

**PJN**

ISSN 1680-5194

PAKISTAN JOURNAL OF  
**NUTRITION**

**ANSI***net*

308 Lasani Town, Sargodha Road, Faisalabad - Pakistan  
Mob: +92 300 3008585, Fax: +92 41 8815544  
E-mail: [editorpjn@gmail.com](mailto:editorpjn@gmail.com)

## Microbial Quality of Formulated Infant Milk Powders

Imran Rashid Rajput<sup>1</sup>, M. Khaskheli<sup>2</sup>, S. Rao<sup>3</sup>, S.A. Fazlani<sup>1</sup>, Q.A. Shah<sup>1</sup> and G.B. Khaskheli<sup>2</sup>

<sup>1</sup>Faculty of Veterinary and Animal Sciences, University of Agriculture,  
Water and Marine Sciences, Uthal Balochistan, Pakistan

<sup>2</sup>Department of Dairy Technology, <sup>3</sup>Faculty of Animal Husbandry and Veterinary Sciences,  
Sindh Agriculture University, Tandojam, Pakistan

**Abstract:** Study was carried out to examine the microbiological quality of Infant formula milk powder. Total 60 of dried milk powders, 20 each of Group A (1-6), B (7-12) and C (13-18 months). Infant formula milk powders were purchased from Hyderabad, Sindh and evaluated for microbiological examination, like Total Viable Count (TVC), Enterobacteriaceae Count (EbC) and Yeasts and Moulds Count (YMC). Total viable count, ( $3.4 \times 10^3 \pm 5.0 \times 10^2$  cfu/g) were significantly higher than thermophilic count, ( $< 10 \pm 2.3 \times 10^1$  cfu/g) Thermophilic spores ( $1.4 \times 10^1 \pm 2.4 \times 10^1$  cfu/g) Enterobacteriaceae count, ( $< 5 \pm 1.0 \times 10^0$  cfu/g) and Yeasts and Moulds ( $< 5 \pm 1.0$  cfu/g) respectively in all samples of infant formula of milk powders. Total Viable Counts group A, B and C having non significant difference. Even incase of thermophilic thermophilic spores, enterobacteriaceae, yeast and moulds were non significant recorded. The obtained averaged results compared to Indian Standard Institution (ISI) values. In Group A total viable counts were (12.18 folds), Group B (14.70 folds) and in Group C (15.15folds) lower than the ISI standard. thermophilic counts averaged compared with ISI standards and results were found lower in Group A (12.5 folds), Group B (16.0 folds) and Group C (14.2 folds) as compared to Indian Standards Institution, thermophilic spores in Group A (7.14 folds), Group B (6.6 folds) and Group C (6.25 folds) as compared to ISI. Enterobacteriaceae were lower than ISI standards (33 folds) in Group A, (33 folds) Group B and (25 folds) in Group C. Yeast and Moulds were lower than the ISI standards Group A (33 folds), Group B (33 folds) and in Group C (25 folds). Although Total Viable Count were within the range of standard of specification of (ISI) and the counts of thermophilic Spores, enterobacteriaceae counts and yeast and moulds also indicates the hygienic condition of Infant formula milk powders without risk level for human health.

**Key words:** Infant milk powder, enterobacteriaceae count, yeasts and moulds count

### INTRODUCTION

Every parent heed about their health and feeding. The best source of babies feeding is their mother's milk (breast feeding). But in few cases the mother naturally fails to fulfill the breast feeding requirement of baby due to disease factor or hormonal imbalance. But in some cases the mother itself is not interested in breast feeding, reasons manifested, the working females can not provide proper feeding to their baby. In rare cases modern world females do not feed their babies just to maintain their apparently beauty. So they follow the infant formula (available in market) suggested by their nutritionist or doctor. No doubt, the Infant milk powders are generally considered as product of good microbiological quality with no risk of spoilage, but several factors may contribute to change its physical and chemical properties which reduce shelf-life and thus its commercial value (Cousins *et al.*, 1987). Although the micro-organisms in infant milk cannot grow due to its low moisture content and do not play any direct role in their spoilage. But their occurrence in infant milk powder is of great significance and serves as an index of hygienic standards maintained during production,

processing and handling. The infant milk provides a highly nutritious substrate that can support the wide variety of bacteria as well as yeast and molds for their growth and reproduction (Phillips and Griffiths, 1990). The contamination role of bacteria during the preparation of infant milk powder has been well documented. The thermophilic can have significant economic consequences when they exceed specification limits and may result in down grading of the products (Ronimus *et al.*, 2005). Because these have ability to produce extremely heat resistant spores, and thus are significant source of pre- and post pasteurization (White *et al.*, 1993). Since, no work has been reported on any aspects of Infant milk powders in the province of Sindh. Thus, present study has been designed for evaluating microbiological quality of milk powders.

