

Reinforcement of Condensation Reaction using DCCI to Synthesis New Compounds Derived from Phthalyl DL-Leucine and Sulfa Drugs

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Abstract: New compounds derived from N-phthalyl DL-leucine as amino acid derivative and some kinds of sulfa drugs like sulfanilamide, sulfaisoxazole, sulfamethaxazole, sulfadiazine, sulfathiazole, sulfapyridine and sulfacetamide sodium were prepared by utilizing N, N-Dicyclohexylcarbodiimide (DCCI) as condensing agent to promoted condensation reaction to appointed the corresponding amide. All the synthesized compounds have been characterized by elemental analysis (C, H, N, S), FT-IR and H NMR. The antimicrobial activity of these compounds were determined against two kinds of bacteria gram positive (*Staphylococcus aureus*) and gram negative (*Aeromonas veronii*) by using different concentration of each synthesized compound in addition the MIC was determined.

Key words: N-phthalyl, DL-leucine, sulfa drugs, DCCI, amide, sulfamethaxazole, antimicrobial

INTRODUCTION

Amino acids constitute an important class of nitrogen-containing, naturally occurring compounds. Also it can be called α -amino acid because every amino acid has a carboxyl group $-\text{COOH}$ and an amino group NH_2 and each group can exist in an acidic form or a basic form depending on the Ph of the solution in which the amino acid is dissolved. The biological function of amino acids is well established they are the constituent monomer units from which the biopolymers based on the peptide bond are built, for example, polypeptides and proteins including enzyme (Fox and Whitesell, 2004). Peptides and proteins are polymers of amino acids linked together by amide bond. Gramicidin S is an example of antibiotic peptide produced by a strain of bacteria, it is a cyclic decapeptide that contains Ornithine and some amino acids such as L-phenylalanine, L-valine and L-Leucine (Bruce, 2004). Another examples of amino acids derivatives is penicillin (Lipmann *et al.*, 1941). Moreover the amino acids can enter into many reaction with some metals to form many types of complexes (Al-Salami *et al.*, 2017). The first antibiotics are sulfonamide commonly known as Sulfa drugs were introduced clinically in 1934 as the first effective antibiotics. Sulfa drugs such as sulfanilamide is a bacteriostatic drug which inhibits the further growth of bacteria.

The effect of the sulfa drugs is inhibition the bacterial enzyme that synthesizes folic acid of that bacteria (Otten, 1986, Tilles, 2001). Sulfa drugs are widespread among many compounds which possess many requisition like diuretic or antiglaucoma, also many kinds of sulfa drugs such as sulfadiazine used for treatment of

anthrax and urinary tract infection and prevent incidence of a meningococcal carriers (Hassan *et al.*, 1974; Schabel *et al.*, 1946).

The newfangled study of this research localize about the formation of amide from amino acid derivative, phthalyl DL-leucine and selected sulfa drugs by using N, N-Dicyclohexylcarbodiimide (DCCI) as condensing agent to furnish the coinciding amide as we explained in our previous research (Al-Salami, 2009; Al-Salami *et al.*, 2014). Linking the carboxylic terminal of phthalyl DL-Leucine with the NH_2 -terminal of sulfa drugs together requires the formation of an amide. This formation of an amid bond can be accomplished in a number way but all require that the carboxylic acid be converted to an activated acyl derivative that is more susceptible to nucleophilic acyl substitution. A particularly good choice for the acyl derivative is one formed from (DCCI) and the carboxylic acid.

Nucleophilic acyl substitution takes place in the normal addition elimination fashion and produces a urea (derived from carbodiimide) in addition to the desired amide (Al-Salami, 2009; Al-Salami *et al.*, 2014).

MATERIALS AND METHODS

Experimental

Materials and reagents: All the chemicals and solvent used were of Analytical grade (AR) and of highest purity obtainable which included phthalicanhydride (MERCK) DL-Leucine (MERCK), all kinds of sulfa drugs (Aldrich). All solvent used equipped by (BDH) company. The measurements of melting point were done on Bauchi 510.

