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Research Article Antioxidant and Anti-Inflammatory Activity of Ethanol Extract Stem of *Etlingera rubroloba* A.D. Poulsen

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Abstract

Background and Objective: *Etlingera rubroloba* A.D. Poulsen (*E. rubroloba*) is a plant endemic to South-East Sulawesi which is used empirically by local people as a pain reliever for joints and to increase endurance after typhoid fever. This study aimed to determine the antioxidant and anti-inflammatory activity of the ethanol extract stem of *E. rubroloba*. **Materials and Methods:** The stem of the ethanol extract of *E. rubroloba* with evaluated for antioxidants using the FRAP method and anti-inflammatory activity with the parameter level of Tumor Necrosis Factor-Alpha (TNF-α) *in vivo* by using 6 groups of rats, namely the normal, the negative (Na-CMC 0.5%), the positive control (Diclofenac sodium) and the ethanol extract group at doses of 200, 300 and 400 mg kg⁻¹ b.wt. **Results:** The results of this study indicated that the antioxidant activity value of the IC₅₀ ethanol extract was $12.720\pm0.12~\mu g~mL^{-1}$ and ascorbic acid (vitamin C) as a standard control was $3.14\pm0.12~\mu g~mL^{-1}$. The TNF- α normal group (7.83 pg mL⁻¹), negative control (250.92 pg mL⁻¹), positive control (123.66 pg mL⁻¹), treatment group dose 200 (192.20 pg mL⁻¹), 300 (97, 95 pg mL⁻¹) and 400 mg kg⁻¹ b.wt. (28.78 pg mL⁻¹). **Conclusion:** This study concluded that the ethanol extract of the stem of *E. rubroloba* has a very strong antioxidant activity and is an anti-inflammatory which is the best in reducing levels of TNF-α at a dose of 400 mg kg⁻¹ b.wt.

Key words: E. rubroloba, ethanol extract, antioxidants, anti-inflammatory, Tumor Necrosis Factor-Alpha (TNF- α), FRAP method, in vivo

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Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Inflammation is a normal protective response caused by injury or tissue damage caused by physical trauma, damaging chemicals or the invasion of pathogenic microorganisms. There are two phases of inflammation, namely acute and chronic. Acute inflammation is the initial response to tissue injury that triggers local vasodilation and increases capillary permeability resulting in fluid accumulation in the injured area. Various mediators contribute to the inflammatory process such as serotonin, histamine, leukotrienes, prostaglandins and pro-inflammatory cytokines such as interferon (IFN), interleukin 1 β and Tumor Necrosis Factor-Alpha (TNF- α)¹.

Cytokines are inflammatory mediators that function in the mobilization of other leukocytes to the injured tissue. The more neutrophils and macrophages that carry out phagocytosis, the more cytokines will be produced. Tumour Necrosis Factor Alpha (TNF- α) is a major cytokine in the acute inflammatory response. TNF- α will function as a protein that gives a signal if there is interference due to infection. Severe infections can trigger the production of large amounts of TNF- α causing systemic reactions².

Changes in environmental conditions and unhealthy lifestyles can make the body susceptible to various types of diseases. One of the causes of disease in the body is free radicals. To protect the body from free radical attack, we need a material that functions as an antioxidant. Antioxidants are secondary metabolites that are used to prevent free radicals. The higher the antioxidant activity, the more free radicals are prevented³.

One of the plants studied is *Etlingera rubroloba* A.D. Poulsen from the genus Etlingera, this plant is thought to have various pharmacological activities. Etlingera species exist in the world about 150-200 species of which about 54 are found in Indonesia, including 48 species on the island of Sulawesi and 6 species from the island of Java. Species of this plant have been used in traditional medicine to treat various diseases and the presence of volatile and nonvolatile entities in these species is gaining research interest among scientists⁹.

