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## ***In silico Structural Analysis of 16S rDNA Sequences of Bacteria Isolated from Keratitis Patients***

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### **ABSTRACT**

Keratitis is a condition in which the cornea of the eye becomes inflamed. It is caused by bacteria, viruses, fungi and parasites. The diagnosis of the causative organism of keratitis remains a problem due to lack of advanced techniques. In this study PCR and sequence analysis of 16S rDNA was used to ascertain the bacteria isolated in eye swabs of keratitis patients. The bacterial genes were sequenced and deposited in the GenBank (NCBI) with the accession numbers JN378393, JN378392, HM204502, JQ039348, JN652127, JN652129, JQ039350, JN652128, JQ039349 and HQ404365 for *Streptococcus viridan*, *Staphylococcus aureus*, *Micrococcus* sp., *Moraxella* sp., *Citrobacter koseri*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, *Klebsiella* sp., *Propionibacterium* sp. and *Staphylococcus epidermidis*, respectively. Sequences were analyzed and aligned by ClustalX and a phylogenetic relationship was studied. The secondary structure of the 16S DNA was developed using GeneBee which produces secondary structures prediction through the minimization of the potential energy of the system. These results may be useful in characterizing the micro evolutionary mechanisms of the species for researchers.

**Key words:** 16S rDNA, keratitis, GeneBee, phylogenetic analysis

### **INTRODUCTION**

Microbial keratitis is a leading cause of visual loss in developing countries. It is normally predisposed by ocular trauma and/or contact lens wear. Corneal scarring listed second only to cataracts as an important cause of blindness and visual impairment in many developing countries in Asia, Africa and the Middle East (Whitcher and Srinivasan, 1997). Bacterial keratitis is the most common form of suppurative corneal ulceration. A number of organisms are able to cause infection (Dunlop *et al.*, 1994; Upadhyay *et al.*, 1991; Wilhelmus, 1996) and analyses of them are essential to understand how they set up the infection.

Approximately 65-90% of all corneal infections is bacterial keratitis (Asbell and Stenson, 1982; McDonnell and Green, 1990; Jones, 1981; Laibson, 1972; Liesegang and Forster, 1980). Although, these bacteria may vary in incidence according to geographical locale (Baum, 1979) the most common organisms include *Staphylococcus aureus*, *Streptococcus epidermidis*, *Streptococcus pneumoniae*, *Pseudomonas aeruginosa* and other Gram negative bacilli (Morlet and Daniell, 2003; Jones, 1979; Baum, 1978). Of these, *S. aureus* is the predominant pathogen isolated from the majority of cases of keratitis, (Morlet and Daniell, 2003; Baum, 1978) but *P. aeruginosa*, a potentially devastating ocular pathogen, is the most common cause of hypopyon corneal ulcers, (Hyndiuk, 1981) ulcerative keratitis associated with contact lens wear and severe necrotic corneal ulceration (Laibson, 1972).

In this study, 16S rDNA sequences of bacteria isolated from infectious keratitis in eye. Clinic of the General Hospital of Tiruchirappalli Tamil Nadu were sequenced. A comparative phylogenetic analysis of the 16S rDNA sequences were performed by using ClustalX and the phylogenetic tree was constructed using NJ method. The secondary structure of the 16S rDNA was constructed and minimal free energy was calculated by Genebee tool. The ribosomal RNA mainly 16S r DNA has proven to be a stable and specific molecular marker of the identification of bacteria. The 16S rDNA is a common target for the taxonomical purpose, mainly due to the mosaic composition of phylogenetically conserved and variable regions.

## MATERIALS AND METHODS

**Sample collection:** In this study, bacteria were isolated from keratitis patients in Eye Clinic of the General Hospital of Tiruchirappalli Tamil Nadu during 2010-2012 All patients were examined under a slit-lamp biomicroscope by an ophthalmologist. Corneal scrapings were collected after instillation of 4% lignocaine without preservative under aseptic conditions from each ulcer by an ophthalmologist using a sterile Bard Parker blade (No. 15). Scrapings were performed under magnification of slit-lamp or operating microscope. Leading edge and base of each ulcer were scraped initially and the material obtained were directly inoculated onto the surface of solid media such as sheep blood agar, chocolate agar and Sabouraud Dextrose Agar (SDA) in a row of C-shaped streaks and also deep inoculation in the liquid media such as Brain Heart Infusion (BHI) broth without gentamicin sulphate and thioglycollate medium. Subsequent scrapings were spread onto labelled slides in a thin, even manner for 10% potassium hydroxide (KOH) wet mount and Gram staining.

**Culture:** All bacterial cultures were incubated aerobically. Cultures on blood agar and chocolate agar were evaluated at 24 hours and at 48 hours and then discarded if no growth was seen. All media's were incubated at 35°C ( $\pm 1$ ) except SDA which are incubated at 27°C ( $\pm 1$ ) in BOD incubator. Cultures inoculated in BHI broth were examined for turbidity in similar fashion which was subsequently subcultured and Gram stained for identification. However, liquid media were prone to contamination and were not used for interpretation in isolation. The criteria described by Bharathi *et al.* (2002) were used for determining culture positive samples.

The specific identification of bacterial pathogens was based on microscopic morphology, staining characteristics and biochemical properties using standard laboratory criteria.

**PCR and DNA sequencing:** Genomic DNA was isolated from sample and loaded on 1% agarose gel. PCR amplified was performed using 16s rDNA primers (PCR amplification gel Photo attached). The PCR product was gel eluted. The purified PCR product was sequenced using 2 primers. The conditions used for PCR amplification of DNA: 1, 16S Forward Primer 400 ng, 16S Reverse Primer 400 ng, dNTPs (2.5 mM each) 4, 10X Taq DNA Polymerase Assay Buffer 10, Taq DNA Polymerase Enzyme (3 U L<sup>-1</sup>) 1, water X total reaction volume: 100, the two universal primers and 1.5 mM MgCl<sub>2</sub>. Amplification was done by initial denaturation at 94°C for 5 min, followed by 35 cycles of denaturation at 94°C for 30 sec, annealing temperature of primers was 55°C for 30 sec and extension at 72°C for 2 min. Final extension was at 72°C for 15 min All PCR reagents were of Chromous Biotech, Ltd.

The PCR product were detected by using 1% agarose gel containing Ethidium bromide and the result recorded by UV transilluminator. Finally, the target PCR product was sliced and recovered. After purification, the PCR product was submitted for sequencing. Sequences have been deposited in GenBank.

**Sequence analysis:** Sequences were analyzed and aligned with other sequences deposited in Genbank by ClustalX (Thompson *et al.*, 1997). The secondary structure and minimal free energy were calculated by Genebee tool (Mathews *et al.*, 1999).

## RESULTS AND DISCUSSION

**Isolation and characterization bacterial of strains:** The bacterial colonies differ greatly in their morphologies and these differences can help us in identifying different species of bacteria. The culture containing *Streptococcus viridans*, *Staphylococcus aureus*, *Micrococcus* sp., *Moraxella* sp., *Citrobacter koseri*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, *Klebsiella* sp., *Propionibacterium* sp., *Staphylococcus epidermidis* bacteria were and isolated from patients and identified by using biochemical tests. It shows that it was cocci shaped and Gram positive and Gram negative bacteria and it has glucose, lactose, sucrose fermentations (acid and gas), catalase enzyme activity (degradation of hydrogen peroxide) and non-acidic in nature.