Instrumentation: The solid state FTIR spectra of the compounds were recorded on Shimadzu FT-IR Model 8400 S spectrophotometer using KBr pellets in the range 4000-400 cm⁻¹. The spectra of HNMR were done in a Bruker spectrophotometer (400 MHz) and using DMSO-d₆ as solvent and TMS as internal standard. Elemental analysis (C, H, N, S) were recorded by Euro vector model 3000 A (Italy).

Preparation of N-Phthalyl DL-Leucine: In this study, we prepared N-Phthalyl DL-Leucine by the method described by Ing and Fox (Doering and Weil, 1947). An equimolar amount of DL-Leucine and finely ground phthalic anhydride are heated in oil-bath at 150 for 25 min until frothing ceased. After cooling, the light brown mass was extracted with boiling ether. Hexane was added to the filtered ether extracts until the appearance of white precipitation, recrystallized from cyclohexane. The N-phthalyl DL-Leucine was prepared (melting point) was (118-119) with yield 83%.

Synthesis of compounds (F1-F7): The new compounds (F1-F7) were prepared from conforming phthalyl DL-leucine and different types of sulfa drugs according to literature procedure curini (Curini *et al.*, 2002). The preparation methods for a composite can be illustrated as follows:

- F1: 2-(1, 3-dioxoisindolin-2-yl)-4-methyl-N-(4-sulfamoylphenyl) pentanamide
- F2: N-(4-(3, 4-dimethylisoxazole-5-yl) sulfamoyl) phenyl-2-(1,3-dioxoisindoline-2-yl)-4-methyl pentanamide

- F3: 2-(1,3-dioxoisindolin-2-yl)-4-methyl-N-(4-(N-5-methyl-isoxazol-3-yl) sulfamoyl) phenyl) pentanamide
- F4: 2-(1,3-dioxoisindoline-2-yl)-4-methyl-N-(4-(N-thiazole-2-yl)sulfamoyl) phenyl) pentanamide.
- F5: Sodium acetyl(4-(2-(1, 3-dioxoisindoline-2-yl)-4-methyl pentanamido) phenyl) sulfamoyl) amide
- F6: 2-(1, 3-dioxoisindoline-2-yl)-4-methyl-N-(4-(N-(pyridin-2-yl) sulfamoyl) phenyl) pentanamide
- F7: 2-(1, 3-dioxoisindoline-2-yl)-4-methyl-N-(4-(N-(pyrimidin-2-yl) sulfamoyl) phenyl) pentanamide

These compounds (F1-F7) were prepared by mixing the congruous phthalyl DL-leucine 1 mmol (0.1722 g) with 1 mmol of sulfanilamide (0.261 g), sulfaisoxazole (0.267 g), sulfamethaxazole (0.253 g), sulfathiazole (0.2553 g), sulfacetamide sodium (0.236 g), sulfapyridine (0.249 g) and sulfadiazine (0.250 g), respectively in 50 mL of ethylacetate, to this stirred solution has been added dropwise solution of DCCI (0.206 g, 1 mmol) in 5 mL ethylacetate over period 20 min at room temperature with continuous stirring until white precipitate formed (dicyclohexylurea). The solid precipitate was filtered off and the filtrate was washed with 5% citric acid solution (10 mL), two layers formed and separation by extraction. The aqueous layer was neglected and the organic layer was taken. The solvent was evaporated and the residue was purified by silica gel column chromatography (eluent ethylacetate: n-hexane 2:8). The structural formula, elemental analysis and physical properties were listed and conferred in Table 1. The synthetic compounds were summarized in Fig. 1.

In vitro antimicrobial activity: The biological activity of the new compounds (F1-F7) were studied against two types of bacteria which included positive bacteria (*Staphylococcus aureus*) and gram negative bacteria

Table 1: Symbol, structural formula, IUPAC name, analytical and physical data of the synthetic compounds