Various species of Etlingera that have been reported related to biological and pharmacological aspects are *E. elatior* as hepatoprotective and phytochemical screening results contain alkaloids, flavonoids, tannins and terpenoids⁴. *Etlingera calophrys* contains Yakuchinone A, p-Hydroxybenzoic acid and stigmasterol compounds and is active as an antifioxidant⁵. On the island of Sulawesi, the fruit of *E. elatior* is used as a medicine for nausea and treating typhoid ever⁶. *E. elatior* is also active as an antibacterial

antioxidant^{7,8}. E. elatior fruit is active as Antihyperuricemia⁹. Rhizome of *E. elatior* as antioxidant and antibacterial^{7,10}. E. elatior is active as an antibacterial, deodorant, wound medicine, tyrosinase inhibitor and antioxidant¹¹. E. elatior flower is a fungal antioxidant and antibacterial¹². E. coccinea is an antioxidant in the rhizome, stem and leaves¹³. Meanwhile, E. rubrostriata, E. Littoralis and E. Fulgens are active as antibacterial and antioxidants14. E. sessilanthera and *E. coccinea* as antibacterial¹⁵. *E. pubescens* as antibacterial 16,17, flower extract of E. elatior as antihyperuricemia, antioxidant and chemotherapy against melanoma^{18,19}. Etlingera rubroloba immunomodulator²⁰, fruit *E. rubroloba* has Immunomodulatory Potential on Diabetic-Infected Tuberculosis²¹, E. rubrolaba stem methanol extract as antioxidant²² and Xanthine Oxidase inhibitor²³. E. brevilabrum as cholesterol-lowering²⁴. E. rubroloba fruit as Immunostimulator with increased CD8²⁵.

This study was conducted to examine the potential of ethanol extract of *E. rubroloba* A.D. Poulsen as an antioxidant and anti-inflammatory with parameters of TNF- α levels. The results of this study can be developed as traditional medicine.

MATERIALS AND METHODS

Study area: This research was conducted in the laboratory of the UHO Faculty of Pharmacy, the Laboratory of the UHO Faculty of Medicine and the Faculty of Medicine, Universitas Brawijaya Malang Indonesia, from January to April, 2022.

Materials: The materials used were *E. rubroloba* stem, aqua dest (WaterOne®), 96% ethanol, filter paper, carrageenan, Aqua Pro Injection (Otsuka®), diclofenac sodium 50 mg, Na. CMC 0.5%, Potassium Ferricyanide (K₃Fe(CN)₆, FeCl₃, NaOH, KH₂PO₄, TCA, oxalic acid, pure vitamin C, ethanol pa, 1 cc and 3 cc syringe (OneMed®), microplate flat-bottom polystyrene 96 well (Iwaki, Japan), 1.5 mL Eppendorf tube (Onemed®), Rat TNF-α ELISA KIT. Eppendorf tube (Onemed®, Germany), Rat TNF-α ELISA KIT (Shanghai Korain Biotech Co., Ltd.).

Methods

Sample preparation and extraction: A total of 20 kg of stems of *E. rubroloba* A.D. Poulsen were collected from Laiwoi Village, Laeya District, Konawe Selatan Regency. The sample was determined at the Research Center for Biology, LIPI, Cibinong, Bogor, Indonesia. The samples were cleaned, dried under direct sunlight and then powdered. Then extracted with 96% ethanol (12 L, 3×24 hrs), using the maceration method. The obtained filtrate was evaporated with an evaporator (50°C) and a thick extract was obtained ²⁶.

Antioxidant activity test FRAP method

Activity measurement: The stock solution of extract and vitamin C samples with five concentration variants, namely 25, 20, 15, 10 and 5 μ g mL⁻¹ 1 mL of each concentration, was taken, then 1 mL of 0.2 M phosphate buffer and 1 mL of 1% K₃Fe(CN) 6 were added, then incubated. Then 1 mL of 10% TCA was added with the aim that the potassium ferricyanide complex precipitated, then centrifuged at 3000 rpm for 10 min to speed up the precipitation process. After that, 1 mL of the top layer was taken and put in a test tube, adding 1 mL of distilled water and 0.5 mL of 0.1% FeCl₃. The solution was allowed to stand for a few minutes and its absorption was measured with an ultraviolet-visible (UV-Vis) spectrophotometer at the maximum wavelength obtained.