**Molecular characterization of bacteria:** Molecular characterization of the bacterial strains was carried out by sequencing their 16S rDNA after its amplification and sequencing. Most strains showed high sequence identities with the same species on GenBank and some strains showed high identities with other species. This analysis made the identification of reference strains more reliable. The bacterial gene was sequenced and it deposited in the GenBank (NCBI) with the accession numbers JN378393, JN378392, HM204502, JQ039348, JN652127, JN652129, JQ039350, JN652128, JQ039349, HQ404365 for *Streptococcus viridan*, *Staphylococcus aureus*, *Micrococcus* sp., *Moraxella* sp., *Citrobacter koseri*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, *Klebsiella* sp., *Propionibacterium* sp., *Staphylococcus epidermidis*, respectively are shown in Table 1.

**Sequence analyses and phylogenetic tree constructing:** The 16S rRNA gene sequences of the bacterial strains studied by us were subject to multiple sequence alignment using ClustalX and the phylogenetic tree was drawn using Neighbour Joining plot. Multiple sequence alignment (Fig. 1) confirmed sequence conservation of the highly conserved regions of 16S rDNA of the causative bacteria of keratitis. The phylogenetic analysis of these genes from the phylogenetic tree (Fig. 2) shows that these genes are closely related.

Table 1: Bacteria isolated from keratitis patients

Microorganism	Accession No.
<b>Gram-negative bacteria</b>	
<i>Pseudomonas aeruginosa</i>	JQ039350
<i>Acinetobacter baumanii</i>	JN652129
<i>Klebsiella</i> sp.	JN652128
<i>Propionibacterium</i> sp.	JQ039349
<i>Citrobacter koseri</i>	JN652127
<b>Gram-positive bacteria</b>	
<i>Staphylococcus aureus</i>	JN378392
<i>Staphylococcus epidermidis</i>	HQ404365
<i>Micrococcus</i> sp.	HM204502
<i>Streptococcus viridians</i>	JN378393
<i>Moraxella</i> sp.	JQ039348

JN652127	-----CCTGGCTCAGATTGAGGTGTGGCGGCAGGCCTAACACATGCAAGTCGAACG 51
JN652128	-----
JN378392	-----
HQ404365	-----AGGATGAACACTGGCGCGTGCCTAACATGCAGGTCGAGCG 43
JQ039348	-----CTGGCGGCAGGCTAACACATGCAAGTCGAACG 33
JN652129	-----
JQ039350	-----
HM204502	-----A 1
JQ039349	-----CGGCGTGCTTAACACATGCAAGTCGAACG 29
JN378393	GAGTTTGATCCTGGCTCAGGACGAACGCTGGCGCGTGCCTAACATGCAAGTAGAACG 60
JN652127	GTAACAG---GAAGCAGCTTGCTGCTTGCT---GACGAGTGGCGGACGGGTGAGTAATG 105
JN652128	-----
JN378392	-ACAG---ACGAGAAGCTTGCTCTG---ATGTTANCGCGGACCGGTGAGTAACA 51
HQ404365	AACAG---ACGAGGAGCTTGCTCTG---ACGTTAGCGCGGACGGGTGAGTAACA 95
JQ039348	ATGAA----CTCTAGCCTGCT---AGACG---GATTAGTAGCGAACGGGTGAGTAATG 81
JN652129	-----CGAAAGGG----- 8
JQ039350	-----
HM204502	GAGAA----GCCAGCTTGCTGCGGGTG---GATTAGTGGCGAACGGGTGAGTAACA 51
JQ039349	GAAAG----GCCCTGTTTGTGGGTG---CTCGAGTGGCGAGCGGGTGAGTGACA 79
JN378393	CACAGTTTACCGTAGCTTGCTACACCATAGACTGTGAGTTGCGAACGGGTGAGTAACG 120
JN652127	TCTGGG-AAACTGCCTGATGGAGGGGATAACTACTGGAAACGGTAGCTAACACGCATA 164
JN652128	-----
JN378392	CGTGGATAACCTACCTATAAGACTGGGATAACTTCGGGTAACCGGAGCTAACACGGATA 111
HQ404365	CGTGGATAACCTACCTATAAGACTGGGATAACTTCGGGAAACCGGAGCTAACACGGATA 155
JQ039348	C-TAGGAATCTGCCTATTAGTGGGGATAACGTAGGAAACTCACGCTAACACGCATA 140
JN652129	-----ATGCTAACACGCATA 24
JQ039350	-----
HM204502	CGTGAGTAACCTGCCCTAACCTGGGATAAGCCTGGAAACTGGGTCTAACACCGGATA 111
JQ039349	CGTGAGTAACCTGCCCTGACTTTGGGATAACTTCAGGAAACTGGCGCTAACACCGGATA 139
JN378393	CGTAGGTAACCTGCCTGGTAGCGGGGGATAACTATTGGAAACGATAGCTAACACGCATA 180
JN652127	ACGTCGCAAGACCAAAGAGGGGGACCTTCGGGCCTCTT-----GCCATCAGATGTGC 216
JN652128	-----
JN378392	ATATTTGAACCGCATGGTCAAAAGTGAAAGACGGTCTTGCTGTCACTTAGATGGAT 171
HQ404365	ATATATTGAACCGCATGGTCAATAGTGAAAGACGGTTTGCTGTCACTTAGATGGAT 215
JQ039348	CGTCTTACGAGAGAAA---GGGGGCCT-TTAGCTCT-----CGCTAACAGATGAGC 187
JN652129	CGCCTACGGAGAAAGCAGGGATCT-CCAAACCTG-----CGCTAACAGATGAGC 76
JQ039350	-----
HM204502	GGAGCGTCCACCGCATGGTGGGTGTTGAAAGATTAT-----CGGTTTTGGATGGAC 164
JQ039349	GGAGCTTCTGCTGCATGGTGGGGTTGGAAAGCTTCGG-----CGGTTGGGGATGGAC 192
JN378393	ATATTAATTATTGCTGATAATTAATTGAAAGGTGCAATTGCA-CCACTACCAGATGGAC 239
JN652127	CCAGATGGGATTAGCTNGTGGTGGGTAACGACTCACCAAGGGACGATCCCTAGCTGG 276
JN652128	-----
JN378392	CCCGCGCTGCATTACCTAGTTGGTAAGGTAACGGCTTACCAAGGCAACGATGCTAGCCGA 231
HQ404365	CCCGCGCGCATTAGCTAGTCGGTAGGGTAACGGCTTACCAAGGCAACGATGCGTAGCCGA 275
JQ039348	CTAAGTCGGATTAGCTAGTTGGTGGGTAAGGCTAACAGGCGATCTGTAGCTAG 247

