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Review Article

Methamphetamine Neurotoxicity: Neurotoxic Effects, Mechanism of Toxicity, Molecular Mechanisms and Treatment Strategies

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Abstract

Methamphetamine (METH) is a highly addictive and dangerous drug that mainly affects neurotransmitters in the brain and leads to feelings of alertness and euphoria. The METH use can lead to addiction, which has become a worldwide problem, resulting in a slew of public health and safety issues. Recent studies showed that chronic METH use can lead to neurotoxicity, neuro-inflammation and oxidative stress which can lead to neuronal injury. This review discussed the history of METH use, the link between METH use and neurotoxicity, the molecular mechanism and the different treatment strategies. This study attempted to discuss some of the drug's principal impacts and gave proof in favor of a few of the cellular and molecular causes of METH neurotoxicity. In addition, it demonstrates the most recent treatment strategies involving mitigating METH-induced neurotoxicity. However, future studies are needed to better understand the mechanism by which METH use induced neurotoxicity.

Key words: Methamphetamine, neurotoxicity, neuroinflammation, excitotoxicity, treatment, oxidative stress

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Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Methamphetamine (METH) is a highly addictive and dangerous drug that mainly affects neurotransmitters in the brain and leads to feelings of alertness and euphoria¹. The METH is classified as a Schedule II drug under the Controlled Substances Act due to its high potential for abuse and addiction². The METH was first synthesized in Japan in 1919 and was used during World War II to keep soldiers awake and alert. The METH has both short- and long-term impacts on the body. In the short term, users report an intense rush of euphoria, decreased appetite and increased energy, blood pressure and heart rate. However, these effects are short-lived and are followed by a crash that can last for days^{3,4}. The long-term use of METH can lead to severe physical and mental health problems. Chronic use can cause damage to the brain's dopamine (DA) system, which can result in symptoms similar to Parkinson's disease. Recently, Vandenberg *et al.*⁵ showed that chronic METH exposure can lead to long-lasting neuro-inflammation and neuronal injury but the exact mechanism is still unclear. It can also cause tooth decay, skin sores, weight loss, insomnia, paranoia, aggression, hallucinations and delusions. High-dose or chronic METH exposure causes excessive dopaminergic activity that can lead to the development of psychotic features⁶. Methamphetamine stimulates the release of monoamine neurotransmitters, including norepinephrine, DA and serotonin, which has an impact on the Central Nervous System (CNS). Users often require professional treatment to overcome their addiction. Treatment options include behavioral therapy, medication-assisted treatment, support groups such as Narcotics Anonymous or a combination of these approaches. However, a drug called levo-tetrahydropalmatine has recently been approved to be an important therapeutic drug for METH addiction and neurotoxicity⁷. In addition, there are new trends to develop anti-METH vaccines⁸. The addictive nature of METH and its effects on humans need a way forward to overcome it. To include articles in this review, a broad literature search was conducted in January, 2024 and repeated in March and April of the same year within the PubMed databases. The search terms "methamphetamine", "METH" and field-specific terms such as "pharmacokinetics, excitotoxicity, oxidative stress and treatment" were used. Finally, the research articles for the last 10 years ago have been narrowly distilled to provide an up-to-date summary of the current knowledge.

Chemical structure and names: The METH belongs to the amphetamine class because the amino group of

(S)-amphetamine has a methyl substituent (Fig. 1). The full systematic name, according to IUPAC, is (2S)-N-methyl-1-phenylpropan-2-amine, which had the molecular formula $C_{10}H_{15}N$ and a molecular mass of 149.233 g/mol. The asymmetric carbon atom produces two enantiomers. Previously known as the [-]- or l-stereoisomer and the [+] or d-stereoisomer, these two forms are now known as the R- and S-stereoisomers. The other names of METH include "crank", "snappy", "crystal", "glass" (Philippines), "tik" (South Africa), "yaa baa" (Thailand) and "shabu" (Arabia)⁹.

Prevalence and epidemiology: The METH belongs to the amphetamine-type stimulant family, which also includes amphetamine, methylene deoxy methamphetamine and other amphetamines¹⁰. The METH was first manufactured in the early 1900s and was unregulated as a nasal decongestant, alertness enhancer and weight loss aid. It was widely utilized by several armed forces throughout World War II, the Korean War and the Vietnam War. In the 1950s, Japan had a high prevalence of abuse, followed by the United States in the 1960s. The term "crank" alludes to biker gangs' transportation of METH hidden in the crankcase of their motorcycles. From the 1970s through the 1990s, the Southwestern and West Coast States (including Hawaii) had the highest rate of abuse. Over the last decade, all regions of the United States have seen a considerable increase in the number of people taking drugs and visiting emergency rooms. According to a recent report, 27 million people used METH and related amphetamine-type stimulants in 2019, accounting for 0.5% of the global adult population. As per UNODC (2020), the rate of METH use in the population of North America, Australia and Asia was 2.3, 1.3 and 0.5%, respectively¹¹.

It is regarded as the second most prevalent drug after cannabis¹². Adolescent abuse has recently been regarded as rampant. When reporting their drug past, METH addicts are often dishonest and distrustful of medical personnel¹³. According to a recent report of Al-Asmari¹⁴, the number of METH-related deaths in Jeddah, Saudi Arabia, increased by more than 500%. Most of the cases were violent, unintentional and suicides.

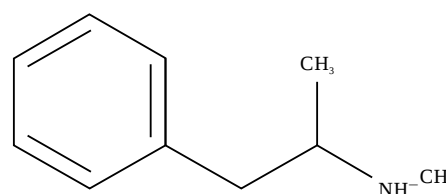


Fig. 1: Chemical structure of METH

Data from the National Survey on Drug Use and Health (NSDUH) for 2021 show that 6.0% of the population or about 16.8 million persons aged 12 or older, had used methamphetamine at least once in their lifetime (2021 DT 1.1). According to recent reports 36 million people are predicted to have used amphetamines in 2021¹⁵.

Compared to non-users, young people who used methamphetamine had a higher likelihood of having conduct disorder. An estimated 95,000 children in Australia between the ages of 4 and 17 were reported to have a conduct disorder in 2022¹⁶. In addition, extreme methamphetamine use was linked to a sudden death in a recent case report by Chansaengpetch *et al.*¹⁷.

A major problem in the Middle East and North Africa (MENA) region is methamphetamine usage. For example, methamphetamine was found to be the most commonly abused illicit drug in Kuwait, where amphetamines were found to be collected in greater amounts than other psychoactive substances¹⁸.

There is no one clear explanation for the seemingly lower rates of METH abuse in the MENA region when compared to the rest of the world due to the complexity of the data. Here are a few potential motivators:

- Because of religious and cultural reasons, METH usage can be severely stigmatized in MENA nations. Because of this stigma, fewer people may ask for assistance or confess to using, which could result in underreported cases
- In the MENA region, strict drug prohibitions and severe punishments may discourage some people from using METH. But this can also drive the drug trade underground, which complicates the process of gathering statistics
- Some MENA nations may have less developed public health data collection and reporting systems than other regions. This may result in incomplete or erroneous data regarding drug use

Methamphetamine abuse is still a major issue in the world, having a terrible impact on both individuals and communities. It is a major cause of violent crime, the spread of infectious diseases and societal evils including child neglect and unemployment. Knowing its effects aids in the development of treatment and preventative plans.

MECHANISM OF TOXICITY

Methamphetamine-induced neurotoxicity: As a lipophilic substance, METH may easily cross the blood-brain barrier and

enter the brain, where it predominantly increases the release of central and peripheral neurotransmitters such as DA, serotonin, norepinephrine and glutamate (Glu)¹⁹. The effects of METH on the Central Nervous System (CNS) have been extensively studied and it has long been known that METH promotes DA neurotransmission by regulating the activity of DA transporters²⁰. Additionally, it inhibits monoamine oxidase, which lowers monoamine metabolism²¹. The METH causes blocking the transport of endogenous neurotransmitters, inhibiting synaptic reuptake and decreasing the expression of transporters at the cell surface²². Since Glu is a key excitatory neurotransmitter in the brain, it has been suggested that Glu contributes significantly to the excitotoxicity brought on by METH²³. At low or moderate dosages, METH induces euphoria, faster heart rate, reduced weariness, increased temperature, higher blood pressure, pupil dilation, reduced appetite, behavioral disinhibition and increased alertness and energy. While higher doses result in hypertension, paranoia, confused speech, nervousness and sweating²⁰. Acute or chronic consumption of METH can lead to several neurotoxic effects due to oxidative stress which happens mainly due to the release of DA²¹. According to Kohno *et al.*²⁴, METH is also known to cause excessive release of DA and Glu, which damages the presynaptic membrane by causing oxidative stress and the production of peroxide radicals which lead eventually to brain degeneration directly or indirectly. According to Chao *et al.*²⁵, Sig-1Rs can activate microglia in response to METH stimulation by generating reactive oxygen species (ROS) and activating the PI3K/Akt and mitogen-activated protein kinase pathways. A recent study looked at the ability of calcitriol to protect against METH-induced reductions in striatal serotonin (5-HT) release and content in male rats. These findings imply that calcitriol can give significant protection against the 5-HT depleting effects of neurotoxic METH dosages²⁶.