Inhibition percentage: The antioxidant activity of the sample is determined by the amount of free radical uptake FRAP by calculating the percentage of solution absorption inhibition using the formula²⁷:

Inhibition (%) =
$$\frac{\text{Blanko absorbance} - \text{Sample absorbance}}{\text{Blanko absorbance}} \times 100$$

The IC_{50} value was obtained from the % inhibition and concentration of the extract of *E. rubroloba* A.D. Poulsen by plotting the calculated values in a linear regression equation with concentration (ppm) as the X-axis and the percentage of inhibition as the Y-axis, so that the Eq:

$$Y = aX + b$$

Where:

 $Y = IC_{50}$ value

X = Sample

a = Slope/gradient

b = Intercept

Antioxidant activity is expressed by the IC_{50} value (50% Inhibition Concentration), which is the concentration of the sample that can reduce 50% FRAP radicals²⁷.

Anti-inflammatory activity test

Grouping of test animals: The test animals were divided into 6 test groups and the grouping of test animals was carried out entirely randomly with the number following the Federer formula. This study used a sample of 4 rats for each group and as a backup, if there was a sample failure in the study. So, the total number of samples used in this study was 30 individuals in 6 treatment groups.

Induction of inflammation: Edema was made on the right hind paw of rats by inducing 0.1 mL of 1% carrageenan solution subplantar. Carrageenan was chosen as an irritant for making edema because it has several advantages. Namely, it does not leave scars, does not cause tissue damage and can provide a more sensitive response to anti-inflammatory drugs than other irritants, so it is suitable to be chosen as an inductor for edema²⁸.

Inflammation measurement: Inflammation was measured by dipping the mice's feet into a tube containing the measuring solution to the mark. The change in the volume of the solution was recorded for a certain time (Vt) of the mice's feet. Inflammation volume is the difference in the volume of the mice's feet at a certain time (Vt) with the initial leg volume (Vo).

Anti-inflammatory potency test based on TNF- α

Treatment and collection and storage of rat blood: After the rats were induced with 1% carrageenan and experienced edema after 1 hr, the rats were given treatment in the form of administration of *E. rubroloba* stem ethanol extract 200, 300 and 400 mg kg $^{-1}$ b.wt., as the test group, administration of Na-CMC 0 and 5% as a negative control and administration of sodium diclofenac as a positive control with a dose of 1.13 mg. After that, blood samples were taken in the second hour to assess the levels of the inflammatory mediator TNF-α. Blood sampling was carried out on the hearts of rats. After taking the blood, the blood is put in a tube containing the anticoagulant EDTA.

ELISA test: Enzyme-Linked Immunosorbent Assay (ELISA) for measuring levels of TNF- α follows protocol Rat Tumor Necrosis Factor, TNF- α ELISA Kit which measured absorbance at a wavelength of 450 nm.

Data analysis: The data obtained were analyzed statistically using the SPSS program. The effect of the ethanol extract of *E. rubroloba* stems and the decrease in TNF- α levels in all treatment and comparison groups was analyzed by using the One-way ANOVA (Analysis of Variance) statistical test. Then proceed with the *post hoc* Tests to compare between groups.

RESULTS AND DISCUSSION

Antioxidant activity of FRAP method: Measurement of antioxidant activity was carried out using the FRAP test, where the ethanol extract of the stem of *E. rubroloba* was used as a

sample and vitamin C as a standard solution. Antioxidant testing is expressed by the IC_{50} (inhibition concentration) parameter. The amount of antioxidant activity is indicated by the IC_{50} value, which is the concentration of the sample solution required to inhibit 50% of free radicals. The smaller the IC_{50} value of the compound, the greater the ability of the compound to ward off free radicals.