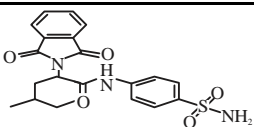
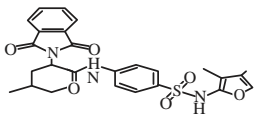
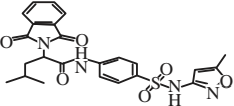
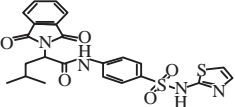
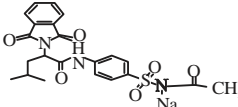
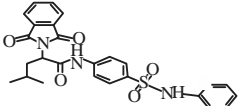
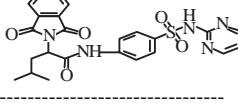
Com. symbol	Structural formula IUPAC name	Color	Molecular weight	Mol. formula	M.P (°C)	Yield (%)	Elemental analysis CHNS partical (theoretical)			
							C (%)	H (%)	N (%)	S (%)
F1	 2-(1, 3-dioxoisindolin-2-yl)-4-methyl-N-(4-sulfamoylphenyl) pentanamide	Pale yellow	415.46	C ₂₀ H ₂₁ N ₃ O ₅ S	140	72	57.32 57.82	5.24 5.09	10.24 10.11	7.79 7.72
F2	 N-(4-(N-(3, 4-dimethylisoxazol-5-yl)sulfamoyl)phenyl)-2-(1, 3-dioxoisindolin-2-yl)-4-methyl-N-(4-sulfamoylphenyl) pentanamide	Pale yellow	510.56	C ₂₅ H ₂₆ N ₄ O ₆ S	93	58	59.21 58.81	5.29 5.13	11.09 10.97	6.67 6.28

Table 1: Continue

Com. symbol	Structural formula		Molecular weight	Mol. formula	M.P (°C)	Yield (%)	Elemental analysis CHNS partical (theoretical)			
	IUPAC name	Color					C (%)	H (%)	N (%)	S (%)
F3	dioxoisindolin-2-yl)-4-methyl pentanamide		496.54	C ₂₄ H ₂₄ N ₄ O ₆ S	85	52	58.68	5.22	11.39	6.58
		Pale yellow					58.05	4.87	11.28	6.46
F4	2-(1, 3-dioxoisindolin-2-yl)-4-methyl-N-(4-(N-(5-methylisoxazol-3-yl) sulfamoyl) phenyl) pentanamide		498.57	C ₂₃ H ₂₂ N ₄ O ₅ S ₂	105	60	55.42	4.56	11.36	12.99
		Pale yellow					55.41	4.45	11.24	12.86
F5	2-(1, 3-dioxoisindolin-2-yl)-4-methyl-N-(4-(N-(thiazol-2-yl) sulfamoyl) phenyl) pentanamide		479.48	C ₂₂ H ₂₂ N ₃ NaO ₆ S	125	71	55.30	4.70	8.91	7.01
		Yellowish white					55.11	4.62	8.76	6.69
F6	sodium acetyl((4-(2-(1, 3-dioxoisindolin-2-yl)-4-methylpentanamido) phenyl) sulfonyl) amide		492.55	C ₂₅ H ₂₄ N ₄ O ₅ S	80	67	60.06	4.99	11.49	6.70
		Pale yellow					60.96	4.91	11.37	6.51
F7	2-(1, 3-dioxoisindolin-2-yl)-4-methyl-N-(4-(N-(pyridin-2-yl) sulfamoyl) phenyl) pentanamide		493.53	C ₂₄ H ₂₃ N ₅ O ₅ S	83	76	58.59	4.81	14.32	6.69
		Yellowish white					58.41	4.70	14.19	6.50
	2-(1, 3-dioxoisindolin-2-yl)-4-methyl-N-(4-(N-(pyrimidin-2-yl) sulfamoyl) phenyl) pentanamide									

(*Aerompnas veroni*) by using agar plate diffusion (Al-Salami, 2018). Dimethyl sulfoxide was used as a solvent and as control for disc sensitivity. The antibiotic tetracycline 25 mg L⁻¹ was used for the purpose of calibration and comparison with the bacteria stuff. Bacteria have been cultured by providing the appropriate habitat for their growth at 37°C for 24 h. This method involves the detection of the zone of inhibition toward the diffusion of microorganism on agar plate. The solution of compounds (F1-F7) were prepared in different concentration to estimated the

antibacterial activity by using 0.1 mL of particular bacterial culture. The plate were incubated for 24 h at 37°C (Al-Salami, 2018). Zone of inhibition were determined in mm. Also, the values of Minimum Inhibitory Concentration (MIC) of these compounds were calculated (Al-Salami, 2018). The MIC can be defined as the lowest concentration of the compound in a media which prevent visible growth of the test bacteria in each concentration (25-300 mg L⁻¹). The MIC is determined by preparing solution of the chemical *in vitro* at increasing concentrations incubating