Methamphetamine-induced oxidative stress: The oxidative stress is caused by an imbalance between a biological system's ability to detoxify these reactive byproducts and the production and accumulation of ROS in cells and tissues disrupting oxidation-reduction (redox) reactions²⁷. In normal and pathological situations, ROS are primarily created by mitochondria through the generation of free radicals, like superoxide radicals, hydrogen peroxide, hydroxyl radicals and singlet oxygen radicals inside endothelium and inflammatory cells²⁸. The neurotoxic effects of METH are, in part, mediated by oxidative stress, which plays a crucial role in cellular toxicity. When METH enters neurons, it displaces DA from vesicles, resulting in elevated intracellular and synaptic DA levels which leads to the auto-oxidation and increased

metabolism of DA, producing ROS such as hydrogen peroxide as by-products which are responsible for oxidative damage¹⁹. Additionally, DA can be further oxidized into quinones and semiquinones, which generate nitrogen and superoxide radicals that contribute to oxidative stress²⁹. Others have demonstrated that rats exposed to METH had higher levels of malondialdehyde, a byproduct of lipid peroxidation by ROS, in certain brain areas leading to the destruction of dopaminergic neurons³⁰. In addition, oxidative stress caused by METH leads to the oxidation of cellular macromolecules including proteins, lipids and DNA³¹. Protein oxidation results in the formation of disulfuric bridges and misfolded proteins while lipid oxidation produces highly reactive 4-hydroxynonenal, which further contributes to cellular damage³¹. Also, epigenetic changes in DNA methylation levels were stimulated by METH indicating nuclear damage and neurological disorders^{31,32}. The METH-induced oxidative stress also disturbs the redox balance and inhibits mitochondrial complex II, leading to elevated oxidative stress and increased mitochondrial damage. This process contributes to neurodegeneration and the release of neuro-melanin, which exacerbates neuro-inflammation and the neurodegenerative process^{31,33}. Furthermore, METH increases nitric oxide production through nitric oxide synthase activation leading to the upregulation of alpha-synuclein expression which enhances cellular oxidative stress and contributes to neuronal damage³⁴.

Methamphetamine-induced neuro-inflammation: An inflammatory response in the brain or spinal cord is referred to as neuro-inflammation. The synthesis of cytokines, chemokines, ROS and secondary messengers mediates this inflammation³⁵. The METH has been demonstrated to cause inflammatory reactions in regions that have damaged DA and Glu^{24,36}. The molecular mechanism of METH-induced neuro-inflammation involves the activation of microglia. The exact mechanism of microglial activation is still unknown, but it is believed that dopaquinones (DAQs), metabolites of DA, play a significant role in gene expression of microglial cells³⁷. Activated microglia release various neurotoxic molecules, including pro-inflammatory cytokines, proteinases and ROS, which contribute to neuro-inflammation³⁸. The secretion of excite-toxic Glu by activated microglia also plays a role in mediating excite-toxicity and neuro-inflammation, ultimately leading to neurodegeneration³⁹. Repeated administration of METH activates Glu receptors and promotes Glu release which leads to activation of transcription factor NF- κ B. The NF- κ B then triggers inflammatory mediators, such as Tumor Necrosis Factor- α (TNF- α), Interleukin-1 β (IL-1 β) and Interleukin-6 (IL-6). These cytokines can further enhance

extracellular Glu levels by inhibiting uptake and increasing release from microglial cells, creating a feed-forward loop that amplifies neurotoxicity⁴⁰. The effects of METH include the neuronal release of Damage-Associated Molecular Patterns (DAMPs), which can trigger neuro-inflammatory processes (Fig. 2). Moreover, the activation of microglia by METH is reported to be linked with Toll-Like Receptor 4 (TLR4), which is involved in the immune response to pathogens⁴¹. Billod *et al.*⁴² reported that the TLR4 receptor can activate Myd88 pathways. As a result, Tumor Necrosis Factor Receptor-Related Factor 6 (TRAF6) and Interleukin-1 Receptor Related Kinase (IRAK) is activated, which results in the activation of Nuclear Factor- κ B (NF- κ B)⁴³. According to White *et al.*⁴⁴, Sig-1Rs can activate microglia in response to METH stimulation by generating ROS and activating the PI3K/Akt and MAPK pathways. The NF- κ B signaling system and the MAPK signaling pathway share a strong relationship⁴⁵ and both are crucial for the activation of inflammatory cytokines by METH⁴⁶. Kobeissy *et al.*⁴⁷ have demonstrated the increase of these cytokines following METH intoxication, highlighting their significant role in mediating brain injury associated with METH-induced neurotoxicity.

Methamphetamine-induced excitotoxicity: The excitotoxicity is defined as cell death caused by excitatory amino acid toxicity. Neuronal excitotoxicity usually refers to the destruction and death of neurons caused by prolonged Glu exposure. It follows an excessive ion influx into the cell which activates enzymes that destroy proteins, membranes and nucleic acids⁴⁸. In addition, Glu terminals repeatedly undergo depolarization-repolarization cycles. As a result, the degeneration of post-synaptic neurons occurs through the activation of NMDA (N-Methyl-D-Aspartate) ionotropic receptors which play a significant role in METH-induced excitotoxicity⁴⁹. The molecular mechanisms underlying METH-induced excitotoxicity involve excessive release and accumulation of the neurotransmitter Glu, particularly the ionic form L-Glu, from both neuron and glial cells to the extracellular space⁵⁰ which contribute to excitotoxicity¹⁰. Another study has shown that high doses of METH stimulate the production of ER stress-related genes, including ATF4, CHOP and caspase-12 (Fig. 3). Furthermore, METH-induced ER stress has been linked to dopaminergic toxicity through the activation of DA receptor D. The apoptosis or programmed cell death, is induced by ER stress through the activation of death receptors and the involvement of mitochondrial-dependent apoptosis pathways⁵¹. Also, METH indirectly increases proteolysis of the cytoskeletal protein spectrin which contributes to DA terminal degradation in the striatum⁵².