The result of determining the maximum wavelength of the FRAP solution is 582 nm with an absorbance of 0.576 nm. Determination of the antioxidant activity of the ethanolic stem extract of *E. rubroloba* A.D. Poulsen compared with ascorbic acid (vitamin C) standard²⁹.

The measurement results of antioxidant activity indicated that the comparison of vitamin C and the ethanol extract of stem E. rubroloba A.D. Poulsen has antioxidant activity values in the very strong category. The result of percentage inhibition in ascorbic acid has a higher percentage (%) of inhibition than the ethanol extract of E. E0 rubroloba stem A.D. Poulsen, which indicates that the greater the percentage (%) of inhibition, the smaller the E1 rubroloba value, which indicates the greater the ability of the sample to capture free radicals. Thus, vitamin E1 used as a comparison has a stronger free radical scavenging ability than the ethanol extract of E1. E2 rubroloba A.D. Poulsen stem. Judging from the E3 value, it is still a very strong category.

The results showed that the IC₅₀ value of the *E. rubroloba* stem ethanol extract of A.D. Poulsen was $12.71\pm0.12 \,\mu g \,mL^{-1}$ and vitamin C was $3.14\pm0.12~\mu g~mL^{-1}$. The amount of antioxidant activity was indicated by the IC₅₀ value, which is the concentration of the sample solution needed to reduce 50% of free radicals. A compound is said to be a very strong antioxidant if it has an IC_{50} value of less than 50 µg mL⁻¹, strong for an IC₅₀ value of 50-100 µg mL⁻¹ and moderate if it has a value of 101-250 μg mL⁻¹, weak if the IC₅₀ value is 251-500 μ g mL⁻¹ and >500 have no antioxidant activity³⁰. Based on these results, the IC₅₀ value of Vitamin C and the ethanol extract of *E. rubroloba* stem A.D. Poulsen had an IC₅₀ value of less than 50 μ g mL⁻¹. Thus, the stem of *E. rubroloba* A.D. Poulsen is a plant with a very strong antioxidant activity. This study is in line with previous research, where the antioxidant test used methanol extract of E. rubroloba with the DPPH method with a very strong antioxidant value^{22,23}. In addition, it was previously reported that a different species, E. calophrys, has a very strong antioxidant capacity⁵.

Anti-inflammatory activity

Rat edema examination results: Edema is a buildup of fluid in the lower layers of the skin, which is a sign of inflammation. Carrageenan is a strong chemical substance that releases

inflammatory and pro-inflammatory mediators (prostaglandins, leukotrienes, histamine, bradykinin, TNF- α , etc.). Carrageenan was chosen to test anti-inflammatory drugs because it is not antigenic and does not cause systemic effects. Cardinal signs of inflammation, namely edema, hyperalgesia and erythema, occur after injection due to the activity of pro-inflammatory agents, such as bradykinin, histamine, complement and reactive oxygen³¹. Neutrophils immediately migrate to sites of inflammation and can produce pro-inflammatory reactive oxygen species. Based on the plethysmometer test, the average volume measurement results for each treatment group were obtained, as shown in Fig 1.

The samples used were the feet of rats that had edema after being induced by carrageenan with a concentration of 1% and a decrease in the volume of the rat's feet after administration of the test solution. The anti-inflammatory activity of the test material was shown by its ability to reduce edema caused by carrageenan induction in the soles of mice. The research was carried out by injecting 1% carrageenan suspension into the paws of the male Wistar strain test rats intraplantar and then giving the test solution.

Results of examination of TNF- α levels in Wistar strain male

rats: Measurement of TNF-α levels using Enzyme-Linked Immunosorbent Assay (ELISA). The ELISA was performed to see the quantitative or qualitative TNF-α based on colorimetric readings³². Measurement of TNF-α levels was carried out by taking rat blood 1 hr after carrageenan induction. This blood collection time was adjusted to the peak level of TNF-α as a pro-inflammatory cytokine in the 1st hr. The levels obtained will be the initial data on TNF-α levels, namely when inflammation occurs. After that, each group was given their respective treatment. One hour later, blood was drawn and centrifuged to obtain plasma. This plasma is then read on the ELISA reader. The results of the TNF-α levels can be seen in Fig. 2.