Fig. 1: Continue

JN652129	CTAAGTCGGATTAGTTAGTTGGTGGGGTAAAGGCCTACCAAGGCGACAATCTGTAGCGGA	136
JQ039350	-----	
HM204502	TCGCGGCCTATCAGCTTGTGGTGAGGTAATGGCTCACCAAGGCGACGACGGGTAGCCGG	224
JQ039349	TCGAGCTTATCAGCTTGTGGTGAGGCTTACCGAGGCTTGACGGGTAGCCGG	252
JN378393	CTGCGTTGACTAGTAGTAGGTGAGGTAACGGCTACCTAGGCGACGATACATAGCCGA	299
JN652127	TCTGAGAGGGATGACCAGCCACACTGGAAC TGAGACACGGTCCAGACTCCTACGGGAGGCA	336
JN652128	-----	
JN378392	CATGAGAGGGTGATCGGCCACACTGGAAC TGAGACACGGTOCAGACTCCTACGGGAGGCT	291
HQ404365	CCTGAGAGGGTGATCGGCCACACTGGAAC TGAGACACGGTCCAGACTCCTACGGGAGGCA	335
JQ039348	TCTGAGAGGGATGATCAGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCA	307
JN652129	TCTGAGAGGGATGATCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCA	196
JQ039350	-----	
HM204502	CCTGAGAGGGTGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCA	284
JQ039349	CCTGAGAGGGTGACCGGCCACATTGGGACTGAGATA CGGCCAGACTCCTACGGGAGGCA	312
JN378393	CCTGAGAGGGTGATCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCA	359
JN652127	GCAGTGGGAATATTGCA-CAATGGCGCAAGCCTGATGCAGCCATGCCCGTGTATGAA	395
JN652128	-----	
JN378392	GCAGTAGGAAATCTTCTTCAATGGCGAAAGCCTGACGGAGCAACGCCCGTGTGAGTGT	351
HQ404365	GCAGTAGGAAATCTTCCG-CAATGGCGAAAGCCTGACGGAGCAACGCCCGTGTGAGTGT	394
JQ039348	GCAGTGGGAATATTGGA-CAATGGCGAAAGCCTGATTCAGCCATGCCCGTGTGTGAA	366
JN652129	GCAGTGGGAATATTGGA-CAATGGGGAGCCCTGATCCAGCCATGCCG-----A	247
JQ039350	-----	
HM204502	GCAGTGGGAATATTGCA-CAATGGCGCAGGCCTGATGCAGCGACGCCCGTGTGAGGGAT	343
JQ039349	GCAGTGGGAATATTGCA-CAATGGCGGAAGCCTGATGCAGCAACGCCCGTGCGGGAT	371
JN378393	GCAGTAGGAACTTCGG-CAATGGACGAAAGTCTGACCGAGCAACGCCCGTGTGAGTGA	418
JN652127	GAAGG-CCTTCGGTTGTAAA--GTACTTTCAGCGGGGAGGAAG--GTGTTGTGGTTAA	449
JN652128	-----	
JN378392	GAAGGTACTTCGGATCGTAAACTCCTGTTATTAGGGAAGAACATATGTGTAAGT-AAC	410
HQ404365	GAAGG-TCTTCGGATCGTAA- ACT-CTGTTATTAGGGAAGAACAAATGTGTAAGT-AAC	450
JQ039348	GAAGG-CCTTTGGTTGTAAA-GCACTTTAAGTAGGGAGGAAA--AGCTTGTGGTTAA	420
JN652129	GC GG-----	251
JQ039350	-----	
HM204502	GACGG-CCTTCGGTTGTAAA--CCTCTTCAGTAGGGAAGAAG--CGAAAGTG----	392
JQ039349	GACGG-CCTTCGGTTGTAAA--CCGCTTCCGCTGTGACGAAG--CGTGAGTG----	420
JN378393	GAAGG-TTTTCGGATCGTAAA-GCT-CTGTTGTAGAGAAGAACGGGTGTTAGAGTGGAA	475
JN652127	TAACCACAGCAATTGACGTACCCGCA-GAAGAACCGGCTAACTCCGTGCCAGCAGC	508
JN652128	-----	
JN378392	TGTGCACATCCTTGTACGGTACCTACNGCAGAAAGCCACGGCTAA-----	455
HQ404365	TATGCACGTCTG--ACGGTACCTAAT-CAGAAAGCTACGGCTAACTACGTGCCAGCAGC	507
JQ039348	TACCCACAAGCCTGACGTACCCACA-GAATAAGCACGGCTAGCTCTGTGCCAGCAGC	479
JN652129	-----	
JQ039350	-----	
HM204502	-----ACGGTACCTGCA-GAAGAACCGGCTAACTACGTGCCAGCAGC	436

Fig. 1: Continue

JQ039349	-----ACGATAATGGGT-AAAGAACGCCGGCTAAGTGCAGCAGC 464
JN378393	AGTTCACACTGTG-ACGGTATCTTAC-CAGAAAGGGACCGCTAACTAAGTGCCAGCAGC 532
JN652127	TGCGGTAATACGGAGGGTGCAAGCGTTAACCGAATTACTGGCGTAAAGCGCACGCAGG 568
JN652128	-----
JN378392	-----
HQ404365	CGCGGTAATACGTAGGTGGCAAGCGTTAACCGAATTATTGGCGTAAAGCGCGTAGG 567
JQ039348	CGCGGTAATACAGAGGGTGCAAGCGTTAACCGAATTACTGGCGTAAAGCGAGCGTAGG 539
JN652129	-----
JQ039350	-----
HM204502	CGCGGTAATACGTAGGGTGCAGCGTTAACCGAATTATTGGCGTAAAGAGCTCGTAGG 496
JQ039349	CGCGGTATACGTAGGGTGCAGCGTTAACCGAATTATTGGCGTAAAGGGCTCGTAGG 524
JN378393	CGCGGTAATACGTAGGTCCCAGCGTTAACCGAATTATTGGCGTAAA-CGAGCGCAGC 591
JN652127	CGGTCTGTCAAGTCAGATGTGAAATGCCCGGCTAACCTGGAACTGCATCTGATACTG 628
JN652128	-----
JN378392	-----
HQ404365	CGGTTTTTAAGTCTGATGTGAAAGCCCACGGCTAACCGTGGAGGGTATTGGAGACTG 627
JQ039348	TGGTCATTTAAGTCAGATGTGAAAGCCCCGGCTAACCTGGAACTGCATCTGATACTG 599
JN652129	-----
JQ039350	-----
HM204502	CGGTTGTCGCGTCTGCGTAAAGTCCGGGCTTAACCTCGGATCTGGTGGGTACGG 556
JQ039349	TGGTTGACCGCGTCGGAAGTGTAACTCTGGGCTAACCGTGGCTTCGATAACGG 584
JN378393	CGGTTAGATAAGTCTGAAGTAAAGGCTGTGGCTAACCATAGTACG-CTTGGAAACTG 650
JN652127	GCAGGCTTGAGTCTCGTAGAGGGGGTAGAATTCCAGGTAGCGGTAAATGCGTAGAG 688
JN652128	-----
JN378392	-----
HQ404365	GAAAACCTGAGTGCAGAACAGGAAAGTGAATTCCATGTAGCGGTAAATGCGCAGAG 687
JQ039348	GGTGAAGTGTAGGTGAGAGGGAAAGTAGAATTCCAGGTAGCGGTAAATGCGTAGAG 659
JN652129	-----
JQ039350	-----
HM204502	GCAGACTAGAGTGCAGTAGGGAGACTGAAATTCTGGTAGCGGTGAAATGCGCAGAT 616
JQ039349	GTTGACTCGAGGAAGGTAGGGAGAAATGAAATTCTGGTAGCGGTGAAATGCGCAGAT 644
JN378393	TTAACCTGAGTGCAGAACAGGGAGAGTG-AATTCCATGTGTAGCGGT---AGCGTAGAT 705
JN652127	ATCTGGAGGAATACCGTGGCGAAGGCCGGCCCCCTGGACGAAGACTGACGCTCAGGTGCG 748
JN652128	-----
JN378392	-----
HQ404365	ATATGGAGGAACACCGTAGCGAAGCGACTTTCTGGCTGTAACTGACGCTGATGTGCG 747
JQ039348	ATCTGGAGGAATACCGATGGCGAAGGCAGCTTCTGGCATCATACTGACACTGAGGTTCG 719
JN652129	-----
JQ039350	-----G 1
HM204502	ATCAGGAGGAACACCGATGGCGAAGGCAGGTCTCTGGCTGTAACTGACGCTACGGAGCG 676
JQ039349	ATCAGGAGGAACGCCAGTGGCGAAGGCAGGTCTCTGGCCTTCCTGAC----- 693
JN378393	ATATGGAGGAACACCGTGGCGAAAGCGGCTCTGGCTGTAACTGACGCTGAGGCTCG 765