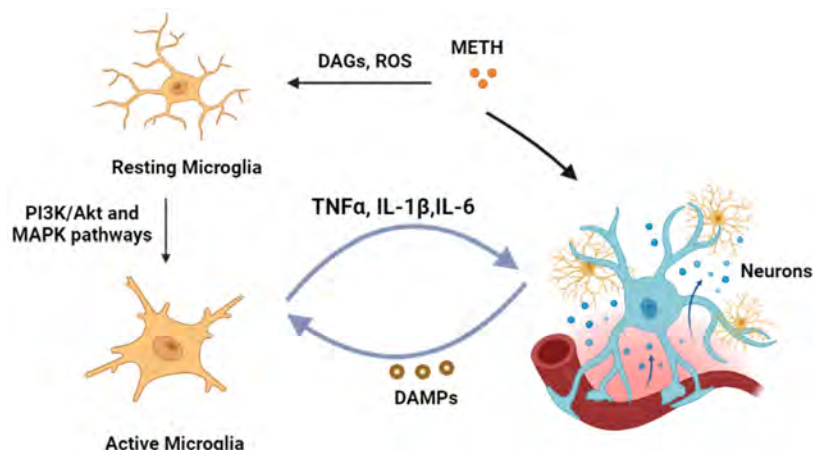


Fig. 2: Illustrating diagram of METH-induced neuro-inflammation
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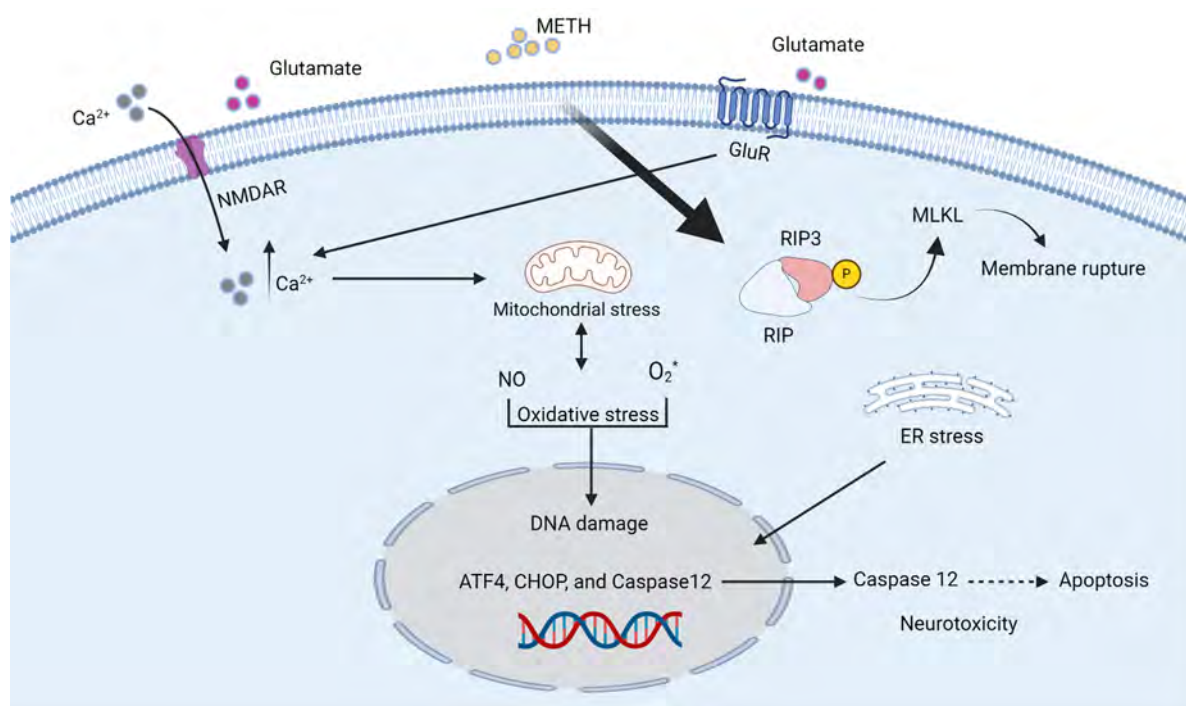


Fig. 3: Illustrating diagram of oxidative stress, mitochondrial stress, ER stress, membrane rupture and apoptosis involved in METH-induced neurotoxicity
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Methamphetamine-induced mitochondrial toxicity:

Mitochondrial toxicity is a phenomenon that arises as a side effect of some medications in which the mitochondria of a body's cells become damaged or decrease considerably in number⁵³. In METH-induced mitochondrial toxicity, the lipophilic nature of METH makes it easy to enter cell membranes, including mitochondria. Inside the mitochondria,

METH impairs their function and leads to neuropathological effects in the brain⁵⁰. According to Lenzi *et al.*⁵⁴, METH-induced mitochondrial changes are associated with a decrease in fission/mitophagy proteins Fis1 and DRP1 and an increase in Pink1 and Parkin *in situ*, indicating that mitochondria are targets of METH-induced toxicity. Another study by Abdullah *et al.*⁵⁵ demonstrated that METH self-administration

has a significant impact on mitochondrial ultrastructure, OXPHOS SCs assembly and redox activity, as well as enhanced PDH activity, which could explain reported cellular dysfunction. One of the key effects of METH-induced mitochondrial toxicity is the reduction in mitochondrial respiratory chain complex activity. This reduction impairs the production of ATP, the cell's primary energy source. Additionally, METH exposure increases the production of ROS and promotes oxidative stress within the mitochondria⁵⁶. The elevated ROS levels can cause damage to mitochondrial components and further impair mitochondrial function⁵⁷. The METH exposure also affects mitochondrial dynamics, specifically leading to an increase in mitochondrial fission. This imbalance between fusion and fission processes disrupts the normal structure and function of mitochondria, contributing to their fragmentation. Mitochondrial fragmentation is associated with apoptotic cell death, as it releases pro-apoptotic proteins such as cytochrome c from the mitochondria into the cytosol⁵⁸. Furthermore, METH exposure alters the expression of proteins involved in mitochondrial biogenesis, such as Nuclear Respiratory Factor 1 (NRF1), Peroxisome Proliferator-Activated Receptor Gamma Coactivator-1 α (PGC1 α) and Mitochondrial Transcription Factor A (TFAM). These changes negatively impact mitochondrial biogenesis and impair the overall health of

mitochondria⁵⁹. Moreover, METH can induce neuronal programmed necrosis through the activation of signaling pathways involving the activation of RIP3 leading to the phosphorylation of Mixed Linkage Domain-Like Protein (MLKL), which disrupts cell membranes and promotes necrotic cell death (Fig. 3). This process involves the formation of a complex between RIP3 and RIP1 and ultimately results in mitochondrial damage and neuronal necrosis⁶⁰. Recently, Graves *et al.*⁶¹ showed that METH therapy increased both axonal and somatic mitochondrial oxidative stress in substantia nigra pars compacta dopaminergic neurons, which is associated with a small but substantial increase in firing frequency and resulted in degeneration after drug withdrawal⁶¹.

TREATMENT OF METH-INDUCED NEUROTOXICITY

Several techniques have been developed to achieve effective pharmacological output in the treatment of METH-induced neurotoxicity. The processes of neurotoxicity induction serve as the basis for treatment options. The most recent studies on strategies used in the treatment of METH-induced neurotoxicity were summarized as shown in Table 1.

Table 1: Summary of recent studies (the last five years) on METH-induced neurotoxicity and treatment strategies in different experimental models

Treatment agent	Type of study	Experimental model	References
Calcitriol	<i>In vivo</i>	Rat	Cass and Peters ²⁶
Mesenchymal stem cells with antioxidant compound H-290/51	<i>In vivo</i>	Rat	Lafuente <i>et al.</i> ⁶²
α -Pinene	<i>In vivo</i>	Mice	Lee <i>et al.</i> ⁶³
Thiopentamide	<i>In vivo</i>	Mice	Luo <i>et al.</i> ⁶⁴
Thyroid hormones	<i>In vivo</i>	Primary hippocampal neurons	Tamijani <i>et al.</i> ⁶⁵
Erythropoietin	<i>In vivo</i>	Rat	Garmabi <i>et al.</i> ⁶⁶
Melatonin	<i>In vivo</i>	Human neuroblastoma cells	Nopparat <i>et al.</i> ⁶⁷
	<i>In vivo</i>	Rat	Kraiwananapirom <i>et al.</i> ⁶⁸ , Hein <i>et al.</i> ⁶⁹ and Namyen <i>et al.</i> ⁷⁰
	<i>In vivo</i>	Mouse hippocampus	Lwin <i>et al.</i> ⁷¹
Icariside II	<i>In vivo</i>	Mice	Huang <i>et al.</i> ⁷²
Cinnamaldehyde	<i>In vivo</i>	PC12 Cells	Saeed <i>et al.</i> ⁷³ and Rashidi <i>et al.</i> ⁷⁴
Curcumin	<i>In vivo</i>	Rat	Hadizadeh-Bazaz <i>et al.</i> ⁷⁵ ,
	<i>In vivo</i>	PC12 cell line	Ottonelli <i>et al.</i> ⁷⁶ and Ryskalin <i>et al.</i> ⁷⁷
6,7,4'-trihydroxyflavanone	<i>In vivo</i>	SH-SY5y Cells	Lee and Jeong ⁷⁸
Aromadendrin	<i>In vivo</i>	SH-SY5y cells	Lee <i>et al.</i> ⁷⁹
Resveratrol	<i>In vivo</i>	Mice	Zeng <i>et al.</i> ⁸⁰
Rosmarinic acid	<i>In vivo</i>	Zebrafish	Shahrestani <i>et al.</i> ⁸¹
Thymoquinone	<i>In vivo</i>	Mice	Roohbakhsh <i>et al.</i> ⁸²
Tertiary butylhydroquinone	<i>In vivo</i>	Rat	Meng <i>et al.</i> ⁸³
α -Synuclein	<i>In vivo</i>	Mice	Ding <i>et al.</i> ⁸⁴
Gastrodin	<i>In vitro</i>	Primary cortex neuronal culture	Ma <i>et al.</i> ⁸⁵
	<i>In vivo</i>	SH-SY5y cells	Yang <i>et al.</i> ⁸⁶
Lupenone	<i>In vivo</i>	SH-SY5y Cells	Lee <i>et al.</i> ⁸⁷
Tea polyphenols	<i>In vivo</i>	PC12 Cells	Ru <i>et al.</i> ⁸⁸
Obestatin	<i>In vivo</i>	PC12 cells	Foroughi <i>et al.</i> ⁸⁹
Apelin-13	<i>In vivo</i>	PC12 Cells	Foroughi <i>et al.</i> ⁹⁰

Anti-oxidative stress therapy: A lot of investigations have found that oxidative stress is a major source of METH-induced neurotoxicity. The METH-induced dopaminergic cell death occurs through boosting ROS generation and decreasing cellular ATP levels. The METH inhibits DA reuptake, causing auto-oxidation, which results in the generation of different ROS, superoxide and nitrogen radicals, causing cell death³⁵. Various pharmacotherapies have been investigated in a search for successful therapeutic techniques that will protect cells from the oxidative stress caused by METH. A variety of antioxidants have been shown to have anti-neurotoxic actions against METH.