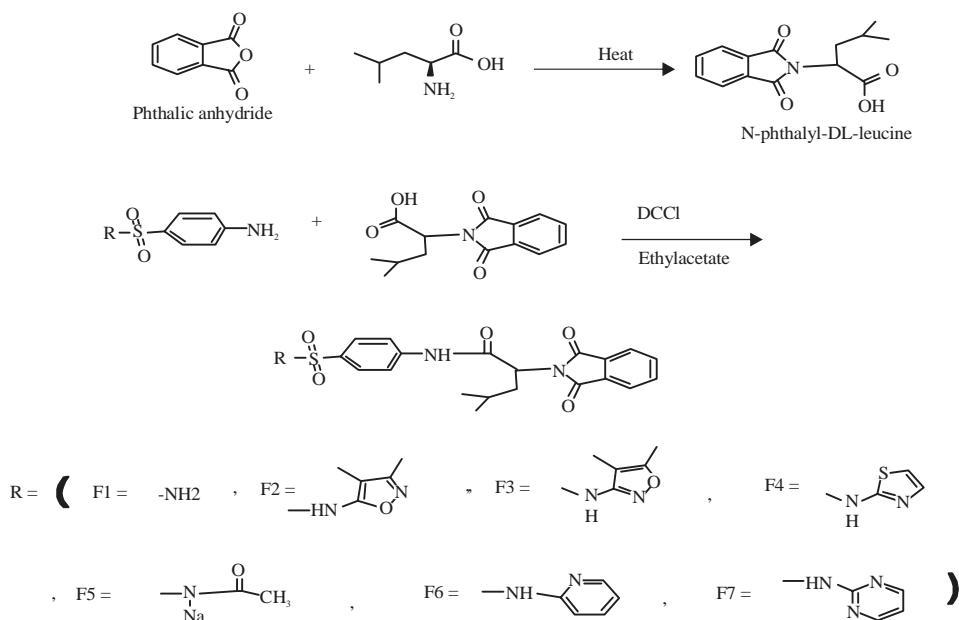


Fig. 1: The synthetic compounds

the solutions with the separate batches of cultured bacteria, and measuring the results using agar dilution or broth microdilution (Andrews, 2001).

RESULTS AND DISCUSSION

In this research, focus on synthesis of amide from amino acids were these amino acids are classified as a kind of carboxylic acid. The amide compounds are one of the important derivatives of carboxylic acid. The reaction and mechanism of formation of amide from amino acid has already been explained in our previous work (Al-Salami, 2009; Al-Salami *et al.*, 2014). The mechanism shows that the DCCI works as condensing agent to equip the compatible amide. One of the important analysis in this research was elemental analysis and the outcomes of it shows in Table 1, we will find that there is a great match with the propose structure.

FT-IR spectra: FT-IR spectra data are listed in Table 2 and Fig. 2-4. Derivatives of amino acids such as phthalyl amino acids are characterized by appearance of OH band but when configured the amide from this derivatives we saw disappearance of the OH band and instead of that the new amides display a strong band at the range (3381-3327 cm^{-1}) which attributed to NH group of amides which attributed to the range 3381-3327 cm^{-1} of new amides. Also the formation of amide can be characterized by appearance of amide II (νNH) band in the range (1658-1593 cm^{-1} while the amide I ($\nu\text{C}=\text{O}$) appear as a strong bands in the range 1685-1616 cm^{-1} .

Furthermore, the new amides contain sulfa component in particular contain SO₂ moiety appears featured bands at the range 1388-1336 and in 1188-1138 cm^{-1} which assigned to asymmetrical and symmetrical stretching vibration, respectively of SO₂ (Al-Salami, 2009; Al-Salami *et al.*, 2014). Also the sulfa component possess another group which was C = N moiety of aromatic ring, thus, the band of C = N appear at the range (1329-1333 cm^{-1}). In addition, appearance of bands at the range 1776-1772 and at 1708-1714 cm^{-1} which attributed to C = O anhydride moiety.