Fig. 1: Continue

JN652127	AAAGCGTGGGAGCAAACAG---GATTAGATAACCCTGATAGTCACGCCGTAAACGATGT	805
JN652128	-----	
JN378392	-----	
HQ404365	AAAGCGTGGGATCAAACAG---GATTAGATAACCCTGGTAGTCCACGCCGTAAACGATGA	804
JQ039348	AAAGCGTGGGTAGCAAACAG---GATTAGATAACCCTAGTAGTCCACGCCGTAAACGATGT	776
JN652129	-----	
JQ039350	AAAGCGTGGGAGCAGACAG---GATTAGATAACCCTGGTAGTCCACGCCGTAAACGATGT	58
HM204502	AAAGCATGGGAGTGAACAG---GATTAGATAACCCTGGTAGTCCACGCCGTAAACGTTGG	733
JQ039349	-----	
JN378393	AGAGCGTGGGAGCGAACAGGAAGGTTAGATAACCCTGGTAGTCCACGCCGTAAACGATGA	825
 JN652127	CGACTTGGAGGTTG-TGCCCTTGAGGGG-TGGCTTCCGGAGCTAACCGGTTAAG-TCGAC	862
JN652128	-GATT CGGAGGTTG-TGCCCTTGAGGCAGTGGCTTCCGGAGCTAACCGGTTAAAATCGAC	58
JN378392	-----	
HQ404365	GTGCTAAGTGTAGGGGTTCCGCCCTTAG-TGCTGCAGCTAACGCAATTAAAG-CACTC	862
JQ039348	CTACCAGTCGTTGGGTCTCTGAAG---ACTTAGTGACGCAGTTAACGCAATAAG-TAGAC	833
JN652129	-----	
JQ039350	CGACTAGCCGTGG-GATCCTTGAGATCTTAGTG-GCGCAGCTAACGCGATAAG-TCGAC	115
HM204502	GCAC TAGGTGTGGGACCATTCACGGTTCCGC CGCAGCTAACGCAATTAAAG-TGCC	792
JQ039349	---CTAGGC-----	699
JN378393	GTGCTAGGTGTTAGGCCCTTCCGGGCTTAG-TGCCGCAGCTAACGCAATTAAAG-CACTC	883
 JN652127	CGCCTGGGAGTACGCCCGCAAGGTTAAACTCAAATGAGTTGACGGGGCCCGACAAG	922
JN652128	CGCCTGGGAGTACGCCCGCAAGGTTAAACTCAAATGAATTGACGGGGCCCGACAAG	118
JN378392	-----	
HQ404365	CGCCTGGGAGTACGCCCGCAAGGTTAAACTCAAAGGAAATTGACGGGGCCCGACAAG	922
JQ039348	CGCCTGGGAGTACGCCCGCAAGGTTAAACTCAAATGAATTGACGGGGCCCGACAAG	893
JN652129	-----	
JQ039350	TGCCTGGGAGTACGCCCGCAAGGTTAAACTCAAATGAATTGACGGGGCCCGACAAG	175
HM204502	CGCCTGGGAGTACGCCCGCAAGGCTAAACTCAAAGGAAATTGACGGGGCCCGACAAG	852
JQ039349	-----	
JN378393	CGCCTGGGAGTACGCCCGCAAGGTTAAACTCAAAGGAAATTGACGGGGCCCGACAAG	943
 JN652127	CGGTGGAGCATGTGGTTAATTGATGCAACCGCAGGAATCTTACCTGGTCTTGACATCC	982
JN652128	CGGTGGAGCATGTGGTTAATTGATGCAACCGCAGGAACCTTACCTGGTCTTGACATCC	178
JN378392	-----	
HQ404365	CGGTGGAGCATGTGGTTAATTGCAAGCAACCGCAGGAACCTTACCAAATCTTGACATCC	982
JQ039348	CGGTGGAGCATGTGGTTAATTGATGCAACCGCAGGAACCTTACCTGGTCTTGACATAG	953
JN652129	-----	
JQ039350	CGGTGGAGCATGTGGTTAATTGCAAGCAACCGCAGGAACCTTACCTGGCCTTGACATGC	235
HM204502	CGGCGGAGCATGCCGATTAATTGATGCAACCGCAGGAACCTTACCAAGGCTTGACATGT	912
JQ039349	-----	
JN378393	CGGTGGAGCATGTGGTTAATTGCAAGCAACCGCAGGAACCTTACCAAGGCTTGACATCC	1003
JN652127	ACAG-AAGTTKTCAGAGATGAGMATGTGCCCTCGGGA--ACCGTGAGACAGGTGCTGCAT	1039
JN652128	ACAG-AACTTAGCAGAGATGGTTGGTGCCTTCGGGA--ACTGTGAGACAGGTGCTGCAT	235
JN378392	-----	
HQ404365	T-CTGACCCCTAGAGATAGAGTTCCCTTCGGGGACAGAGTGACAGGTGGTGCAT	1041
JQ039348	TGAGGATC-CTGCAGAGATGCCGGAGTGCCTTCGGGA--ATCCACATACAGGTGCTGCGT	1010

Fig. 1: Continue

JQ039350	TGAG-AACTTCCAGAGATGGATTGGTGCCTCGGGA--ACTCAGACGCAGGTGCTGCAT 292
HM204502	TCTCGATGCCGTAGAGATAACGG-TTTCCCCTTGGG--GCGGGTTCACAGGTGGTGCAT 969
JQ039349	-----
JN378393	CGATGCCCGCTCTAGAGATAAGAGTTT--ACTTCGGTACATCG-GTGACAGGTGGTGCAT 1060
JN652127	GGCTGTCGTAGCTCGTGTGAAATGTTGGGTTAAGTCCCGAACGAGCGCAACCCTT 1099
JN652128	GGCTGTCGTAGCTCGTGTGAAATGTTGGGTTAAGTCCCGAACGAGCGCAACCCTT 295
JN378392	-----
HQ404365	GGTTGTCGTAGCTCGTGTGAGATGTTGGGTTAAGTCCCGAACGAGCGCAACCCTT 1101
JQ039348	GGCTGTCGTAGCTCGTGTGAGATGTTGGGTTAAGTCCCGAACGAGCGCAACCCTT 1070
JN652129	-----
JQ039350	GGCTGTCGTAGCTCGTGTGAGATGTTGGGTTAAGTCCCGAACGAGCGCAACCCTT 352
HM204502	GGTTGTCGTAGCTCGTGTGAGATGTTGGGTTAAGTCCCGAACGAGCGCAACCCTC 1029
JQ039349	-----
JN378393	GGTTGTCGTAGCTCGTGTGAGGTGTTGGGTTAAGTCCCGAACGAGCGCAACCCTT 1120
JN652127	ATCCTTTGTTGCCAGCGGTCGG-GCCGGGAACCTCAAAGGAGGCTGCCAGTGATAAACTGG 1158
JN652128	ATCCTTTGTTGCCAGCGGTTGG-GCCGGGAACCTCAAAGGAGGACTGCCAGTGATAAACTGG 354
JN378392	-----
HQ404365	AAGCTTAGTTGCCA--TCATTAAGTTGGGCACTCTAAGTTGACTGCCGGTGACAAACCGG 1159
JQ039348	TTCCCTTAGTTAACAGCACTTCG-GGTGGGAACCTCAAGGATACTGCCAGTGACAAACTGG 1129
JN652129	-----
JQ039350	GTCCTTAGTTACAGCACCTCG-GGTGGGCACTCTAAGGAGACTGCCGGTGACAAACCGG 411
HM204502	GTTCCATGTTGCCAGCACGTCGTGGGGACTCATGGGAGACTGCCGAAGTCAACTCGG 1089
JQ039349	-----
JN378393	ATTGTTAGTTGCCA--TCATTCAGTTGGGCACTCTAGCGAGGACTGCCGGTAATAAACCGG 1178
JN652127	AGGAAGGTGGGATGACGTCAAGTCGTATGGCCCTTACGACCAGGGCTACACACGTGCT 1218
JN652128	AGGAAGGTGAAATGACGTCAAGTCATCATGGCCCTTATGACCAGGGCTACACACGTGCT 414
JN378392	-----
HQ404365	AGGAAGGTGGGATGACGTCAAATCATCATGGCCCTTATGATTGGGCTACACACGTGCT 1219
JQ039348	AGGAAGCGGGGACGACGTCAAGTCATCATGGCCCTTACGACCAGAGCTACACACGTGCT 1189
JN652129	-----
JQ039350	AGGAAGGTGGGATGACGTCAAGTCATCATGGCCCTTACGGCCAGGGCTACACACGTGCT 471
HM204502	AGGAAGGTGAGGACGACGTCAAATCATCATGGCCCTTATGCTTGGGCTTCACGCATGCT 1149
JQ039349	-----
JN378393	AGGAAG----- 1184
JN652127	ACAATGGCATATAACAAAGAGAAGCGACCTCGCGAGAGCAAGCGGACCTCATAAAGTATGT 1278
JN652128	ACAGTGGCATATAACAAAGAGAAGCGACCTCGCGAGAGCAAGCGGACCTCATAAAGTATGT 474
JN378392	-----
HQ404365	ACAATGGACAATAACAAAGGGCAGCGAAACCGCGAGGTCAAGCAAATCCCATAAAGTTGTT 1279
JQ039348	ACAATGGTTGGTACAAAGGGTTGCTACACAGCGATGTGATGCTAATCTCAAAAAGCCAAT 1249
JN652129	-----
JQ039350	ACAATAGTCATAACAAAGGGTTGCCAAGCCGCGAGGTGGAGCTAATCCCATAAAACCGAT 531
HM204502	ACAATGGCCGGTACAATGGGTTGCGATACTGTGAGGTGGAGCTAATCCCAAAAGCCGGT 1209
JQ039349	-----
JN378393	-----