According to Huang *et al.*⁹¹ vitamin C protects cortical cells from the neurotoxicity caused by METH through the attenuation of the expression of cleaved caspase-3 and the reduction in the number of METH-induced apoptotic cells. The rosmarinic acid combined with ZnO nanoparticles showed high potential in counteracting the METH-mediated elevation in caspase-3 mRNA level in zebrafish⁸¹. Furthermore, Kish *et al.*⁹² suggested that DA replenishment could potentially reverse the METH induced neurotoxicity.

Non-enzymatic antioxidants such as flavonoids have been proven to be beneficial in attenuation of METH-induced neurotoxicity. For example, anthocyananine protects neurons by avoiding cell death⁹³. According to Ru *et al.*⁸⁸ findings, tea polyphenol supplementation may be a useful dietary preventive approach for METH-induced neurotoxicity and neurodegenerative diseases. Also, according to Foroughi *et al.*⁹⁰, apelin-13, an endogenous ligand, may be able to lessen the neurotoxicity caused by METH by reducing oxidative damage, apoptotic and autophagic cell death. A further investigation was planned to find out if the antioxidant Tertiary Butylhydroquinone (TBHQ) may lessen the neurotoxicity caused by METH in male Wistar rats. The findings show that TBHQ can improve the antioxidative stress and antiapoptotic actions of phosphatidylinositol 3-kinase and nuclear factor erythroid 2-related factor-2, which can reduce the ROS and apoptosis caused by METH⁸³. Ma *et al.*⁸⁵ investigate gastrodin's neuroprotective effects against METH-induced neurotoxicity and show that it regulates the cAMP/PKA/CREB signaling pathway Cinnamaldehyde, a naturally occurring flavonoid found in the plant *Cinnamomum cassia*, similarly shown neuroprotective benefits against METH-induced cytotoxicity^{73,74}. Thymoquinone, a significant component of the plant *Nigella sativa*, has also been demonstrated to protect against METH toxicity⁸². Also, Lee and Jeong⁷⁸ reported that 6,7,4'-trihydroxyflavanone, a compound isolated from the *Dalbergia odorifera* plant

showed protective effects against METH-induced neurotoxicity. In addition, Lee *et al.*⁷⁹ found that aromadendrin, isolated from *Chionanthus retusus*, protected cells from METH-induced neurotoxicity via modulating ER stress and the PI3K/AKT/mTOR pathways. Also, Zeng *et al.*⁸⁰ demonstrated that pretreatment with natural polyphenol resveratrol (10 or 100 mg/kg) significantly improves memory impairments by preventing METH-induced oxidative damage and apoptosis in mice. Lafuente *et al.*⁶² recently demonstrated that nano delivery of H-290/51 with mesenchymal stem cells significantly increased cerebral blood flow and decreased blood-brain barrier breakdown, edema development and brain pathology after METH exposure in a hot environment. Thioperamide, according to Luo *et al.*⁶⁴ has a neuroprotective effect against METH-induced cognitive impairment in rats and toxicity in HT22 cells via the raf-MEK-ERK signaling pathway.

According to Tamijani *et al.*⁶⁵, thyroid hormones activate the cell surface receptors for integrin $\alpha v \beta 3$, which prevents METH-induced apoptosis in primary hippocampal neurons. To examine erythropoietin's (EPO) potential defense against METH neurotoxicity in male Wistar rats. Garmabi *et al.*⁶⁶ EPO showed significant neuroprotective effects on METH neurotoxicity due to its anti-inflammatory, antioxidant and antiapoptotic potential.

Anti-excitotoxicity therapy: The METH excitotoxicity is induced by increased Glu release, which activates NMDA receptors and other Glu receptors (Fig. 3), resulting in a high calcium ion influx and activation of a variety of cellular processes. As a result, these receptors are regarded as promising targets for METH-induced excitotoxicity and have sparked considerable interest in the development of targeted treatments. Neuropeptide Y is neuroprotective in a variety of disease situations. Baptista *et al.*⁹⁴ demonstrated that activating Y1 or Y2 receptors prevents METH-induced cell death and that the Y1 subtype is responsible for suppressing METH-induced neuronal differentiation. Melatonin, a pineal hormone, may protect against oxidative stress and regulate calcium ion intracellular transport in the CNS. Many recent studies reported the protective effect of melatonin on METH-induced changes in the expression levels of NMDA receptor subunits as well as Ca^{2+} -dependent protein kinase in human and experimental animals^{67,68}. Nopparat *et al.*⁶⁷ found that melatonin can help reduce amyloid deposits inside the neuron in a cellular model of METH-induced toxicity. Many researchers have discovered that N-acetylcysteine, a precursor of the antioxidant glutathione, exhibits an

antioxidant effect and reduces Glu excitatory toxicity in animals treated with METH^{95,96}. Also, dizocilpine, a powerful non-competitive NMDA receptor inhibitor, protects against METH-induced toxicity by inhibiting NMDA receptor overactivity downstream of DA release¹⁰. In addition, topiramate, an anticonvulsant, has a greater ability to reduce METH-induced excitatory toxicity by antagonizing numerous Glu receptors and inhibiting carbonic anhydrase¹⁰. It has been discovered that Levo-tetrahydropalmatine, an active component of herbal plant species from the genera *Corydalis* and *Stephania*, is neuroprotective against METH-induced neurotoxicity⁷.

Anti-mitochondrial toxicity therapy: Increasing mitochondrial activity may be beneficial to alleviate cellular resilience and neuronal plasticity impairments which are linked to a number of neuropsychiatric illnesses. In a study by Bachmann *et al.*⁹⁷, demonstrated that SH-SY5Y cells protected against METH-induced mitochondrial damage when treated with lithium or valproate in a long-term therapy. Park *et al.*⁹⁸ showed that METH-induced translocation of NF- κ B/STAT3 and ERK phosphorylation and proteolytic cleavage of caspase-3 were significantly reduced by asiatic acid. According to the results of Lee *et al.*⁸⁷, lupenone, naturally occurring triterpenoids, can prevent METH-induced neuronal death in SH-SY5y cells by blocking the PI3K/Akt pathway. According to Ru *et al.*⁸⁸, peptide hormone obestatin may be able to lessen METH-induced neurotoxicity in PC12 cells by reducing apoptotic and autophagic responses. In addition, Li *et al.*⁹⁹ investigated oxytocin's neuroprotective properties against METH-induced neuronal injury in rats. They discovered that pre-incubation with oxytocin prevents the damage to hippocampus neurons caused by METH intoxication. In addition, Chen *et al.*¹⁰⁰ reported that quercetin exhibited antipsychotic efficacy by modulating mitochondrial function and neuro-inflammation, implying that it has the potential to be further developed as a treatment in METH-induced anxiety. Recently, METH-induced lipid peroxidation was substantially reduced by alpha-Pinene via activation of nuclear factor-erythroid 2-related factor, which up-regulated antioxidant enzymes⁶³.

Anti-neuroinflammation therapy: Neuroinflammation is a primary cause of neurotoxicity caused by METH intoxication and is considered to be the most probable outcome of METH-induced damage. There are multiple endpoints that show METH-induced injury because there are many pathways and mediators for neuroinflammation¹⁰¹. The primary cause of METH's neurotoxicity is the inflammatory response brought

on by activated microglia. According to Sekine *et al.*¹⁰², METH users may experience reactive microgliosis in their brains as a result of continuous self-administration of the drug. Zhang *et al.*¹⁰³ reported that minocycline notably attenuated the rise in extracellular DA levels following multiple METH injections. They suggested that a number of symptoms linked to METH exposure could be treated by using minocycline to inactivate microglia.