### MATERIALS AND METHODS

**Collection of infant powder samples:** A total of sixty samples of infant milk powders i.e 20 form each category A (1-6), B (7-12) and C for (7-18 months) babies were purchased from Hyderabad and brought to

the Laboratory of Dairy Microbiology, Department of Dairy Technology, Faculty of Animal Husbandry and Veterinary Sciences, Sindh Agriculture University Tandojam, for microbial examination.

**Preparation of test samples:** Milk powder (10 g) was diluted in warm (45°C) sterile diluents peptone water solution (90 ml) to make primary dilution ( $10^{-1}$ ). Then a series up to  $10^{-5}$  dilution was prepared by transferring primary dilution (1 ml) into test tube containing sterile diluents (9 ml) to obtain  $10^{-2}$  dilution and repeating the operations with sterile diluents (9 ml) using the  $10^{-2}$  and further dilutions to obtain  $10^{-3}$ ,  $10^{-4}$  and/or  $10^{-5}$ .

**Enumeration of total viable count (Colony count technique at 30°C):** Total viable counts were enumerated according to the method of International Dairy Federation (IDF, 1991). Pre prepared test sample (1 ml) of  $10^{-3}$ ,  $10^{-4}$  and/or  $10^{-5}$  dilutions (section-3.9.1) was transferred into sterile petri dishes in duplicate through sterile graduate pipette and/or dispensing pipette (1000  $\mu$ l) with sterile plastic tips and warm (45 $\pm$ 1°C) sterile plate count agar medium (15 ml) was mixed with inoculum. The mixture was allowed to solidify and incubated (30°C) for 72 $\pm$ 2 h. Parallel to that, control plates were also prepared using similar medium (15 ml) to check the sterility. The dishes containing more than 30 and/or fewer than 300 colonies were selected and counted using colony counter. The result was calculated using following formula.

**Enumeration of Enterobacteriaceae counts (Colony count technique at 37°C):** Enterobacteriaceae counts were enumerated according to the method of British Standard Institute (BSI, 1993). Pre prepared test sample (1ml) of  $10^{-1}$ ,  $10^{-2}$  and/or  $10^{-3}$  dilution (section 3.9.1) was transferred into sterile petri dishes through dispensing pipette (1000  $\mu$ l) with sterile plastic tips and warm (45 $\pm$ 1°C) sterile violet red bile agar medium (15 ml) was mixed with inoculum. The mixture was allowed to solidify and incubated (37°C) for 24 $\pm$ 2 h. Parallel to that control plates were also prepared using similar medium (15 ml) to check its sterility. The dishes containing more than 10 and/or fewer than 200 colonies were selected and counted using colony counter. The result was calculated using formula as mentioned in section 3.10.

**Enumeration of yeasts and moulds counts (Colony count technique at 25°C):** Yeasts and moulds count were enumerated according to the method of IDF (1990). Pre prepared test sample (1 ml) of  $10^{-1}$ ,  $10^{-2}$  and/or  $10^{-3}$  dilution (section-3.9.1) was transferred into sterile petri dishes through dispensing pipette (1000  $\mu$ l) with sterile plastic tips and warm (45 $\pm$ 1°C) sterile potato dextrose agar medium (15 ml) was mixed with inoculum. The mixture was allowed to solidify and incubated (25°C) for

5 days. Parallel to that control plates were also prepared using medium (15 ml) to check the sterility. The dishes containing more than 10 and/or fewer than 150 colonies were selected and counted using colony counter.