Fig. 1: Continue

JN652127	CGTAGTCCGGATTGGAGTCTGCAGCTGACTCCATGAAGTCGGAATCGCTAGTAATCGTG	1338
JN652128	CGTAGTCCGGATTGGAGTCTGCAACTCGACTCCATGAAGTCGGAATCGCTAGTAATCGTA	534
JN378392	-----	
HQ404365	CTCAGTTGGATTGTAGTCTGCAACTCGACTATATGAAGCTGGAATCGCTAGTAATCGTA	1339
JQ039348	CGTAGTCCGGATTGGAGTCTGCAACTCGACTCCATGAAGTCGGAATCGCTAGTAATCGCA	1309
JN652129	-----	
JQ039350	CGTAGTCCGGATCGCAGTCTGCAACTCGACTGCGTGAAGTCGGAATCGCTAGTAATCGTG	591
HM204502	CTCAGTCCGGATTGGGGCTGCAACTCGACCCCATGAAGTCGGAAGTCGCTAGTAATCGCA	1269
JQ039349	-----	
JN378393	-----	
 JN652127	GATCAG-AATGCCACGGTGAATACGTCCCCGGGCCTGTACAACCG-CCCGTCACACCAT	1396
JN652128	GATCAG-AATGCTACGGTGAATACGTCCCCGGGCCTGTACACACGGCCCGTCACACCAT	593
JN378392	-----	
HQ404365	GATTAGCATG-CTACGGTGAATACGTCCCCGGGCCTGTACACACCGCCCGTCACACCAC	1398
JQ039348	GATCAG-AATGCTGCGGTGAATACGTCCCCGGGCCTGTACACACCGCCCGTCACACCAT	1368
JN652129	-----	
JQ039350	AATCAG-AATGTCACGGTGAATACGTCCCCGGGCCTGTACACACCGCCCGTCACACCAT	650
HM204502	GATCAGCAACGCTGCGGTGAATACGTCCCCGGGCCTGTACACACCGCCGGTCAAGTCAC	1329
JQ039349	-----	
JN378393	-----	
 JN652127	GGGAGTGGTTGCAAAAGAAGTAGGTAGCTAACCTTCGGGAGGGCGCTTACCAACTTGT	1456
JN652128	GGGAGTGGTTGCAAAAGAGGTAGGTAGCT-AACCTTCGGGAGGGCGCTTACCAAGG--	649
JN378392	-----	
HQ404365	GAGAGTTGATCTCACCAAGAAGTGGTTAGCTAACGCAAGAGGGCGATCACACGGTGGGG	1458
JQ039348	GGGAGTTGATCTCACCAAGAAGTGGTTAGCTAACGCAAGAGGGCGATCACACGGTGGGG	1428
JN652129	-----	
JQ039350	GGGAGTGGTTGCTCCAGAAGTAGCTAGTCTAACCG-CAAAGGAAACGG-TACCA-----	702
HM204502	GAAAGTCGTTAACACCCGAAGCCGGTGGCCTAACCAAAGTCGTTAACACCCAAGCCGGTG	1389
JQ039349	-----	
JN378393	-----	
 JN652127	GATTCACTGAGGTGAAGTCGTATCAAGGTAAACCGTAGG	1497
JN652128	-----	
JN378392	-----	
HQ404365	AACAAATGATTGGG-----	1472
JQ039348	TCGATGACTGGAGTGG-----	1445
JN652129	-----	
JQ039350	-----	
HM204502	GCCTAAC-----	1396
JQ039349	-----	
JN378393	-----	

Fig. 1: Multiple sequence alignment of 16S rDNA sequences of bacteria causing keratitis

**GeneBee-secondary structure prediction:** In the multiple alignment the algorithm used by GeneBee produces structures through the minimization of the potential energy of the system. The algorithm divides a sequence S, into subsequences (S<sub>ij</sub>) and calculates the minimal energy for two cases. W(i, j) defines the minimal free energy of all allowed structures formed from S<sub>ij</sub> and V(i, j)

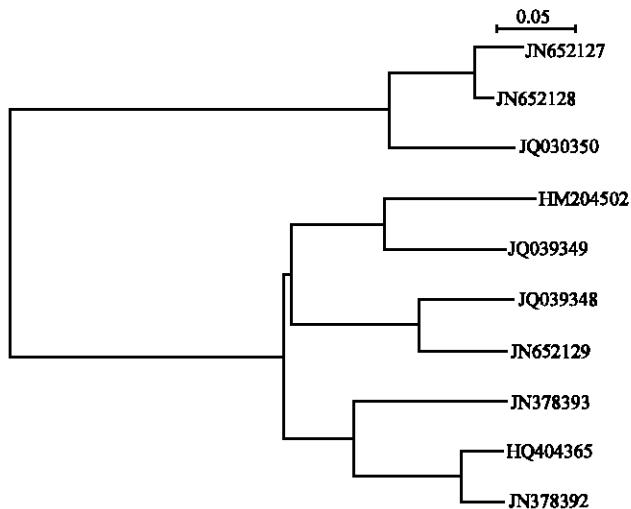


Fig. 2: Phylogenetic analysis of 16S rDNA sequences of bacteria causing keratitis