The use of minocycline and modafinil after multiple METH injections attenuates the rise in extracellular DA and reduced METH-induced activation of microglial in brain cells, respectively. It results in the release of neuroinflammation¹⁰⁴.

Modafinil, a medication that improves cognition, also reduces METH-induced activation of microglial in brain cells, lowering the risk of neuroinflammation¹⁰⁴. White *et al.*⁴⁴ found that anti-METH/AMP mAb4G9 protected rat brains from METH-induced neuroinflammation. In another study by Kraiwattanapirom *et al.*⁶⁸, tested the anti-neuroinflammation potential of melatonin by pretreating with 10 mg/kg of melatonin before administering METH. This treatment was able to decrease an increase in Neuron-Glia2 levels and activation in microglia and astrocytes. Furthermore, Namyen *et al.*⁷⁰ demonstrated the protective effects of melatonin against METH neurotoxic profiles, which include reactive gliosis, increased upregulation of primary pro-inflammatory cytokines, activation of neuroinflammatory signaling and suppression of anti-oxidative signaling. These characteristics may exacerbate structural impairment of the blood brain barrier. A recent study conducted by Hadizadeh-Bazaz *et al.*⁷⁵ reported that curcumin, the main component of *Curcuma longa*, could increase anti-oxidative enzymes and decrease malondialdehyde in mice neurons injected with METH. They also reported that curcumin could reduce the TNF- α levels to attenuate the neuroinflammation induced by METH. Furthermore, Lwin *et al.*⁷¹ demonstrated that the administration of melatonin caused the METH-induced markers to return to control levels. Thus, they conclude that melatonin may be utilized as an anti-inflammatory medication to treat METH use disorder in people⁶⁹. More recently, Huang *et al.*⁷² administered Icariside II (ICS), the primary active component of the traditional Chinese medicine Epimedium. It results in the activation of Keap1-Nrf2 pathway thereby reducing glial cell activation and mitigating neurotoxicity. Also, Ottonelli *et al.*⁷⁶ found that cerebral edema, neuronal damage and blood brain barrier leakage in rat brains were dramatically reduced by curcumin alone (50 mg/kg, i.v.) and curcumin delivered via TiO₂ nanowired administration (25 mg/kg, i.v.).

CONCLUSION

Methamphetamine (METH) is a highly addictive drug that affects brain neurotransmitters, leading to severe physical and mental health issues. Short-term effects include euphoria, increased energy, decreased appetite and elevated heart rate and blood pressure, followed by a prolonged crash. Long-term use results in significant health problems, including neuro-inflammation and neuronal injury due to excessive dopaminergic activity. The METH's lipophilic nature allows it to cross the blood-brain barrier, causing neurotoxicity through interactions with cell surface receptors. Key factors in METH-induced neurotoxicity include dopamine depletion, oxidative stress and activation of microglia and astrocytes. While the exact chemical processes remain unclear, research focuses on mitigating these effects. Current studies explore targeted therapies like gene therapy, immunotherapy and nanoparticle-based treatments. This review highlights recent findings on METH-induced neurotoxicity and treatment strategies, emphasizing the need for further research to develop effective therapeutics.

SIGNIFICANCE STATEMENT

This review provides a comprehensive analysis of the neurotoxic effects of Methamphetamine (METH) use, highlighting the underlying molecular mechanisms and the different treatment strategies. By elucidating the pathways of METH-induced neurotoxicity and exploring innovative treatment strategies, this research aims to contribute to the development of more effective therapeutic interventions. Understanding these mechanisms is crucial for mitigating the public health crisis posed by METH addiction and improving outcomes for affected individuals.

REFERENCES

1. Yasaei, R. and A. Saadabadi, 2023. Methamphetamine. StatPearls, Treasure Island.
2. Bonson, K.R., T. Dalton and D. Chiapperino, 2019. Scheduling synthetic cathinone substances under the controlled substances act. *Psychopharmacology*, 236: 845-860.
3. Salani, D., B. Valdes, J. de Santis and M. Zdanowicz, 2020. Back with a vengeance: The reappearance of methamphetamine and its implications for health care providers. *J. Nurse Pract.*, 16: 483-488.
4. Sahab Uddin, M., M.A. Sufian, M.T. Kabir, M.F. Hossain and M. Nasrullah *et al.*, 2017. Amphetamines: Potent recreational drug of abuse. *J. Addict. Res. Ther.*, Vol. 8. 10.4172/2155-6105.1000330.
5. Vandenbark, A.A., R. Meza-Romero, G. Benedek and H. Offner, 2019. A novel neurotherapeutic for multiple sclerosis, ischemic injury, methamphetamine addiction, and traumatic brain injury. *J. Neuroinflammation*, Vol. 16. 10.1186/s12974-018-1393-0.
6. Moszczynska, A. and S.P. Callan, 2017. Molecular, behavioral, and physiological consequences of methamphetamine neurotoxicity: Implications for treatment. *J. Pharmacol. Exp. Ther.*, 362: 474-488.
7. Liu, L., M. Liu, W. Zhao, Y.L. Zhao and Y. Wang, 2021. Tetrahydropalmatine regulates BDNF through TrkB/CAM interaction to alleviate the neurotoxicity induced by methamphetamine. *ACS Chem. Neurosci.*, 12: 3373-3386.
8. Hossain, M.K., M. Hassanzadeganroudsari, K. Nurgali and V. Apostolopoulos, 2020. Vaccine development against methamphetamine drug addiction. *Expert Rev. Vaccines*, 19: 1105-1114.
9. Kazemifar, A.M., H. Solhi, D. Badakhshan and M. Eidi, 2011. Analysis of the drug crystals discovered by police 23 Sep 2008 to 20 Mar 2009 in Arak. *ARPN J. Eng. Appl. Sci.*, 6: 48-51.
10. Yang, X., Y. Wang, Q. Li, Y. Zhong and L. Chen *et al.*, 2018. The main molecular mechanisms underlying methamphetamine-induced neurotoxicity and implications for pharmacological treatment. *Front. Mol. Neurosci.*, Vol. 11. 10.3389/fnmol.2018.00186.
11. Ahmed, B., F.N. Yousaf, M. Saud and A. Ahmad, 2020. Youth at risk: The alarming issue of drug addiction in academic institutions in Pakistan. *Children Youth Serv. Rev.*, Vol. 118. 10.1016/j.childyouth.2020.105385.
12. Stoneberg, D.M., R.K. Shukla and M.B. Magness, 2018. Global methamphetamine trends: An evolving problem. *Int. Crim. Justice Rev.*, 28: 136-161.
13. Turner, C., D. Chandrakumar, C. Rowe, G.M. Santos, E.D. Riley and P.O. Coffin, 2018. Cross-sectional cause of death comparisons for stimulant and opioid mortality in San Francisco, 2005-2015. *Drug Alcohol Depend.*, 185: 305-312.
14. Al-Asmari, A.I., 2021. Methamphetamine-related postmortem cases in Jeddah, Saudi Arabia. *Forensic Sci. Int.*, Vol. 321. 10.1016/j.forsciint.2021.110746.
15. Erdoğan, A. and S. Karabulut, 2024. Frequency of methamphetamine use in patients admitted to an addiction center and clinical factors associated with methamphetamine use. *J. Drug Issues*, 10.1177/00220426241263256.
16. Guerin, A.A., T. Bridson, H.M. Plapp and G. Bedi, 2023. A systematic review and meta-analysis of health, functional, and cognitive outcomes in young people who use methamphetamine. *Neurosci. Biobehav. Rev.*, Vol. 153. 10.1016/j.neubiorev.2023.105380.
17. Chansaengetch, N., W. Worasuwannarak and S. Worawichawong, 2023. Methamphetamine-induced profound rhabdomyolysis and myoglobin cast nephropathy: A case report and a literature review. *J. Forensic Legal Med.*, Vol. 96. 10.1016/j.jflm.2023.102530.