Based on Fig. 2, It can be seen that the normal group is the group that has the lowest levels, namely 7.83 pg mL $^{-1}$, because it is the level of TNF- α in the physiological state of the body. Where TNF- α will increase as the degree of inflammation increases. Inflammation is the body's defense response against the entry of foreign substances in the form of carrageenan so that pro-inflammatory cytokines such as TNF- α appear as an immune response. The TNF- α acts as an immune response modulator that can mediate the induction of adhesion molecules. Increased induction of adhesion molecules serves to facilitate leukocytes' adherence to the endothelium surface and activate leukocytes. These

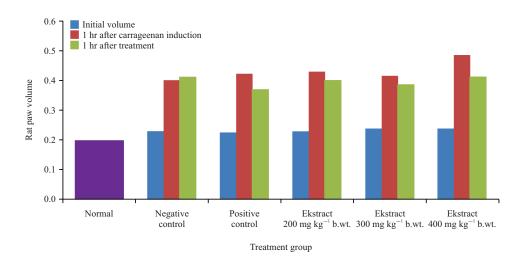


Fig. 1: Volume of edema before and after treatment

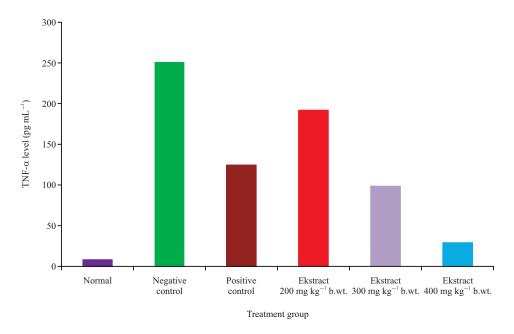


Fig. 2: TNF- α levels in the treatment and normal group

leukocytes will neutralize inflammatory agents at the site of injury, so the higher the degree of inflammation, the more leukocytes needed and the more TNF- α levels must play a role.

These data indicated that the administration of negative Na-CMC control was the initial comparison of the entire treatment group. The level of negative control was 250.92 pg mL⁻¹. This value is not much different from the decrease in levels caused by the administration of the ethanol extract of stem *E. rubroloba* A.D. Poulsen 200 mg kg⁻¹ b.wt., equal to 192,20 pg mL⁻¹. This indicated that the extract with a dose of 200 mg kg⁻¹ b.wt., has less effect as an anti-

inflammatory. However, the decrease in TNF- α levels occurred as the dose of the extract was increased. In the extract with a dose of 300 mg kg⁻¹ b.wt., the levels of TNF- α decreased to 97.95 pg mL⁻¹, where this value is lower than the dose of 200 mg kg⁻¹ b.wt. This shows that a 300 mg kg⁻¹ b.wt., dose has a better anti-inflammatory effect than 200 mg kg⁻¹ b.wt. At the same time, the lowest value of all dose variations is 400 mg kg⁻¹ b.wt., which can reduce TNF- α levels to 28,78 pg mL⁻¹. This showed that a 400 mg kg⁻¹ b.wt., dose has a better anti-inflammatory effect than a dose of 200 and 300 mg kg⁻¹ b.wt. and positive control (123.66 pg mL⁻¹). The

anti-inflammatory activity of the ethanolic stem extract of *E. rubroloba* A.D. Poulsen was probably due to the presence of flavonoids, which can decrease the secretion of pro-inflammatory cytokines such as TNF- α^{33} .