**Enumeration of thermoduric and thermophilic spore counts (Colony count technique at 55°C):** Thermoduric and thermophilic count was enumerated according to the method of Marshall (1993). Milk powder (10 g) was reconstituted in peptone water diluents (90 ml) and heated (80°C or 100°C) for 10 or 30 min to eliminate the vegetative cells. Heat treated sample (1 ml) of  $10^{-1}$ ,  $10^{-2}$  and/or  $10^{-3}$  dilution was transferred into petri dishes (in duplicate) through sterile pipette automatic pipette (1000  $\mu$ l) and warm (45 $\pm$ 1°C) sterile nutrient or milk starch agar medium (15 ml) was mixed with inoculum. The mixture was allowed to solidify and incubated (55°C) for 48 h. Parallel to that control plates were also prepared using medium (15 ml) to check the sterility. The dishes containing more than 10 and/or fewer than 200 colonies were selected and counted using colony counter.

**Total viable count:** Total viable count of Group A (1-6 months), Group B (7-12 months) and Group C (13-18 months) was evaluated and the results are presented in Fig. 1. No wide variation was observed in TV counts in all types of infant powders were examined in the present study. The concentration of TV count in Group A, ranged between  $2.3 \times 10^3$  to  $4.8 \times 10^3$  cfu/g and averaged  $3.9 \times 10^3 \pm 3.0 \times 10^2$  cfu/g. While in case of Group B, the TV counts were observed in between  $1.5 \times 10^3$  to  $5.3 \times 10^3$  cfu/g with mean value of  $3.4 \times 10^3 \pm 5.0 \times 10^2$  cfu/g. Where ever, TV count in Group C, varied between  $1.2 \times 10^3$  to  $6.5 \times 10^3$  cfu/g and averaged  $3.3 \times 10^3 \pm 6.1 \times 10^2$  cfu/g.

Moreover, the results of statistical analysis showed no significant difference ( $p > 0.05$ ), in TV counts in Group A, Group B and Group C. The concentration of TV counts were lower in Group A (12.18 folds), Group B (14.70 folds) and in Group C (15.15 folds) compared to Indian Standards Institute, ISI and Pakistan Standard Institution i.e  $\leq 5.0 \times 10^4$  cfu/g.

**Thermoduric count:** Thermoduric count of (Group A), (Group B) and (Group C) was evaluated and the results are presented in Fig. 2. A wide variation was not observed in TD counts in all types of infant formula powders examined in the present study. The concentration of TD count in Group A ranged between  $10 \times 0$  to  $2 \times 10$  cfu/g and averaged  $< 10$  and  $> 5 \pm 2.3 \times 10^1$  cfu/g. While in case of Group B, the TD counts were observed in between  $10 \times 0$  to  $1.1 \times 10$  cfu/g with mean value of  $< 10$  and  $> 5 \pm 1.4 \times 10^1$  cfu/g. Where ever, TD count in Group C varied between  $1 \times 0$  to  $1.2 \times 10$  cfu/g and averaged  $< 10$  and  $> 5 \pm 1.5 \times 10^1$  cfu/g.

Table 1: Total Viable Counts (cfu /g) in different infant formula samples compared to ISI/PSI standards.

Sample	Total Viable Count (TVC) cfu/g	
	Observed (a)	Deviation in folds from ISI standard (b) = (x) ÷ (a)
Group A	3900	-12.8
Group B	3400	-14.70
Group C	3300	-15.15

a = Observed Values

x = (Standard Value of PSI/ISI =  $\leq 50000$  cfu/g)

ISI = Indian Standards Institution

Table 2: Thermoduric Counts (cfu /g) in different infant formula samples compared to ISI standards.

Sample	Thermoduric Count (TDC, cfu/g)	
	Observed (a)	Deviation in folds from ISI standard (b) = (x) ÷ (a)
Group A	$\leq 10$	-10
Group B	$\leq 10$	-10
Group C	$\leq 10$	-10

a = Observed Values

x = (Standard Value of ISI (1993) =  $\leq 1.0 \times 10^2$  cfu/g)

ISI = Indian Standards Institution

Table 3: Thermophilic Counts (cfu /g) in different infant formula samples compared to ISI standards.