18. Al-Waheeb, S., N. Al-Omair and A. Mahdi, 2021. Patterns of drug overdose deaths in Kuwait from 2014 to 2018. *Public Health Pract.*, Vol. 2. 10.1016/j.puhip.2021.100181.
19. Thrash, B., K. Thiruchelvan, M. Ahuja, V. Suppiramaniam and M. Dhanasekaran, 2009. Methamphetamine-induced neurotoxicity: The road to Parkinson's disease. *Pharmacol. Rep.*, 61: 966-977.
20. Sambo, D.O., M. Lin, A. Owens, J.J. Lebowitz and B. Richardson *et al.*, 2017. The sigma-1 receptor modulates methamphetamine dysregulation of dopamine neurotransmission. *Nat. Commun.*, Vol. 8. 10.1038/s41467-017-02087-x.
21. Cruickshank, C.C. and K.R. Dyer, 2009. A review of the clinical pharmacology of methamphetamine. *Addiction*, 104: 1085-1099.
22. Blum, K., J.L. Cadet and M.S. Gold, 2021. Psychostimulant use disorder emphasizing methamphetamine and the opioid-dopamine connection: Digging out of a hypodopaminergic ditch. *J. Neurol. Sci.*, Vol. 420. 10.1016/j.jns.2020.117252.
23. Moratalla, R., A. Khairnar, N. Simola, N. Granado and J.R. García-Montes *et al.*, 2017. Amphetamine-related drugs neurotoxicity in humans and in experimental animals: Main mechanisms. *Prog. Neurobiol.*, 155: 149-170.
24. Kohno, M., J. Link, L.E. Dennis, H. McCready, M. Huckans, W.F. Hoffman and J.M. Loftis, 2019. Neuroinflammation in addiction: A review of neuroimaging studies and potential immunotherapies. *Pharmacol. Biochem. Behav.*, 179: 34-42.
25. Chao, J., Y. Zhang, L. Du, R. Zhou, X. Wu, K. Shen and H. Yao, 2017. Molecular mechanisms underlying the involvement of the sigma-1 receptor in methamphetamine-mediated microglial polarization. *Sci. Rep.*, Vol. 7. 10.1038/s41598-017-11065-8.
26. Cass, W.A. and L.E. Peters, 2023. Calcitriol protects against reductions in striatal serotonin in rats treated with neurotoxic doses of methamphetamine. *Neurochem. Int.*, Vol. 169. 10.1016/j.neuint.2023.105590.
27. Pizzino, G., N. Irrera, M. Cucinotta, G. Pallio and F. Mannino *et al.*, 2017. Oxidative stress: Harms and benefits for human health. *Oxid. Med. Cell. Longevity*, Vol. 2017. 10.1155/2017/8416763.
28. Checa, J. and J.M. Aran, 2020. Reactive oxygen species: Drivers of physiological and pathological processes. *J. Inflammation Res.*, 13: 1057-1073.
29. Copley, J.N., M.L. Fiorello and D.M. Bailey, 2018. 13 reasons why the brain is susceptible to oxidative stress. *Redox Biol.*, 15: 490-503.
30. Horner, K.A., Y.E. Gilbert and S.D. Cline, 2011. Widespread increases in malondialdehyde immunoreactivity in dopamine-rich and dopamine-poor regions of rat brain following multiple, high doses of methamphetamine. *Front. Syst. Neurosci.*, Vol. 5. 10.3389/fnsys.2011.00027.
31. Lazzeri, G., F. Biagioni, F. Fulceri, C.L. Busceti and M.C. Scavuzzo *et al.*, 2018. mTOR modulates methamphetamine-induced toxicity through cell clearing systems. *Oxid. Med. Cell. Longevity*, Vol. 2018. 10.1155/2018/6124745.
32. Sundar, V., T. Ramasamy, M. Doke and T. Samikkannu, 2022. Psychostimulants influence oxidative stress and redox signatures: The role of DNA methylation. *Redox Rep.*, 27: 53-59.
33. Rumpf, J.J., J. Albers, C. Fricke, W. Mueller and J. Classen, 2017. Structural abnormality of substantia nigra induced by methamphetamine abuse. *Mov. Disord.*, 32: 1784-1788.
34. Ding, J., Y. Lian, Y. Meng, Y. He, H. Fan, C. Li and P. Qiu, 2020. The effect of α -synuclein and Tau in methamphetamine induced neurotoxicity *in vivo* and *in vitro*. *Toxicol. Lett.*, 319: 213-224.
35. DiSabato, D. J., N. Quan and J.P. Godbout, 2016. Neuroinflammation: The devil is in the details. *J. Neurochem.*, 139: 136-153.
36. Shrestha, P., N. Katila, S. Lee, J.H. Seo, J.H. Jeong and S. Yook, 2022. Methamphetamine induced neurotoxic diseases, molecular mechanism, and current treatment strategies. *Biomed. Pharmacother.*, Vol. 154. 10.1016/j.biopha.2022.113591.
37. Thomas, D.M., D.M. Francescutti-Verbeem and D.M. Kuhn, 2008. The newly synthesized pool of dopamine determines the severity of methamphetamine-induced neurotoxicity. *J. Neurochem.*, 105: 605-616.
38. Smith, J.A., A. Das, S.K. Ray and N.L. Banik, 2012. Role of pro-inflammatory cytokines released from microglia in neurodegenerative diseases. *Brain Res. Bull.*, 87: 10-20.
39. Ambrogini, P., P. Torquato, D. Bartolini, M.C. Albertini and D. Lattanzi *et al.*, 2019. Excitotoxicity, neuroinflammation and oxidant stress as molecular bases of epileptogenesis and epilepsy-derived neurodegeneration: The role of vitamin E. *Biochim. Biophys. Acta Mol. Basis Dis.*, 1865: 1098-1112.
40. Yamamoto, B.K., A. Moszczynska and G.A. Gudelsky, 2010. Amphetamine toxicities: Classical and emerging mechanisms. *Ann. N. Y. Acad. Sci.*, 1187: 101-121.
41. Du, S.H., D.F. Qiao, C.X. Chen, S. Chen and C. Liu *et al.*, 2017. Toll-like receptor 4 mediates methamphetamine-induced neuroinflammation through caspase-11 signaling pathway in astrocytes. *Front. Mol. Neurosci.*, Vol. 10. 10.3389/fnmol.2017.00409.
42. Billod, J.M., A. Lacetera, J. Guzmán-Caldentey and S. Martín-Santamaría, 2016. Computational approaches to toll-like receptor 4 modulation. *Molecules*, Vol. 21. 10.3390/molecules21080994.
43. Shen, Y., H. Qin, J. Chen, L. Mou and Y. He *et al.*, 2016. Postnatal activation of TLR4 in astrocytes promotes excitatory synaptogenesis in hippocampal neurons. *J. Cell Biol.*, 215: 719-734.