HNMR spectra: The HNMR spectral data of new amides (F1-F7) have been summarized in Table 3. The results gave a great match to the expected composition of the structure of these new amides. The spectra of selected HNMR were shown in Fig. 5-7. From HNMR spectrum all compounds (F1-F7) displays a singlet signal at 10.2-10.36 ppm attributed to proton of NHC = O proton which indicates the formation of corresponding amides due to condensation of amino acids derivatives and sulfa drugs (Al-Salami, 2018). Also all synthetic amides except F5 exhibit a broad signal which can assigned to SO₂NH proton that affiliated to sulfa component (Ebrahimi *et al.*, 2013; Kumar and Rani, 2011). As we observe from HNMR the spectrum of compound F1 display a broad signal at 10.29 ppm which attributed to the two protons terminal amine group NH₂ which belong to components of sulfa drugs (Al-Salami, 2009, 2018). Also, the multiple signals that appear in the range (6.09-8.41 ppm) can be assigned to the aromatic protons

Table 2: FT-IR of synthetic amide (cm⁻¹, KBr disc) (vs: very strong, s : strong, m : medium, w: weak, br: broad)

Comp.	ν_{N-H}	ν_{C-H} aromatic	ν_{C-H} aliphatic asy-sym.	$\nu_{C=O}$ anhydride	$\nu_{C=O}$ amide-I	ν_{N-H} amide-II	ν_{C-N} sulfa	ν_{S-O} asym.	ν_{S-O} sym	ν_{S-N}
F1	3369 s	3059 w	2995 m 2929	1772 m 1714 s	1656 s	1649 w	1531 m	1382 s	1155 m	893 w
F2	3373 m	3059 w	2956 s 2933 s	1776 m 1712 m	1683 s	1647 w	1529 s	1386 m	1188 m	923 w
F3	3358 m	3062 m	2958 s 2933 s	1774 m 1712 s	1616 s	1593 w	1533 m	1388 s	1165 m	931 w
F4	3327 m	3111 w	2929 m 2852 m	1774 s 1712 s	1674 m	1658 w	1533 s	1386 s	1145 s	929 w
F5	3381 m	3061 w	2929 s 2854 m	1772 m 1714 s	1685 m	1656 m	1530 m	1386 m	1155 m	858 w
F6	3365 m	3057 w	2929 s 2854 m	1774 w 1712 m	1656 m	1631 m	1533 m	1386 s	1138 m	945 w
F7	3373 w	3035 w	2933 s 2854 m	1776 m 1708 s	1678 m	1658 m	1533 m	1336 s	1161 s	947 m

Table 3: ¹HNMR spectral data of synthetic amides (F1-F7)

Symbol of com.	Structural formula	Chemical shift δ (ppm)
F1		10.29 (s, 1H, NH-CO), 6.88-7.73 (m, 8H, Ar), 10.29 (br, 2H, NH ₂) _{Sulfa} , 5.16 (t, H, CH), 2.075 (dd, 2H, CH ₂), 1.65 (m, H, CH), 1.24 (d, 6H, CH ₃)
F2		10.36 (s, 1H, NH-CO), 6.10-7.90 (m, 8H, Ar), 11.0 (br, H, SO ₂ NH) _{Sulfa} , 2.07 (s, 3H, CH ₃) _{Sulfa} , 2.081 (s, 3H, CH ₃) _{sulfa} , 4.79 (t, H, CH), 2.079 (dd, 2H, CH ₂), 1.23 (m, H, CH), 0.90 (d, 6H, CH ₃)
F3		10.36 (s, 1H, NH-CO), 6.09-7.79 (m, 9H, Ar), 11.09 (br, H, SO ₂ NH) _{Sulfa} , 2.29 (s, 3H, CH ₃) _{Sulfa} , 4.79 (t, H, CH), 2.0759 (dd, 2H, CH ₂), 1.34 (m, H, CH), 0.92 (d, 6H, CH ₃)
F4		10.24 (s, 1H, NH-CO), 6.59-7.93 (m, 10H, Ar), 11.02 (br, H, SO ₂ NH) _{Sulfa} , 3.92 (t, H, CH), 2.0759 (dd, 2H, CH ₂), 1.62 (m, H, CH), 0.90 (d, 6H, CH ₃)
F5		10.34 (s, 1H, NH-CO), 6.18-7.76 (m, 8H, Ar), 1.69 (s, 3H, CH ₃) _{Sulfa} , 4.8(t, H, CH), 2.11 (dd, 2H, CH ₂), 1.45 (m, H, CH), 0.92 (d, 6H, CH ₃)
F6		10.30 (s, 1H, NH-CO), 6.71-8.09 (m, 12H, Ar), 11.02 (br, H, SO ₂ NH) _{Sulfa} , 4.66(t, H, CH), 2.071 (dd, 2H, CH ₂), 1.46 (m, H, CH), 0.961 (d, 6H, CH ₃)
F7		10.28 (s, 1H, NH-CO), 6.92-8.41 (m, 7H, Ar), 11.013 (br, H, SO ₂ NH) _{Sulfa} , 5.01 (t, H, CH), 2.08 (dd, 2H, CH ₂), 1.60 (m, H, CH), 0.99 (d, 6H, CH ₃)