Sample	Thermophilic Spore Count (TPSC cfu/g)	
	Observed (a)	Deviation in folds from ISI standard (b) ÷ (a)
Group A	14	-7.14
Group B	15	-6.6
Group C	16	-6.25

a = Observed Values

x = (Standard Value of ISI (1993) =  $\leq 1.0 \times 10^2$  cfu/g)

ISI = Indian Standards Institution

Furthermore, Analysis of Variance (ANOVA) showed significant difference ( $p < 0.05$ ), in TD counts in Group A, Group B and Group C. It was further observed that TD count of Group A, Group B and Group C was no significantly different ( $p > 0.05$ ) in TD counts observed among all groups. The concentration of TD counts was lower in Group A (12.5 folds), Group B (16.0 folds) and in Group C (14.2 folds) compared to that of Indian Standards Institute (ISI, 1975) i.e  $\leq 1.0 \times 10$  cfu/g (Table 2).

**Thermophilic spore count:** Group A, Group B and Group C were evaluated for thermophilic spore count and the results are presented in Fig. 3. No variation was observed in TPS counts in all groups of infant formula milk powders examined in the present study. The concentration of TPS count in Group A ranged between  $<5$  to  $2.7 \times 10$  cfu/g and averaged  $1.4 \times 10 \pm 2.4 \times 10$  cfu/g. While in case of Group B, the TPS counts were observed in between  $<5$  to  $2.3 \times 10$  and averaged  $1.5 \times 10^2 \pm 2.2 \times 10^1$  cfu/g. TPS count in FCMP varied between  $<10$  to  $2.3 \times 10$  cfu/g and averaged  $1.6 \times 10 \pm 2 \times 10$  cfu/g.

Furthermore, the results of statistical analysis (AOV) showed non significant difference ( $p > 0.05$ ), in TPS

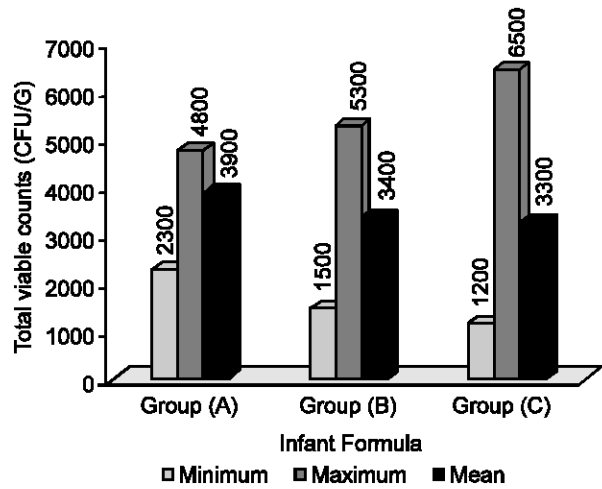


Fig. 1: Graph shows minimum, maximum and mean values of total viable counts (cfu/g) in infant formula

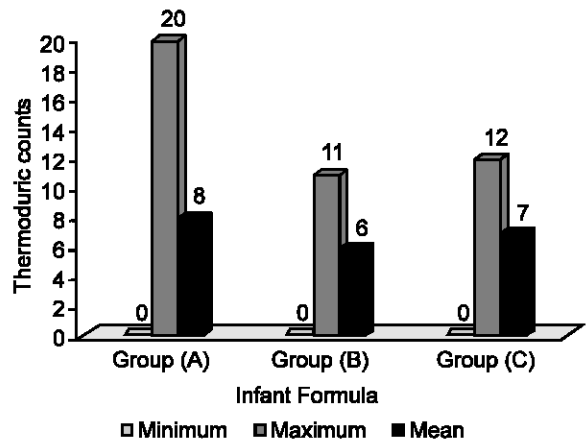


Fig. 2: Graph shows minimum, maximum and mean values of Thermoduric counts (cfu/g) in infant formula

counts in Group A, Group B and Group C. The concentration of TPS counts was lower in Group A (7.14 folds), Group B (6.6 folds) and in Group C (6.25 folds) compared to that of Indian Standards Institute (ISI, 1975) i.e  $\leq 1.0 \times 10^2$  cfu/g.