44. White, S.J., H.P. Hendrickson, W.T. Atchley, E.M. Laurenzana, W.B. Gentry, D.K. Williams and S.M. Owens, 2014. Treatment with a monoclonal antibody against methamphetamine and amphetamine reduces maternal and fetal rat brain concentrations in late pregnancy. *Drug Metab. Dispos.*, 42: 1285-1291.
45. Chen, J., M. Rusnak, P.J. Lombroso and A. Sidhu, 2009. Dopamine promotes striatal neuronal apoptotic death via ERK signaling cascades. *Eur. J. Neurosci.*, 29: 287-306.
46. Liu, X., P.S. Silverstein, V. Singh, A. Shah, N. Qureshi and A. Kumar, 2012. Methamphetamine increases LPS-mediated expression of IL-8, TNF- α and IL-1 β in human macrophages through common signaling pathways. *PLoS ONE*, Vol. 7. 10.1371/journal.pone.0033822.
47. Kobeissy, F.H., Z. Shakkour, S. El Hayek, W. Mohamed, M.S. Gold and K.K.W. Wang, 2022. Elevation of pro-inflammatory and anti-inflammatory cytokines in rat serum after acute methamphetamine treatment and traumatic brain injury. *J. Mol. Neurosci.*, 72: 158-168.
48. Dong, X.X., Y. Wang and Z.H. Qin, 2009. Molecular mechanisms of excitotoxicity and their relevance to pathogenesis of neurodegenerative diseases. *Acta Pharmacol. Sin.*, 30: 379-387.
49. Vishnoi, S., S. Raisuddin and S. Parvez, 2016. Glutamate excitotoxicity and oxidative stress in epilepsy: Modulatory role of melatonin. *J. Environ. Pathol. Toxicol. Oncol.*, 35: 365-374.
50. Tseng, E.E., M.V. Brock, M.S. Lange, J.C. Troncoso and M.E. Blue *et al.*, 2010. Glutamate excitotoxicity mediates neuronal apoptosis after hypothermic circulatory arrest. *Ann. Thoracic Surg.*, 89: 440-445.
51. Valian, N., A. Ahmadiani and L. Dargahi, 2017. Escalating methamphetamine regimen induces compensatory mechanisms, mitochondrial biogenesis, and GDNF expression, in Substantia Nigra. *J. Cell. Biochem.*, 118: 1369-1378.
52. Halpin, L.E., N.A. Northrop and B.K. Yamamoto, 2014. Ammonia mediates methamphetamine-induced increases in glutamate and excitotoxicity. *Neuropsychopharmacology*, 39: 1031-1038.
53. Meyer, J.N., J.H. Hartman and D.F. Mello, 2018. Mitochondrial toxicity. *Toxicol. Sci.*, 162: 15-23.
54. Lenzi, P., F. Biagioni, C.L. Busceti, G. Lazzeri and M. Polzella *et al.*, 2022. Alterations of mitochondrial structure in methamphetamine toxicity. *Int. J. Mol. Sci.*, Vol. 23. 10.3390/ijms23168926.
55. Abdullah, C.S., N.S. Remex, R. Aishwarya, S. Nitu and G.K. Kolluru *et al.*, 2022. Mitochondrial dysfunction and autophagy activation are associated with cardiomyopathy developed by extended methamphetamine self-administration in rats. *Redox Biol.*, Vol. 58. 10.1016/j.redox.2022.102523.
56. Dawson, T.M. and V.L. Dawson, 2017. Mitochondrial mechanisms of neuronal cell death: Potential therapeutics. *Annu. Rev. Pharmacol. Toxicol.*, 57: 437-454.
57. Misrani, A., S. Tabassum and L. Yang, 2021. Mitochondrial dysfunction and oxidative stress in Alzheimer's disease. *Front. Aging Neurosci.*, Vol. 13. 10.3389/fnagi.2021.617588.
58. Ren, L., X. Chen, X. Chen, J. Li, B. Cheng and J. Xia, 2020. Mitochondrial dynamics: Fission and fusion in fate determination of mesenchymal stem cells. *Front. Cell Dev. Biol.*, Vol. 8. 10.3389/fcell.2020.580070.
59. Shen, Y., L. Wu, J. Wang, X. Wu and X. Zhang, 2018. The role of mitochondria in methamphetamine-induced inhibitory effects on osteogenesis of mesenchymal stem cells. *Eur. J. Pharmacol.*, 826: 56-65.
60. Zhao, X., J. Lu, X. Chen, Z. Gao and C. Zhang *et al.*, 2021. Methamphetamine exposure induces neuronal programmed necrosis by activating the receptor-interacting protein kinase 3-related signalling pathway. *FASEB J.*, Vol. 35. 10.1096/fj.202100188R.
61. Graves, S.M., S.E. Schwarzschild, R.A. Tai, Y. Chen and D.J. Surmeier, 2021. Mitochondrial oxidant stress mediates methamphetamine neurotoxicity in substantia nigra dopaminergic neurons. *Neurobiol. Dis.*, Vol. 156. 10.1016/j.nbd.2021.105409.
62. Lafuente, J.V., A. Sharma, L. Feng, D.F. Muresanu and A. Nozari *et al.*, 2023. Retracted Chapter: Nanowired Delivery of Mesenchymal Stem Cells with Antioxidant Compound H-290/51 Reduces Exacerbation of Methamphetamine Neurotoxicity in Hot Environment. In: *Progress in Nanomedicine in Neurologic Diseases*, Sharma, H.S. and A. Sharma (Eds.), Springer, Cham, Switzerland, ISBN: 978-3-031-32997-5, pp: 317-318.
63. Lee, C., J.H. Jang and G.H. Park, 2023. α -Pinene attenuates methamphetamine-induced conditioned place preference in C57BL/6 mice. *Biomol. Ther.*, 31: 411-416.
64. Luo, H., X. Li, R. Fan, Y. Ruan and L. Qian *et al.*, 2023. Neuroprotective effect of histamine H3 receptor blockade on methamphetamine-induced cognitive impairment in mice. *Pharmacol. Biochem. Behav.*, Vol. 222. 10.1016/j.pbb.2022.173512.
65. Tamijani, S.M.S., E. Beirami, S. Dargahi, A. Ahmadiani and L. Dargahi, 2023. Neuroprotective effect of thyroid hormones on methamphetamine-induced neurotoxicity via cell surface receptors. *Neurosci. Lett.*, Vol. 794. 10.1016/j.neulet.2022.137009.
66. Garmabi, B., R. Mohaddes, F. Rezvani, F. Mohseni, H. Khastar and M. Khaksari, 2022. Erythropoietin improve spatial memory impairment following methamphetamine neurotoxicity by inhibition of apoptosis, oxidative stress and neuroinflammation in CA1 area of hippocampus. *J. Chem. Neuroanat.*, Vol. 124. 10.1016/j.jchemneu.2022.102137.

67. Nopparat, C., A. Boontor, J. Panmanee and P. Govitrapong, 2022. Melatonin attenuates methamphetamine-induced alteration of amyloid β precursor protein cleaving enzyme expressions via melatonin receptor in human neuroblastoma cells. *Neurotoxic. Res.*, 40: 1086-1095.
68. Kraiwattanapirom, N., P. Komlao, A. Harnpramukul, K. Promyo, S. Ngampramuan and B. Chetsawang, 2021. The neuroprotective role of melatonin against methamphetamine toxicity-induced neurotransmission dysregulation and cognitive deficits in rats. *Food Chem. Toxicol.*, Vol. 157. 10.1016/j.fct.2021.112610.
69. Hein, Z.M., N. Kraiwattanapirom, S. Mukda and B. Chetsawang, 2020. The induction of neuron-glia2 (NG2) expressing cells in methamphetamine toxicity-induced neuroinflammation in rat brain are averted by melatonin. *J. Neuroimmunol.*, Vol. 344. 10.1016/j.jneuroim.2020.577232.
70. Namyen, J., K. Permpoonputtana, C. Nopparat, J. Tocharus, C. Tocharus and P. Govitrapong, 2020. Protective effects of melatonin on methamphetamine-induced blood-brain barrier dysfunction in rat model. *Neurotoxic. Res.*, 37: 640-660.
71. Lwin, T., J.L. Yang, S. Ngampramuan, K. Viwatpinyo and P. Chanchaen *et al.*, 2021. Melatonin ameliorates methamphetamine-induced cognitive impairments by inhibiting neuroinflammation via suppression of the TLR4/MyD88/NF κ B signaling pathway in the mouse hippocampus. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry*, Vol. 111. 10.1016/j.pnpbp.2020.110109.
72. Huang, J., J. Ding, Z. Wang, Y. Li and Y. He *et al.*, 2022. Icariside II attenuates methamphetamine-induced neurotoxicity and behavioral impairments via activating the Keap1-Nrf2 pathway. *Oxid. Med. Cell. Longevity*, Vol. 2022. 10.1155/2022/8400876.
73. Saeed, M., A. Ghadiri, F. Hadizadeh, A. Attaranzadeh, M.S. Alavi and L. Etemad, 2018. Cinnamaldehyde improves methamphetamine-induced spatial learning and memory deficits and restores ERK signaling in the rat prefrontal cortex. *Iran. J. Basic Med. Sci.*, 21: 1316-1321.
74. Rashidi, R., S.A. Moallem, M. Moshiri, F. Hadizadeh and L. Etemad, 2021. Protective effect of cinnamaldehyde on METH-induced neurotoxicity in PC12 cells via inhibition of apoptotic response and oxidative stress. *Iran. J. Pharm. Res.*, 20: 135-143.
75. Hadizadeh-Bazaz, M., G. Vaezi, M. Khaksari and V. Hojati, 2021. Curcumin attenuates spatial memory impairment by anti-oxidative, anti-apoptosis, and anti-inflammatory mechanism against methamphetamine neurotoxicity in male Wistar rats: Histological and biochemical changes. *NeuroToxicology*, 84: 208-217.
76. Ottonelli, I., A. Sharma, B. Ruozzi, G. Tosi and J.T. Duskey *et al.*, 2023. Retracted Chapter: Nanowired Delivery of Curcumin Attenuates Methamphetamine Neurotoxicity and Elevates Levels of Dopamine and Brain-Derived Neurotrophic Factor. In: *Progress in Nanomedicine in Neurologic Diseases*, Sharma, H.S. and A. Sharma (Eds.), Springer, Cham, Switzerland, ISBN: 978-3-031-32997-5, pp: 385-386.
77. Ryskalin, L., S. Puglisi-Allegra, G. Lazzeri, F. Biagioni and C.L. Busceti *et al.*, 2021. Neuroprotective effects of curcumin in methamphetamine-induced toxicity. *Molecules*, Vol. 26. 10.3390/molecules26092493.
78. Lee, H.S. and G.S. Jeong, 2021. 6,7,4'-Trihydroxyflavanon mitigates methamphetamine-induced neurotoxicity in SH-SY5y cells via Nrf2/heme oxygenase-1 and PI3K/Akt/mTOR signaling pathways. *Molecules*, Vol. 26. 10.3390/molecules26092442.
79. Lee, H.S., E.N. Kim and G.S. Jeong, 2021. Aromadendrin protects neuronal cells from methamphetamine-induced neurotoxicity by regulating endoplasmic reticulum stress and PI3K/Akt/mTOR signaling pathway. *Int. J. Mol. Sci.*, Vol. 22. 10.3390/ijms22052274.
80. Zeng, Q., Q. Xiong, M. Zhou, X. Tian and K. Yue *et al.*, 2021. Resveratrol attenuates methamphetamine-induced memory impairment via inhibition of oxidative stress and apoptosis in mice. *J. Food Biochem.*, Vol. 45. 10.1111/jfbc.13622.
81. Shahrestani, V.N., M. Haddadi and A.R.S. Kermani, 2021. Behavioral and molecular analysis of antioxidative potential of rosmarinic acid against methamphetamine-induced augmentation of Casp3a mRNA in the zebrafish brain. *Basic Clin. Neurosci.*, 12: 243-254.
82. Roohbakhsh, A., M. Moshiri, A.S. Kakhki, M. Iranshahy, F. Amin and L. Etemad, 2021. Thymoquinone abrogates methamphetamine-induced striatal neurotoxicity and hyperlocomotor activity in mice. *Res. Pharm. Sci.*, 16: 391-399.
83. Meng, X., C. Zhang, Y. Guo, Y. Han and C. Wang *et al.*, 2020. TBHQ attenuates neurotoxicity induced by methamphetamine in the VTA through the Nrf2/HO-1 and PI3K/AKT signaling pathways. *Oxid. Med. Cell. Longevity*, Vol. 2020. 10.1155/2020/8787156.
84. Ding, J., S. Hu, Y. Meng, C. Li, J. Huang, Y. He and P. Qiu, 2020. Alpha-synuclein deficiency ameliorates chronic methamphetamine induced neurodegeneration in mice. *Toxicology*, Vol. 438. 10.1016/j.tox.2020.152461.
85. Ma, C.L., L. Li, G.M. Yang, Z.B. Zhang and Y.N. Zhao *et al.*, 2020. Neuroprotective effect of gastrodin in methamphetamine-induced apoptosis through regulating cAMP/PKA/CREB pathway in cortical neuron. *Hum. Exp. Toxicol.*, 39: 1118-1129.