of these new amides. Furthermore, all the synthesized compounds have been shown signals to CH₃, CH₂ and CH protons with different locations as shown in Table 3, the appearance and interpreted of these signals depend on the site of these components in the organic molecule. Also the signal of CH₃ (sulfa moiety) protons in compound s F2 and F3 where observed at 2,07 and 2.29 ppm (Ahmad *et al.*, 2015).

Antibacterial testing: *In vitro* antimicrobial activity of the compounds (F1-F7) have been screened against two, kinds of bacteria, Gram positive (*S.aurous*) and Gram negative (*A, Veronil*) as shown in Table 4, Fig. 8. DMSO is used as negative control and solvent. The antibiotic tetracycline 25 g L⁻¹ have been used as a positive

standard. The stock solution from each compound was prepared. Serial dilutions of experimental compounds were prepared in different conc. The 25, 50, 100, 150, 200, 250, 300 mg L⁻¹ in order to calculate the value of Minimum Inhibition Concentration (MIC). Antibacterial activities of each compound were estimated in Table 4. From the results of antibacterial screening of these amides, it has been observed that all the synthesized compounds have a variable antibacterial activity against the two kinds of microorganism, especially, against Gram-negative bacteria, than another type Gram-positive even in low concentration. On the other hand, we note an increase in the antimicrobial activity whenever the concentration of these amides increase. Also, from the current study and the results we observed that compound