**Enterobacteriaceae count:** All the three groups were evaluated for Enterobacteriaceae count and the results are depicted in Fig. 4. TPS counts did not vary greatly in all types of infant formula of milk powders examined in the present study. The concentration of Enterobacteriaceae count in Group A ranged between  $10 \times 0$  to  $<10$  and  $>5$  cfu/g and averaged  $<5 \pm 1.0 \times 10$  cfu/g. While in case of Group B the Enterobacteriaceae counts were observed in between  $10 \times 0$  to  $<10$  and averaged  $<5 \pm 1.0 \times 10$  cfu/g. whereas in Group C, varied between  $<5$  to  $<10$  cfu/g and averaged  $<5 \pm 1.0 \times 10$  cfu/g.

Table 4: Enterobacteraceae Counts (cfu /g) in different infant formula samples compared to ISI standards.

Sample	Enterobacteraceae Count (EbC cfu/g)	
	Observed (a)	Deviation in folds from ISI standard (b) ÷ (a)
Group A	≤5	-20
Group B	≤5	-20
Group C	≤5	-20

a = Observed Values

x = (Standard Value of ISI (1993) =  $\leq 1.0 \times 10^2$  cfu/g)

ISI = Indian Standards Institution

Table 5: Yeast and Mold Counts (cfu /g) in different infant formula samples compared to ISI standards.

Sample	Yeasts and Moulds Count (YMC cfu/g)	
	Observed (a)	Deviation in folds from ISI standard (b) = (x) ÷ (a)
Group A	≤5	-20
Group B	≤5	-20
Group C	≤5	-20

a = Observed Values

x = (Standard Value of ISI (1993) =  $\leq 1.0 \times 10^2$  cfu/g)

ISI = Indian Standards Institution

Moreover, the results of statistical analysis (AOV) showed non significant difference ( $p > 0.05$ ), in Enterobacteriaceae counts in Group A, Group B and Group C. The concentration of Enterobacteriaceae counts was higher in Group A (33 folds), Group B (33 folds) and in Group C (25 folds) compared to that of Indian Standards Institute (ISI, 1993) i.e.  $\leq 1.0 \times 10^2$  cfu/g.

**Yeasts and moulds count:** Yeasts and moulds count of Group A, Group B and Group C was examined and the results are shown in Fig. 5. It was observed that Yeast and Moulds counts in all types of infant milk powders did not show variation. However, the concentration of yeasts and moulds count in Group A ranged between  $10 \times 0$  to  $< 10$  cfu/g and averaged  $< 5$  cfu/g. While in case of the Group B moulds counts were observed in between  $10 \times 0$  to  $< 10$  and averaged  $< 5 \pm 1.0$  cfu/g, whereas in Group C, varied between  $< 5$  to  $< 10$  cfu/g and averaged  $< 5 \pm 1.0$  cfu/g.

Statistical analysis (AOV) revealed non significant difference ( $p > 0.05$ ) in yeasts and moulds counts in Group A, Group B and Group C. The concentration of Yeast and Moulds counts were lower in Group A (33 folds), Group B (33 folds) and in Group C (25 folds) compared to that of Indian Standards Institute (ISI, 1975) i.e.  $\leq 1.0 \times 10^2$  cfu/g.

**RESULTS AND DISCUSSION**

Present study has been conducted to assess the general hygienic quality of Infant milk powders and the extent of microbes has been observed. Although the microorganisms in infant formula milk owing to their low moisture content can not grow and thus do not play any direct role in their spoilage, their occurrence in these

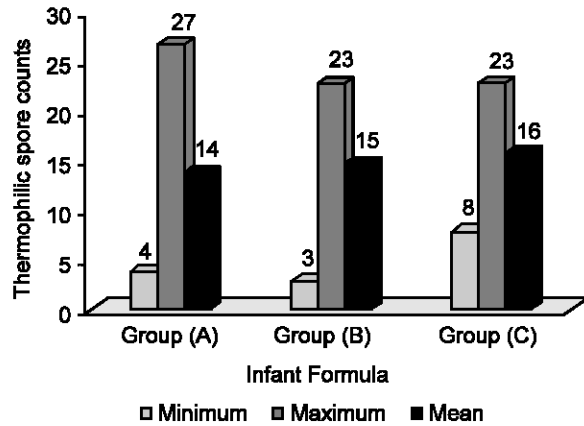


Fig. 3: Graph shows minimum, maximum and mean values of Thermophilic spore counts (cfu/g) in infant formula