The data obtained were carried out by the Shapiro-Wilk Test to see the normality of the data and the Levene test to see the homogeneity of the data. Based on the normality test of the data using the Shapiro-Wilk test, it is known that the data distribution of all groups is normally distributed (p>0.05) and based on the results of the data variance test that the data has the same or homogeneous variance (p>0.05). Therefore, the data were analyzed by the One-way ANOVA (Analysis of Variance) Test. Based on the One-way ANOVA Test, the value (p<0.05) was obtained so that it could be interpreted that there was a significant effect of the administration of ethanol extract of *E. rubroloba* A.D. Poulsen stem on TNF- α levels in male Wistar rats after the 3 hrs of treatment.

CONCLUSION

The ethanol extract of stem *E. rubroloba* A.D. Poulsen has antioxidant activity with a very strong category and has potential as an anti-inflammatory by TNF- α parameters and this research is a reference in the development of traditional medicines. The conclusion of this study showed that the ethanol extract of the stem *Etlingera rubroloba* A.D. Poulsen had a very strong category of antioxidant activity with an IC₅₀ value of 12.71 \pm 0.12 g mL⁻¹ as measured using the FRAP method and had acted as an anti-inflammatory with *in vivo* by reducing TNF- α levels at a dose of the best is 400 mg kg⁻¹ b.wt. This research is a reference in the development of traditional medicine, especially gout, rheumatism, uric acid and natural antioxidants.

SIGNIFICANCE STATEMENT

This study found that the stem of *Etlingera rubroloba* A.D. Poulsen has very strong antioxidant and anti-inflammatory activity with the parameter of Tumor Necrosis Factor-Alpha (TNF- α). This research is a reference for the development of traditional medicine.

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REFERENCES

- Dhingra, A.K., B. Chopra, J.S. Dua and D.N. Parsad, 2017. New insight on inflammation and its management: A review. J. Innovations Pharm. Biol. Sci., 7: 117-126.
- 2. Supit, I.A., D.H.C. Pangemanan and S.R. Marunduh, 2015. Profil tumor necrosis factor (TNF-α) berdasarkan indeks massa tubuh (IMT) pada mahasiswa fakultas kedokteran UNSRAT angkatan 2014. eBiomedik, 3: 640-643.
- 3. Sari, T.M., O. Fera and Y.P. Yonedi, 2021. Antioxidant activity of ethanol extract of konyal passion fruit peel (*Passiflora lingularis* f. lobalata). J. Katalisator, 6: 241-253.
- 4. Fristiohady, A., B. Sadarun, W. Wahyuni, M.H. Malaka and F. Ahmad *et al.*, 2020. Isolation and identification of secondary metabolite acetone extract *Aaptos* sp. and its antioxidant properties and acute toxicity. J. Appl. Pharm. Sci., 10: 81-89.
- Sahidin, I., W. Wahyuni, M.H. Malaka, A. Jabbar, I. Imran and M.A. Manggau, 2018. Evaluation of antiradical scavenger activity of extract and compounds from *Etlingera calophrys* stems. Asian J. Pharm. Clin. Res., 11: 238-241.
- Rahayu, M., S. Sunarti, D. Sulistiarini and S. Prawiroatmodjo, 2006. Traditonal use of medicinal herbs by local community of Wawonii Island, Southeast Sulawesi. Biodiversitas J. Biol. Divers., 7: 245-250.
- 7. Chan, E.W.C., Y.Y. Lim, L.F. Wong, F.S. Lianto and S.K. Wong *et al.*, 2008. Antioxidant and tyrosinase inhibition properties of leaves and rhizomes of ginger species. Food Chem., 109: 477-483.
- 8. Lachumy, S.J.T., S. Sasidharan, V. Sumathy and Z. Zuraini, 2010. Pharmacological activity, phytochemical analysis and toxicity of methanol extract of *Etlingera elatior* (torch ginger) flowers. Asian Pac. J. Trop. Med., 3: 769-774.
- Jabbar, A., Wahyuni, M. Leorita, M.I. Yusuf and Sahidin, 2021.
 Antihyperuricemia activity of wualae fruit (*Etlingera elatior* Jack R. M. Smith) ethanol extract *in vivo*. J. Farmasi Sains dan Praktis, 7: 313-320.
- Ficker, C.E., M.L. Smith, S. Susiarti, D.J. Leaman, C. Irawati and J.T. Arnason, 2003. Inhibition of human pathogenic fungi by members of Zingiberaceae used by the Kenyah (Indonesian Borneo). J. Ethnopharmacol., 85: 289-293.
- 11. Chan, E.W.C., Y.Y. Lim, S.K. Wong, K.K. Lim, S.P. Tan, F.S. Lianto and M.Y. Yong, 2009. Effects of different drying methods on the antioxidant properties of leaves and tea of ginger species. Food Chem., 113: 166-172.
- 12. Wijekoon, M.M.J.O., R. Bhat and A.A. Karim, 2011. Effect of extraction solvents on the phenolic compounds and antioxidant activities of bunga kantan (*Etlingera elatior* Jack.) inflorescence. J. Food Compos. Anal., 24: 615-619.
- Shahid-Ud-Daula, A.F.M., A.S. Kamariah, L.B.L. Lim and N. Ahmad, 2015. Phytochemical screening, antioxidant, and antimicrobial activities ofleaves, stems, and rhizomes of *Etlingera coccinea* (Blume) S. Sakai & Nagam. Int. J. Pharmacogn. Phytochem. Res., 7: 873-883.