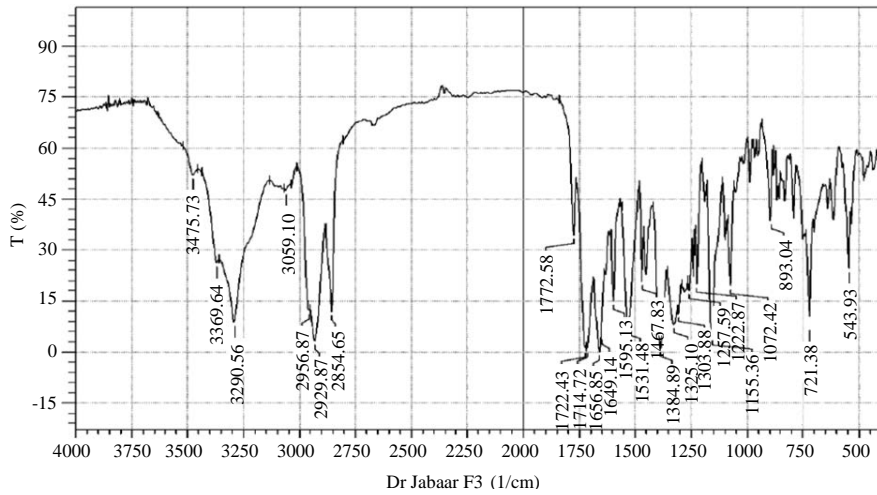


Fig. 2: IR spectrum of F1 compounds

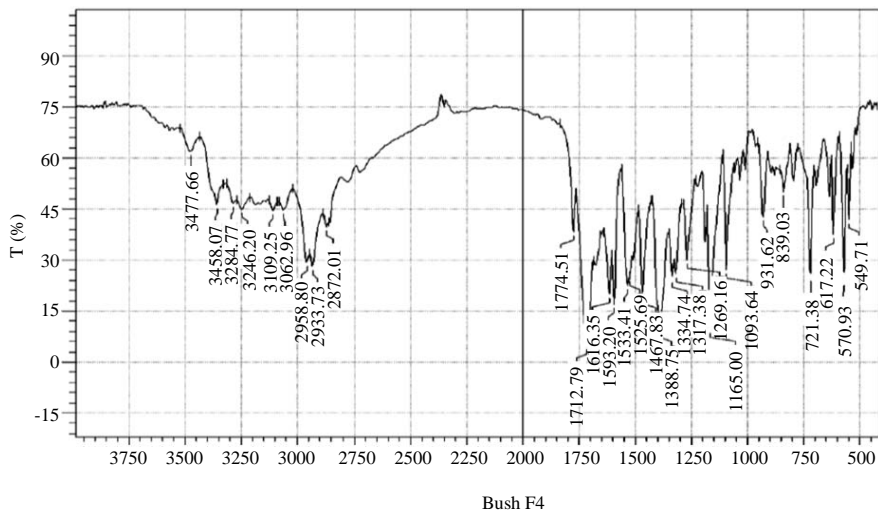


Fig. 3: IR spectrum of F3 compound

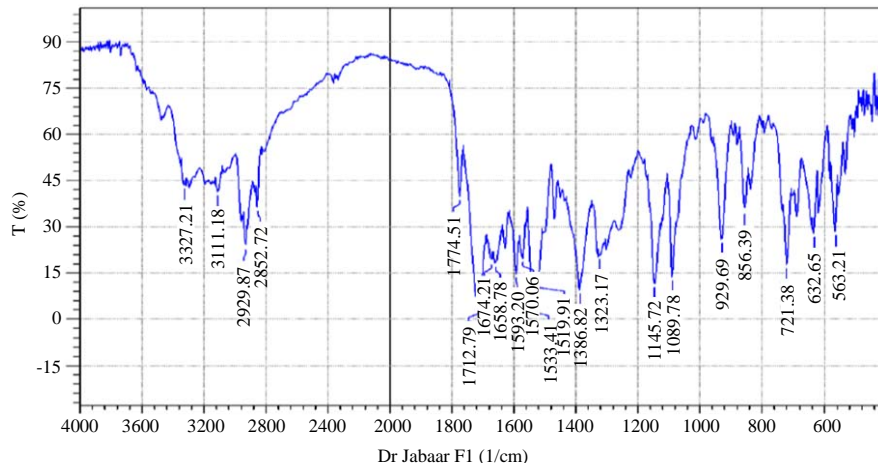


Fig. 4: IR spectrum of F4 compounds

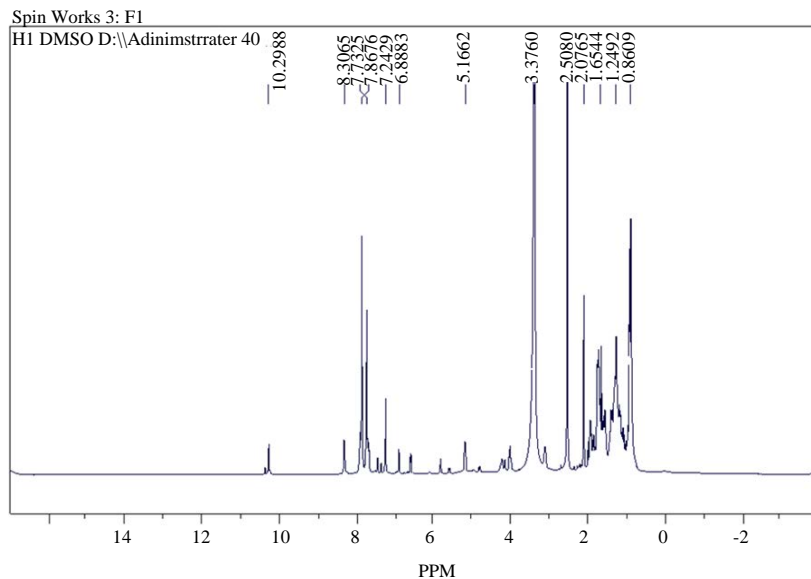


Fig. 5: ¹H NMR spectrum of F1 compound

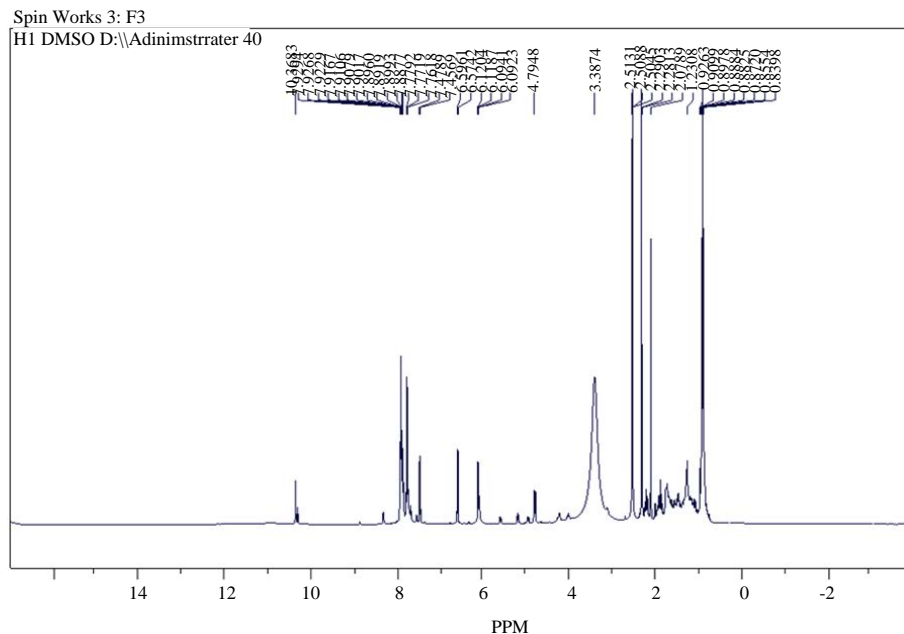


Fig. 6: ¹H NMR spectrum of F3 compound

F4 and F6 owned a better activity than another compounds against the two kinds of microorganisms. The observation of antimicrobial inspection indicate that these synthesized amides can affect on the cell walls of bacteria which consists of two layers, the peptidoglycan which is located outside of cytoplasmic membrane and the another layer which is lipid bilayer membrane which is characterized by the bacterial in general from the another kinds of organism. When compared with tow kinds (-ve) and (+v), the cell wall of gram-positive

bacteria is thick because the matrix substance in this wall contain polysaccharides, ptiptidoglycan which known as murein layer is the main component of this wall and lipotieichoic acid on the