38. Abu El Naga, N., 2012. Transplanted bone marrow modulates injuries in irradiated rat fetuses anatomical, histological and histochemical studies. Az. J. Pharm. Sci., 45: 208-235.
- Singh, V.K., P.K. Singh, S.Y. Wise, A. Posarac and O.O. Fatanmi, 2013. Radioprotective properties of tocopherol succinate against ionizing radiation in mice. J. Radiat. Res., 54: 210-220.
- Shen, Y., X. Jiang, L. Meng, C. Xia, L. Zhang and Y. Xin, 2018. Transplantation of bone marrow mesenchymal stem cells prevents radiation-induced artery injury by suppressing oxidative stress and inflammation. Oxid. Med. Cell Longev., Vol. 59. 10.1155/2018/5942916.

- 41. Limirio, P.H.J.O., P.B.F. Soares, E.T.P. Emi, C.D.C.A. Lopes and F.S. Rocha *et al.*, 2019. Ionizing radiation and bone quality: Time-dependent effects. Radiat. Oncol., Vol. 14, No. 1. 10.1186/s13014-019-1219-y.
- 42. Shediwah, F., 2005. Control of toxicity induced during chemotherapy and radiotherapy using natural plant substance. Ph.D. Thesis, Zoology Department, Faculty of Science, Ain Shams University, Egypt.
- El Salkh, B., 2009. Histological and histochemical studies on the effect of the alternating magnetic field on the mice lung. Egypt. J. Biomed. Sci., 29: 351-366.
- 44. Tins, B.J., M. Garton, V.N. Cassar-Pullicino, P.N. Tyrrell, R. Lalam and J. Singh, 2015. Stress fracture of the pelvis and lower limbs including atypical femoral fractures-a review. Insights Into Imag., 6: 97-110.
- 45. Abdel Naby, Y., 2011. Boosting the immunological responce of irradiated animal by synergism of stem cell transplantation and cytokines administration. M.Sc. Thesis, Faculty of Science, Zoology Department, Al-Azhar University, Egypt.
- 46. Emam, N.M.M., M.A.R. Ibrahim and H.A.K. Mohammed, 2013. The possible protective role of bone marrow transplantation against alternations induced by gamma radiations on heart of pregnant albino rats and their fetuses. J. Biol. Life Sci., 4: 247-272.
- 47. Inglis, S., J.M. Kanczler and R.O. Oreffo, 2018. 3D human bone marrow stromal and endothelial cell spheres promote bone healing in an osteogenic niche. FASEB J., 33: 3279-3290.
- Eid, F. and A. Al Dossary, 2007. Ultrastructural, histological and histochemical studies on the effect of electraomagnetic field on the kidney of pregnant rats and their fetuses. Egypt. J. Hosp. Med., 28: 306-326.
- 49. Al Gahtani, S., 2006. Histological and histochemical studies on the effect of two different types of magnetic field on the liver and kidney of albino rats. M.Sc. Thesis, Girls College of Science, Dammam, K.S.A.
- 50. Yagi, H., A. Soto-Gutierrez, Y. Kitagawa, A.W. Tilles, R.G. Tompkins and M.L. Yarmush, 2010. Bone marrow mesenchymal stromal cells attenuate organ injury induced by LPS and burn. Cell Transplant., 19: 823-830.
- Ittrich, H., C. Lange, F. Tögel, A.R. Zander and H. Dahnke *et al.*, 2007. *In vivo* magnetic resonance imaging of iron oxide-labeled, arterially-injected mesenchymal stem cells in kidneys of rats with acute ischemic kidney injury: Detection and monitoring at 3T. J. Magn. Reson. Imag., 25: 1179-1191.

- Si, J., H. Zhang, Z. Wang, Z. Wu and J. Lu *et al.*, 2013. Effects of ¹²C⁶⁺ ion radiation and ferulic acid on the zebrafish (*Danio rerio*) embryonic oxidative stress response and gene expression. Mutat. Res./Fundam. Mol. Mech. Mutagen., 745-746: 26-33.
- 53. Kheifets, L., M. Repacholi, R. Saunders and E. van Deventer, 2005. The sensitivity of children to electromagnetic fields. Pediatrics, 116: 303-313.
- 54. George, J., K.R. Rao, R. Stern and G. Chandrakasan, 2001. Dimethylnitrosamine-induced liver injury in rats: The early deposition of collagen. Toxicology, 156: 129-138.
- 55. Abedi, M., D.A. Greer, G.A. Colvin, D.A. Demers and M.S. Dooner *et al.*, 2004. Tissue injury in marrow transdifferentiation. Blood Cells Mol. Dis., 32: 42-46.
- Fu, X., G. Liu, A. Halim, Y. Ju, Q. Luo and A.G. Song, 2019. Mesenchymal stem cell migration and tissue repair. Cells, Vol. 8, No. 8. 10.3390/cells8080784.
- 57. Mukhopadhyay, A., 2013. Perspective on liver regeneration by bone marrow-derived stem cells-A scientific realization or a paradox. Cytotherapy, 15: 881-892.
- 58. Hanley, E.S. and V.P. Knutson, 1990. Lidocaine HCL increases glucose uptake in cultured mouse fibroblasts. Anesthesiology, 73: 363-370.
- 59. Godoi, D.F., C.R. Cardoso, M.J.B. Silva, D.B. Ferraz and P.R. Provinciatto *et al.*, 2013. Reappraisal of total body irradiation followed by bone marrow transplantation as a therapy for inflammatory bowel disease. Immunobiology, 218: 317-324.
- Burt, R.K., Y. Loh, W. Pearce, N. Beohar and W.G. Barr *et al.*, 2008. Clinical applications of blood-derived and marrow-derived stem cells for nonmalignant diseases. JAMA., 299: 925-936.
- 61. Pye, D. and D.J. Watt, 2001. Dermal fibroblasts participate in the formation of new muscle fibres when implanted into regenerating normal mouse muscle. J. Anat., 198: 163-173.
- 62. Kirsch, D.G., J. Grimm, A.R. Guimaraes, G.R. Wojtkiewicz and B.A. Perez *et al.*, 2010. Imaging Primary Lung Cancers in Mice to Study Radiation Biology. Int. J. Radiat. Oncol. Biol. Phys., 76: 973-977.
- 63. Ramakrishna, V., P.B. Janardhan and L. Sudarsanareddy, 2011. Stem cells and regenerative medicine-A review. Annu. Rev. Res. Biol., 1: 79-110.