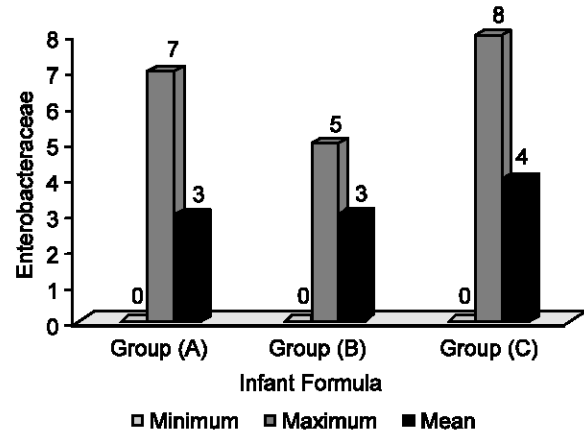


Fig. 4: Graph shows minimum, maximum and mean values of Enterobacteraceae counts (cfu/g) in infant formula

products is of great significance they serve as an index of hygienic standards maintained during Production, Processing and handling (Yadav *et al.*, 1993). In the present study the total viable count of Group A, ( $3.9 \times 10^3 \pm 3.0 \times 10^2$  cfu/g) was not significantly ( $p > 0.05$ ) higher than Group B, ( $3.4 \times 10^3 \pm 5.0 \times 10^2$  cfu/g) and Group C ( $3.3 \times 10^2 \pm 6.1 \times 10^2$  cfu/g).

It is of interest to point out that total viable count of all three groups ( $3.9 \times 10^3 \pm 3.0 \times 10^2$  cfu/g), ( $3.4 \times 10^3 \pm 5.0 \times 10^2$  cfu/g), ( $3.3 \times 10^2 \pm 6.1 \times 10^2$  cfu/g) observed in the infant formula is lower than reported by Rueckert *et al.* (2005); Khaskheli (1998); ISI (1975) and PSI (2007) i.e.  $5.6 \times 10^4 \pm 4.3 \times 10^3$ ,  $8.7 \times 10^3$ ,  $1.0 \times 10^4$  and  $5.0 \times 10^4$  respectively. The vegetative cells normally killed at  $80^\circ\text{C}$  for 10 min. But the present study manifested their presence in infant powders. Because, their ability to attach with stainless steel and folded surface (Flint *et al.*, 2006). Once they attach to the surface, vegetative cells

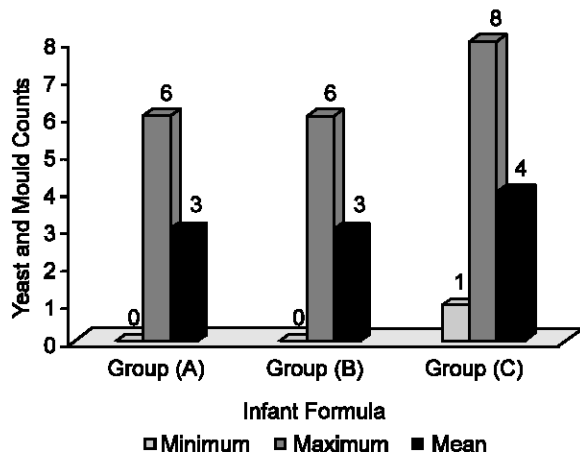


Fig. 5: Graph shows minimum, maximum and mean values of Yeasts and Molds counts (cfu/g) in infant formula

grow with spores by forming biofilms. This biofilms is not completely removable by CIP system but can be decreased; the remaining contaminants present on folded stainless steel transfer into final product, i.e powders milk (Parker *et al.*, 2001).