- Chan, E.W.C., Y.Y. Lim and M. Omar, 2007. Antioxidant and antibacterial activity of leaves of *Etlingera* species (Zingiberaceae) in peninsular Malaysia. Food Chem., 104: 1586-1593.
- Daniel-Jambun, D., J. Dwiyanto, Y.Y. Lim, J.B.L. Tan, A. Muhamad, S.W. Yap and S.M. Lee, 2017. Investigation on the antimicrobial activities of gingers (*Etlingera coccinea* (Blume) S.Sakai & Nagam and *Etlingera sessilanthera* R.M.Sm.) endemic to Borneo. J. Appl. Microbiol., 123: 810-818.
- Daniel-Jambun, D., K.S. Ong, Y.Y. Lim, J.B.L. Tan, S.W. Yap and S.M. Lee, 2019. Bactericidal and cytotoxic activity of a diarylheptanoid (etlingerin) isolated from a ginger (*Etlingera* pubescens) endemic to Borneo. J. Appl. Microbiol., 127:59-67.
- Ghasemzadehk, A., H.Z.E. Jaafar, A. Rahmat and S. Ashkani, 2015. Secondary metabolites constituents and antioxidant, anticancer and antibacterial activities of *Etlingera elatior* (Jack) R.M.Sm grown in different locations of Malaysia. BMC Complementary Altern. Med., Vol. 15. 10.1186/s12906-015-0838-6.
- Dewi, A.R., I. Nur'Aini, I.S. Bahri, H.N. Afifah, A. Fattah and W.A.S. Tunjung, 2016. Antihyperuricemic activity of ginger flower (*Etlingera elatior* Jack.) extract in beef broth-induced hyperuricemic rats (*Rattus norvegicus*). AIP Conf. Proc., Vol. 1755. 10.1063/1.4958573.
- Krajarng, A., M. Chulasiri and R. Watanapokasin, 2017. *Etlingera elatior* extract promotes cell death in B16 melanoma cells via down-regulation of ERK and Akt signaling pathways. BMC Complementary Altern. Med., Vol. 17. 10.1186/s12906-017-1921-y.
- 20. Ilyas, Y.M., A. Diantini, E. Halimah, R. Amalia, M. Ghozali and E. Julaeha, 2021. Potential immunomodulator fraction fruit of *Etlingera rubroloba* A.D Poulsen against macrophage phagocytosis and interleukin-12 levels in BCG-stimulated Balb/C mice. Int. J. Pharm. Res., Vol. 13. 10.31838/ijpr/2021.13.01.478.
- 21. Ilyas, Y.M., A. Diantini, E. Halimah, R. Amalia and M. Ghozali *et al.*, 2022. Phytochemical analysis and immunomodulatory potential on diabetic-infected tuberculosis by fruit *Etlingera rubroloba* A.D. Poulsen. Pak. J. Biol. Sci., 25: 669-675.
- Jabbar, A., S. Wahyuono, I. Sahidin and I. Puspitasari, 2021. Free radical scavenging activity of methanol extract and compounds isolated from stems of *Etlingera rubroloba* A.D. Poulsen. Int. J. Pharm. Res., Vol. 13. 10.31838/ijpr/ 2021.13.01.179.