86. Yang, G., X. Zeng, J. Li, C.K. Leung and D. Zhang *et al*, 2019. Protective effect of gastrodin against methamphetamine-induced autophagy in human dopaminergic neuroblastoma SH-SY5Y cells via the AKT/mTOR signaling pathway. *Neurosci. Lett.*, Vol. 707. 10.1016/j.neulet.2019.134287.
87. Lee, H.S., E.N. Kim and G.S. Jeong, 2020. Lupenone protects neuroblastoma SH-SY5Y cells against methamphetamine-induced apoptotic cell death via PI3K/Akt/mTOR signaling pathway. *Int. J. Mol. Sci.*, Vol. 21. 10.3390/ijms21051617.
88. Ru, Q., Q. Xiong, X. Tian, L. Chen, M. Zhou, Y. Li and C. Li, 2019. Tea polyphenols attenuate methamphetamine-induced neuronal damage in PC12 cells by alleviating oxidative stress and promoting DNA repair. *Front. Physiol.*, Vol. 10. 10.3389/fphys.2019.01450.
89. Foroughi, K., S. Jahanbani, M. Khaksari and A. Shayannia, 2020. Obestatin attenuated methamphetamine-induced PC12 cells neurotoxicity via inhibiting autophagy and apoptosis. *Hum. Exp. Toxicol.*, 39: 301-310.
90. Foroughi, K., M. Khaksari, M. Rahmati, F.S. Bitaraf and A. Shayannia, 2019. Apelin-13 protects PC12 cells against methamphetamine-induced oxidative stress, autophagy and apoptosis. *Neurochem. Res.*, 44: 2103-2112.
91. Huang, Y.N., L.Y. Yang, J.Y. Wang, C.C. Lai, C.T. Chiu and J.Y. Wang, 2017. L-Ascorbate protects against methamphetamine-induced neurotoxicity of cortical cells via inhibiting oxidative stress, autophagy, and apoptosis. *Mol. Neurobiol.*, 54: 125-136.
92. Kish, S.J., I. Boileau, R.C. Callaghan and J. Tong, 2017. Brain dopamine neurone 'damage': Methamphetamine users vs. Parkinson's disease-A critical assessment of the evidence. *Eur. J. Neurosci.*, 45: 58-66.
93. Primatanti, P.A. and I.M. Jawi, 2020. Anthocyanin as neuroprotector for methamphetamine-induced neurotoxicity. *Int. J. Health Med. Sci.*, 3: 11-16.
94. Baptista, S., A.R. Bento, J. Gonçalves, L. Bernardino and T. Summavielle *et al*, 2012. Neuropeptide Y promotes neurogenesis and protection against methamphetamine-induced toxicity in mouse dentate gyrus-derived neurosphere cultures. *Neuropharmacology*, 62: 2413-2423.
95. Scofield, M.D. and P.W. Kalivas, 2014. Astrocytic dysfunction and addiction: Consequences of impaired glutamate homeostasis. *Neuroscientist*, 20: 610-622.
96. McKetin, R., O.M. Dean, A.L. Baker, G. Carter, A. Turner, P.J. Kelly and M. Berk, 2017. A potential role for N-acetylcysteine in the management of methamphetamine dependence. *Drug Alcohol Rev.*, 36: 153-159.
97. Bachmann, R.F., Y. Wang, P. Yuan, R. Zhou and X. Li *et al*, 2009. Common effects of lithium and valproate on mitochondrial functions: Protection against methamphetamine-induced mitochondrial damage. *Int. J. Neuropsychopharmacol.*, 12: 805-822.
98. Park, J.H., Y.H. Seo, J.H. Jang, C.H. Jeong, S. Lee and B. Park, 2017. Asiatic acid attenuates methamphetamine-induced neuroinflammation and neurotoxicity through blocking of NF- κ B/STAT3/ERK and mitochondria-mediated apoptosis pathway. *J. Neuroinflammation*, Vol. 14. 10.1186/s12974-017-1009-0.
99. Li, C., H. Wang, M. Wang, C. Chen, F. Bai, M. Ban and C. Wu, 2021. Oxytocin attenuates methamphetamine-induced apoptosis *via* oxytocin receptor in rat hippocampal neurons. *Front. Pharmacol.*, Vol. 12. 10.3389/fphar.2021.639571.
100. Chen, F., J. Sun, C. Chen, Y. Zhang and L. Zou *et al*, 2022. Quercetin mitigates methamphetamine-induced anxiety-like behavior through ameliorating mitochondrial dysfunction and neuroinflammation. *Front. Mol. Neurosci.*, Vol. 15. 10.3389/fnmol.2022.829886.
101. Shaerzadeh, F., W.J. Streit, S. Heysieattalab and H. Khoshbouei, 2018. Methamphetamine neurotoxicity, microglia, and neuroinflammation. *J. Neuroinflammation*, Vol. 15. 10.1186/s12974-018-1385-0.
102. Sekine, Y., Y. Ouchi, G. Sugihara, N. Takei and E. Yoshikawa *et al*, 2008. Methamphetamine causes microglial activation in the brains of human abusers. *J. Neurosci.*, 28: 5756-5761.
103. Zhang, L., K. Kitaichi, Y. Fujimoto, H. Nakayama, E. Shimizu, M. Iyo and K. Hashimoto, 2006. Protective effects of minocycline on behavioral changes and neurotoxicity in mice after administration of methamphetamine. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry*, 30: 1381-1393.
104. Raineri, M., B. Gonzalez, B. Goitia, E. Garcia-Rill and I.N. Krasnova *et al*, 2012. Modafinil abrogates methamphetamine-induced neuroinflammation and apoptotic effects in the mouse striatum. *PLoS ONE*, Vol. 7. 10.1371/journal.pone.0046599.