Thermophilic count of Group A, ( $<10$  and  $>5\pm 2.3 \times 10^1$  cfu/g) was not significantly ( $p > 0.05$ ) higher than Group B ( $<10$  and  $>5\pm 1.4 \times 10^1$  cfu/g) and C ( $<10$  and  $>5\pm 1.5 \times 10^1$  cfu/g). Moreover, the mean ( $3.7 \times 10^1$ ) of TD counts in the present study is lower than reported values ( $1.8 \times 10^1$  cfu/g) of Infant milk powders. ( $3.8 \times 10^1$  cfu/g), Ronimus *et al.* (2005). The reason of thermophilic growth is processing, if they are present in raw milk their growth accelerate at the time of pasteurization, because temperature of pasteurization is favorable for the growth of thermophilic bacteria (Murphy *et al.*, 1999). The transit time between the silo milk and spray drier is typically 20-30 min there is obviously bacterial growth is clearly associated with processing and bio transfer to the end product (Flint *et al.*, 2006; Wirtanen *et al.*, 1996 and Stadhouders *et al.*, 1982).

The thermophilic spore of Group A ( $1.4 \times 10^2 \pm 2.4 \times 10^1$  cfu/g) was lower than Group B ( $1.5 \times 10^2 \pm 2.2 \times 10^1$  cfu/g) Group C ( $1.6 \times 10^2 \pm 2 \times 10^1$  cfu/g) However, the averages obtained in present study is lower than the mean value reported by Rueckert *et al.* (2005) i.e ( $3.2 \times 10^4 \pm 3.4 \times 10^3$  cfu/g) and ( $2.4 \times 10^4 \pm 5.1 \times 10^3$  cfu/g). If the spores are present in raw milk that rapidly grow, when they obtain favorable temperature during milk processing (pasteurization), the other evidence provided i.e foulant, it is a major source of thermophilic contamination in a full scale milk powder plant (Scott *et al.*, 2007).

Enterobacteriaceae count of Group A ( $<5 \pm 1.0 \times 10^0$  cfu/g) was not significantly ( $p > 0.05$ ) higher than Group B ( $<5 \pm 1.0 \times 10^0$  cfu/g) and Group C ( $<5 \pm 1.0 \times 10^0$  cfu/g). The mean values of all three groups A, B and C obtained in present

study is lower than reported by Taha *et al.* (1972) i.e  $13 \times 10^6$ . It is the general concept that enterobacteriaceae are not present in Infant formula proved by various researches. The Infant formula powder is packed hygienically in large sterilized containers and bags. However, transportation some damaged containers and bags have been observed, probably they can contaminate the milk powders.

The yeasts and moulds count of Group A ( $<5$  cfu/g), Group B ( $<5 \pm 1.0$  cfu/g) and Group C ( $<5 \pm 1.0$  cfu/g) were non significant different. However, the mean value ( $3.6 \times 10^2 \pm 3.8 \times 10^1$ ) of yeasts and moulds in the present study is lower than the results presented by Rossi *et al.* (1974) i.e  $>100000/100$  g in powder milk and reported by Ceittao *et al.* (1973) i.e  $<1000/g$  in milk powder. Presence of yeasts and moulds in milk or milk products, molds may create hazard to one's health, produce an allergen and an irritant to human health (Parihar and Parihar, 2008).

## REFERENCES

- BSI, 1993. Microbiological examination of food and animal feeding stuffs. Enumeration of Enterobacteriaceae, In: British Standards Institution London (U.K). BS, 5763.
- Ceittao., M.F, F. De, I. Dalzani and H. Mazzoni, 1973. Cited Dairy Science. Abstract, 39, 6388.
- Cousins, C.M., A.J. Bramley and R.K. Robinson, 1987. Microbiologia de la leche cruda. microbiologia lactologica. V.1. Acribia, Zaragoza, pp: 109-150.
- Flint, S., J-L. Drocourt, K. Walker, B. Stevenson M. Dwyer, I. Clarke and D. McGill, 2006. A rapid, two-hour method for the enumeration of total viable bacteria in samples from commercial milk powder and whey protein concentrate powder manufacturing plants. Int. Dairy J., 16: 379-384.
- IDF, 1991. Enumeration of microorganism in milk and milk products. Colony counts at 30°C. In: International Dairy Federation, Brussels (Belgium).
- IDF, 1990. Enumeration of yeast and molds in milk and milk products. Yeast and molds at 25°C. In: International Dairy Federation, Brussels (Belgium).
- ISI, 1975. Milk Powder. IS: 1165. Quated by Yadave *et al.* (1993). A Comprehensive Dairy Microbiology. B.V. Gupta, metropolitan book Co Pvt. Ltd., pp: 685-699.
- ISI, 1993. A Comprehensive Dairy Microbiology. Milk Powder. IS: 1165. Quated by Yadave *et al.* (1993) B.V. Gupta, metropolitan book Co Pvt. Ltd., pp: 685-699.
- Khaskheli, M., 1998. Some aspects of the production and quality improvements of fermented milk/cereal mixture (Kishk). Ph.D thesis, University of Glasgow, UK.
- Marshall, R.T., 1993. Tests for groups of microorganisms of dairy products. In: Standard methods for the examination of dairy products, American Public Health Association, Washington, USA., pp: 271-286.