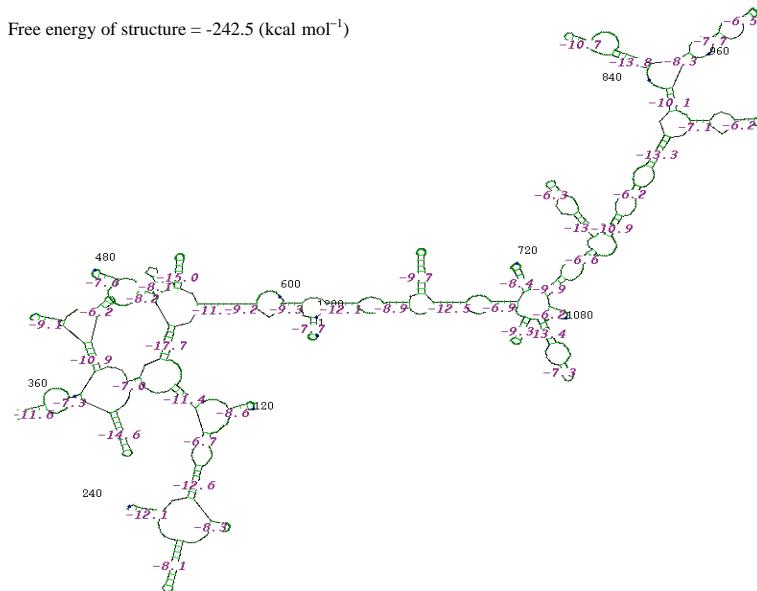


Fig. 3: Structure of *Streptococcus viridan*

the minimum energy if  $S_i$  and  $S_j$  pair with each other. The second term is set to infinity if  $S_i$  and  $S_j$  cannot base pair. Starting with sequences of five nucleotides,  $W(i, j)$  and  $V(i, j)$  are calculated recursively for longer and longer sequences, selecting the optimal structure at each step. The final computation considering the entire sequence is the result.  $W(i, j)$  and  $V(i, j)$  are calculated based upon the stability of base pairs formed and the resulting structures: hairpin loops, stacking regions, bulge loops, interior loops and bifurcation loops as defined by Zuker and Stiegler (1981).

The thermostabilities of the secondary structure were calculated with online software ([http://www.genebee.msu.su/services/rna2\\_reduced.html](http://www.genebee.msu.su/services/rna2_reduced.html)) (Fig. 3-12). The thermostability energy was calculated as -242.5, -74.4, -307.3, -275.2, -326.3, 46.7, -161.2, -234.8, -131.7 and -293.4 kcal mol<sup>-1</sup>, for *Streptococcus viridans*, *Staphylococcus aureus*, *Micrococcus* sp.,

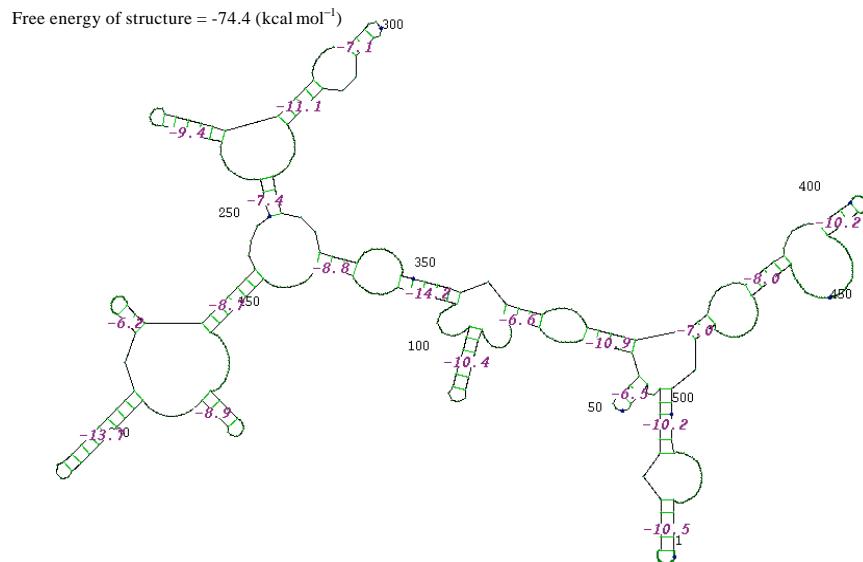


Fig. 4: Structure of *Staphylococcus aureus*

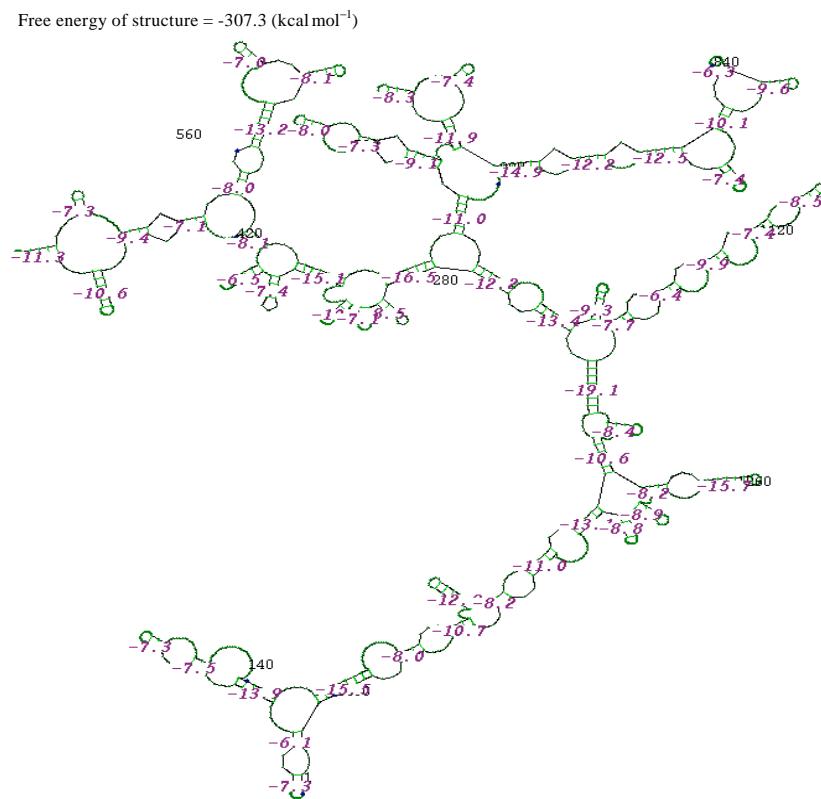


Fig. 5: Structure of *Micrococcus* sp.

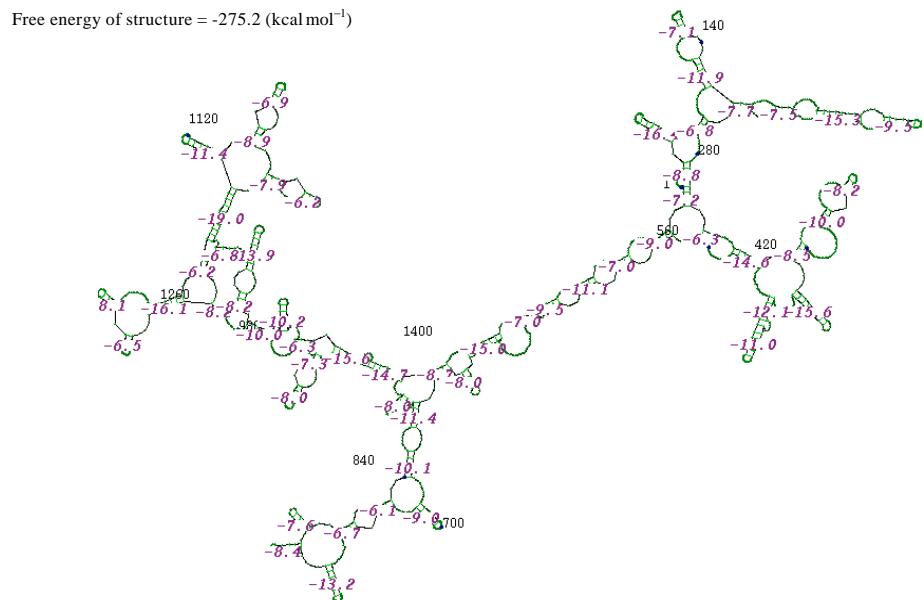


Fig. 6: Structure of *Moraxella* sp.