- Jabbar, A., S. Wahyuono, I. Sahidin and I. Puspitasari, 2021.
 Xanthine oxidase inhibitory activity and DPPH radical scavenging assay of isolated compound from *Etlingera rubroloba* (Blume) A.D Poulsen stem. Int. J. Pharm. Res., Vol. 13. 10.31838/ijpr/2021.13.01.316.
- 24. Mahdavi, B., 2014. Chemical constituents of the aerial parts of *Etlingera brevilabrum* (Zingiberaceae). Der Pharma Chem., 6: 360-365.
- 25. Ilyas, Y.M., A. Diantini, M. Ghozali, I. Sahidin and W.O. Nurfinti, 2021. Immunostimulatory activity of *Etlingera rubroloba* A.D. Poulsen fruit ethanol extract against CD8 levels *in vivo* model. Med. Sains: J. Ilmiah Kefarmasian, 6: 123-132.
- Wahyuni, M.H. Malaka, A. Fristiohady, M.I. Yusuf and Sahidin, 2017. Imunomodulator potential of kecombrang fruit ethanol extract (*Etlingera elatior* (Jack) on macrophag phagocytic activity of male mices, BALB/C strain). Pharmacon J. Ilmiah Farmasi, 6: 350-355.
- 27. Susana, I., A. Ridhay and S. Bahri, 2018. The atioxidant activity of kecombrang (*Etlingera elatior*) stem extract base on various levels of polar solvent. Kovalen: J. Riset Kimia, 4: 16-23.
- 28. Suryandari, S.S., E. de Queljoe and O.S. Datu, 2021. Antiinflammatory activity test of ethanol extract of sesewanua leaves (*Clerodendrum squamatum* Vahl.) towards white rats (*Rattus norvegicus* L.) induced by carrageenan. Pharmacon, 10: 1025-1032.
- 29. Awaluddin, N. and S. Wahyuningsih, 2019. Antioxidant activity testing of children's methanol extract (*Croton oblongus* Burm) using DPPH method. J. Pharm. UIN Alauddin Makassar, 7: 38-45.
- 30. Molyneux, P., 2004. The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity. Songklanakarin J. Sci. Technol., 26: 211-219.
- 31. Alfanda, D., Slamet and S. Prasojo, 2021. Anti-Inflammatory activity test of hexane, ethyl acetate, and ethanol extracts from kecombrang (*Etlingera elatioi*) leaves in Wistar male white rats (*Rattus norvegiucus*). Cerata J. Ilmu Farmasi, 12: 36-41.
- 32. Sakamoto, S., W. Putalun, S. Vimolmangkang, W. Phoolcharoen, Y. Shoyama, H. Tanaka and S. Morimoto, 2018. Enzyme-linked immunosorbent assay for the quantitative/qualitative analysis of plant secondary metabolites. J. Nat. Med., 72: 32-42.
- 33. Ginwala, R., R. Bhavsar, de Gaulle I. Chigbu, P. Jain and Z.K. Khan, 2019. Potential role of flavonoids in treating chronic inflammatory diseases with a special focus on the anti-inflammatory activity of apigenin. Antioxidants, Vol. 8. 10.3390/antiox8020035.