- Murphy, P.M., D. Lynch and P.M. Kelly, 1999. Growth of thermophilic spore forming bacilli in milk during the manufacture of low heat powders. *Int. J. Dairy Technol.*, 52: 45-50.
- Parker, E.L., R.S. Ronimus, N. Turner, S. Poudel, A. Rueckert and H.W. Morgan, 2001. A RAPD-based comparison of thermophilic bacilli from milk powders. *Int. J. Food Microbiol.*, 85: 45-61.
- Parihar, P. and L. Parihar, 2008. Dairy microbiology Chapter, 3. *Microbes*. Agrobios India, pp: 46-50.
- Phillips, J.D. and M.W. Griffiths, 1990. Pasturized dairy products: constraints imposed by environmental contamination. In: *contamination from environmental source*. Wiley, USA, pp: 387-456.
- PSI, 2007. Specification for milk powder (1st revision) PS: 363. Pakistan Standards Institution, Karachi, Pakistan.
- Ronimus, R.S., A. Rueckert and H.W. Morgan, 2005. Survival of thermophilic spore-forming bacteria in a 90+ year old milk powder from Ernest Shackleton's Cape Royds Hut in Antarctica. Thermophile Research Unit., Department of Biological Sciences, University of Waikato, Private Bag 3105, Hamilton New Zealand.
- Rueckert, A., R.S. Ronimus and H.W. Morgan, 2005. Rapid differentiation and enumeration of the total, viable vegetative cell and spore content of thermophilic bacilli in milk powders with reference to *Anoxybacillus flavithermus*. *J. Applied Microbiol.*, 99: 1246-1255.
- Stadhouders, J., G. Hup and F. Hassing, 1982. The conceptions index and indicator organisms discussed on the basis of the bacteriology of spray-dried milk powder. *Netherlands Milk and Dairy J.*, 1: 231-260.
- Scott, S.A., J.D. Brooks, J. Rakonjac, K.M.R. Walker and S.H. Flint, 2007. The formation of thermophilic spores during the manufacture of whole milk powder. *Int. J. Dairy Technol.*, 60: 109-117.
- Rueckert, A., R.S. Ronimus and H.W. Morgan, 2005. Rapid differentiation and enumeration of the total, viable vegetative cell and spore content of thermophilic bacilli in milk powders with reference to *Anoxybacillus flavithermus*. *J. Applied Microbiol.*, 99: 1246-1255.
- Rossi, C., S. Androetto and C. Pellagrino, 1974. Symp. Intern. Sulla. *Mirobiol. Dairy Science*. Abstract, 37: 6377.
- Taha, S., M. Naguib and A. Ghani, 1972. Dried milk microbiology. *J. Milk. Food. Technol.*, 36: 559.
- Wirtanen, G., U. Husmark and T. Mattilasandholm, 1996. Microbial evaluation of the biotransfer potential from surfaces with *Bacillus* biofilms after rinsing and cleaning procedures in closed foodprocessing systems. *J. Food Prot.*, 59: 727-733.
- White, D., R.J. Sharp and F.G. Priest, 1993. A polyphasic taxonomic study of thermophilic bacilli from a wide geographical area. *Antonie van Leeuwenhoek*, 64: 357-386.
- Yadav, J.S., S. Grover and V.K. Batish, 1993. Microbiology of dried milks. *A Comprehensive Dairy Microbiology*. B.V. Gupta, metropolitan book Co Pvt. Ltd. pp: 315-349.