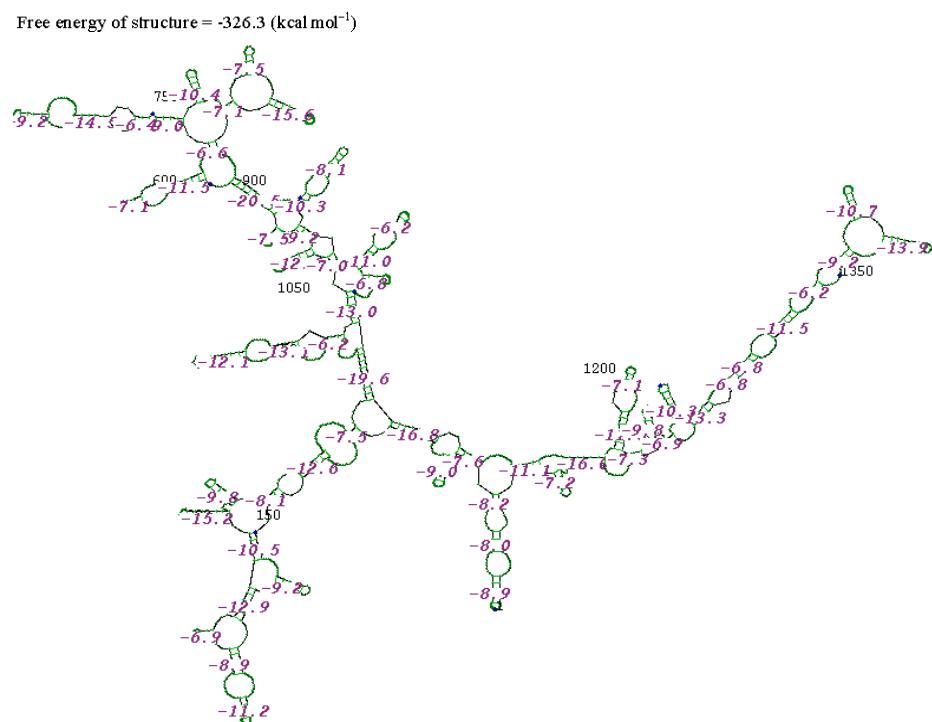


Fig. 7: Structure of *Citrobacter koseri*

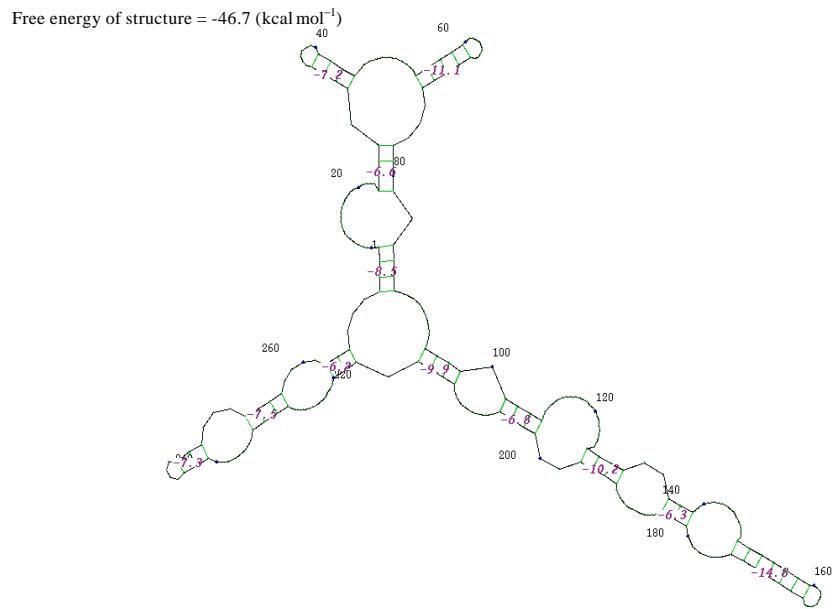


Fig. 8: Structure of *Acinetobacter baumannii*

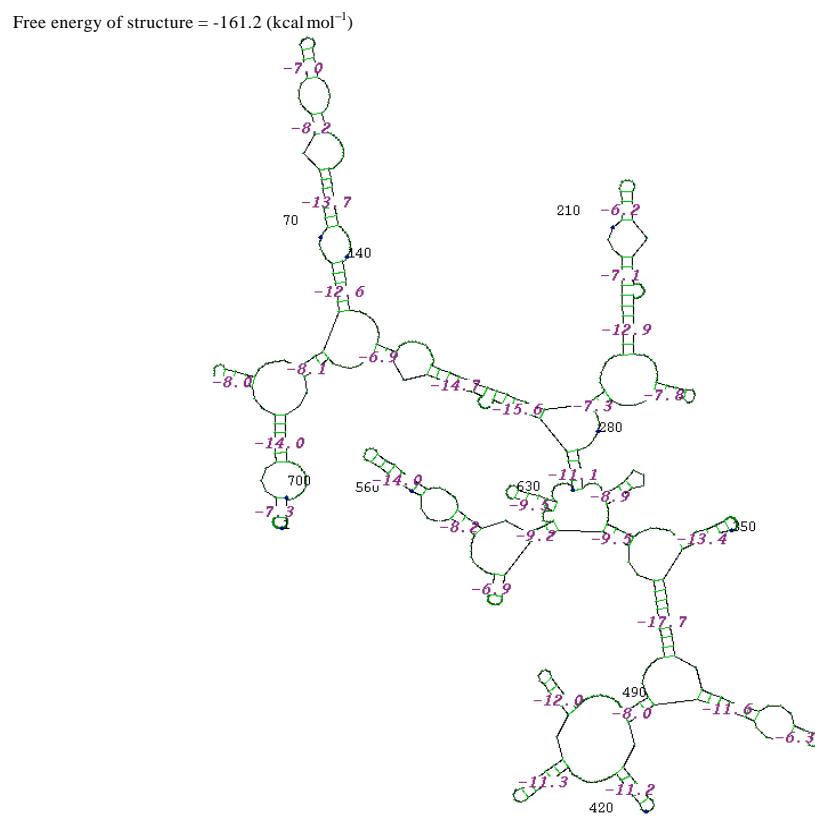


Fig. 9: Structure of *Pseudomonas aeruginosa*

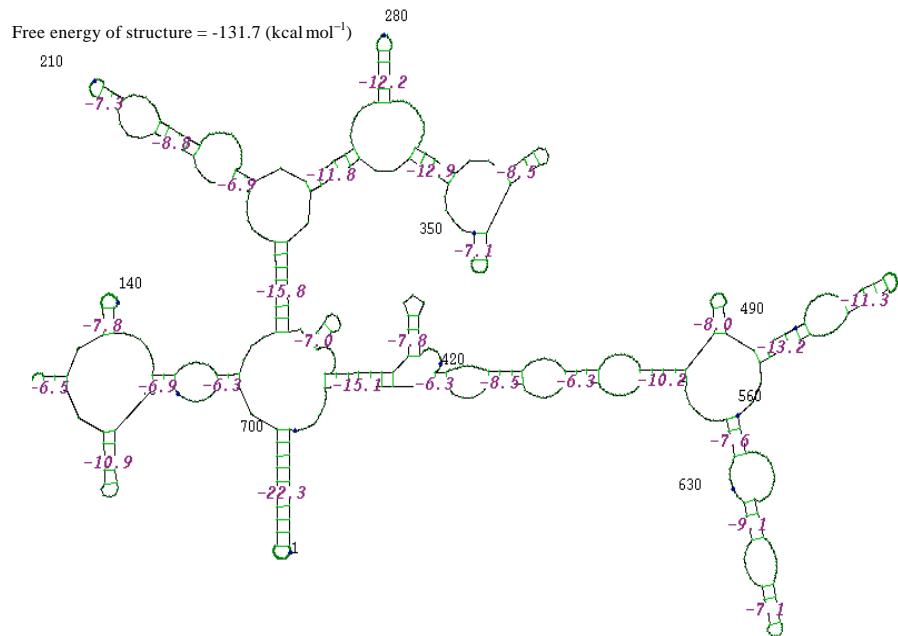


Fig. 10: Structure of *Propionibacterium* sp.

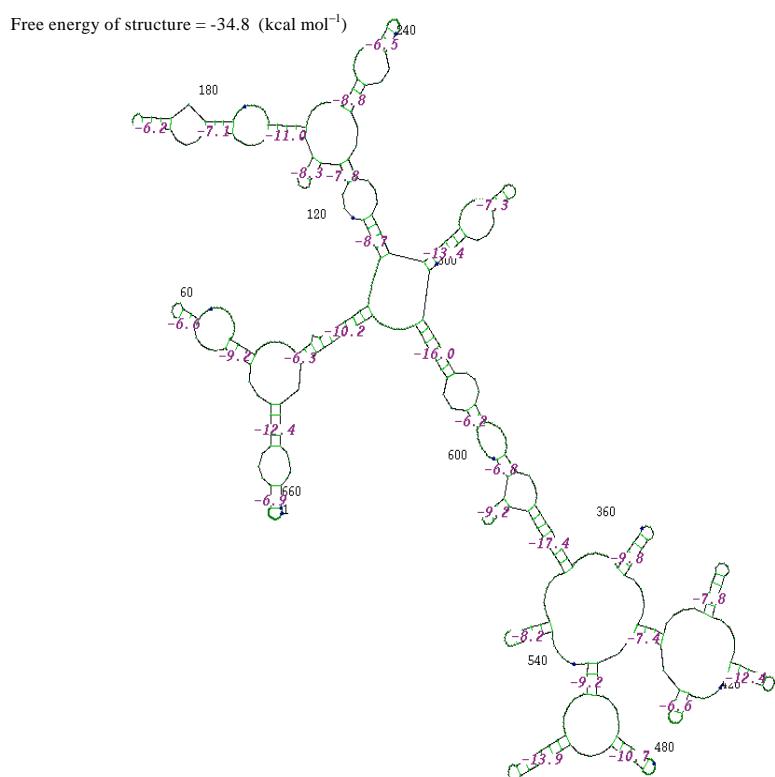


Fig. 11: Structure of *Klebsiella* sp.

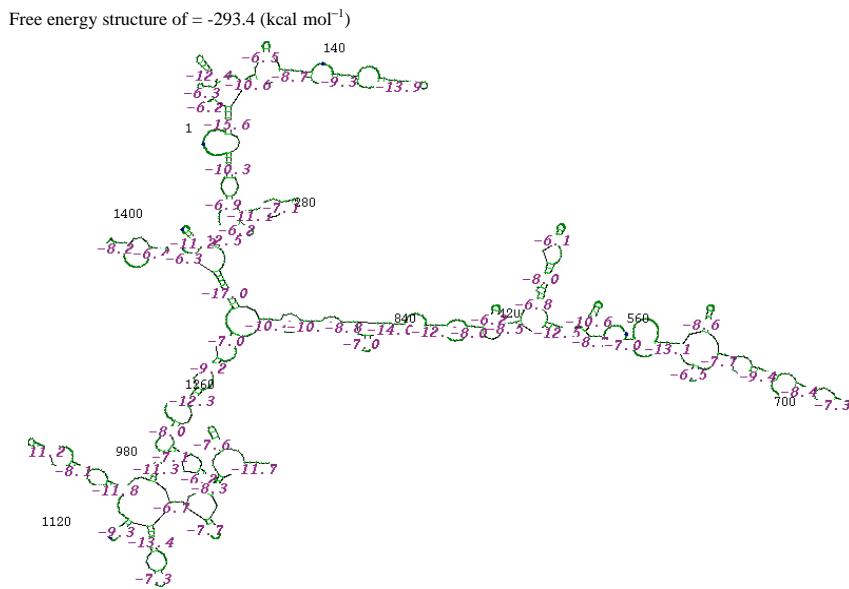


Fig. 12: Structure of *Staphylococcus epidermidis*

Table 2: Secondary structure prediction of rDNA for bacterial sequences by using GeneBee tool

Microorganism	Free energy of structure (kkal mol <sup>-1</sup> )
<i>Streptococcus viridians</i>	-242.5
<i>Staphylococcus aureus</i>	-74.4
<i>Micrococcus</i> sp.	-307.3
<i>Moraxella</i> sp.	-275.2
<i>Citrobacter koseri</i>	-326.3
<i>Acinetobacter baumannii</i>	-46.7
<i>Pseudomonas aeruginosa</i>	-161.2
<i>Klebsiella</i> sp.	-234.8
<i>Propionibacterium</i> sp.	-131.7
<i>Staphylococcus epidermidis</i>	-293.4

*Moraxella* sp., *Citrobacter koseri*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, *Klebsiella* sp., *Propionibacterium* sp., *Staphylococcus epidermidis*, respectively (Table 2). The determination of the structure of the RNA molecule is very important in the determination of its activity.

In the secondary structure prediction of RNA the fold with the more negative free energy, is more stable. From the result, the *Citrobacter koseri*, (-326.3 kkal mol<sup>-1</sup>) has lower minimal free energy. When compared with others species whereas the *Acinetobacter baumannii*, (-46.7 kkal mol<sup>-1</sup>) has high minimal free energy when compared with other species. RNA molecule must be energetically stable to be able to maintain their structure and perform their functions.

rDNA-based analysis is a central method in microbiology, used as a method for bacterial strain identification. Molecular methods, including PCR, genotyping and automated DNA sequencing, are becoming more available and hence their adoption into routine medical microbiology is increasing. Therefore, 16S rDNA is known to be highly conserved in species and it could also be used for the

verification of the thermodynamic stability on the basis of conserved secondary structure of RNA. The structure of RNA plays an important role in the life cycle of bacteria and provides the ability to understand evolution and stability (Zuker and Stiegler, 1981). Earlier studies reported that Molecular Characterization and Sequencing of a Gene Encoding Mannose Binding Protein in an Iranian Isolate of *Acanthamoeba castellanii* as a major agent of *Acanthamoeba* keratitis by PCR techniques (Niyyati *et al.*, 2008).

In this study, we have characterized the bacteria causing keratitis and analyzed the phylogenetic relationship between them. The Multiple sequence analysis reveals that this sequence is highly conserved as all the organism share a region which is highly similar. The rDNA has also been modeled and their free energies have been predicted.

## CONCLUSION

All bacterial species possess at least one copy of the 16S rDNA gene which contains highly conserved as well as hyper variable nucleic acid sequences. Therefore, the PCR targeting of the conserved nucleic acid sequence of 16S rDNA gene of bacterial isolate is used as a molecular tool to identify bacteria causing keratitis.

In conclusion, identification and sequencing of this important gene is the first step to pursue future research such as developing better therapeutic agents, immunization of population at risk or even developing a rapid diagnostic tool by PCR techniques. The sequence analysis of the gene fragment characterized in this study is still under investigation. This technique provides the opportunity to identify non-cultivable, uncommon, or even unknown causative bacteria.

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