contrary the cell wall of Gram-negative bacteria is thin which contain a thin peptidoglycan layer (Demchick and Koch, 1996), so, these materials such as the synthesized compounds can penetrate the wall easily and smoothly thus, affecting the osmotic pressure (Yabe *et al.*, 2011). In addition, the wall membrane of these bacteria formulated with

Table 4: *In vitro* activity and Minimum Inhibitory Concentration (MIC) result of the synthesized compounds (F1-F7)

Compound	Dimeter of inhibition zone (mm) <i>Staphylococcus aureus</i>								MIC
	Concentration (mg L ⁻¹)								
	25	50	100	150	200	250	300		
F1	NI*	4	7	11	15	21	26	50	
F2	NI	NI	4	7	11	17	20	100	
F3	NI	NI	NI	5	8	13	19	150	
F4	4	6	11	16	20	24	39	25	
F5	NI	NI	NI	4	7	12	19	150	
F6	5	9	15	18	22	29	35	25	
F7	4	7	13	17	20	27	31	25	
Tetracycline	4	6	10	14	18	22	27	25	

Compound	Dimeter of inhibition zone (mm) <i>Aerompnas veronii</i>								MIC
	Concentration (mg L ⁻¹)								
	25	50	100	150	200	250	300		
F1	NI	NI	4	7	12	18	23	100	
F2	6	9	16	20	27	33	42	25	
F3	5	7	13	19	22	29	35	25	
F4	4	6	11	17	21	27	31	25	
F5	NI	7	12	18	25	29	35	50	
F6	5	8	15	21	27	35	40	25	
F7	NI	5	9	16	20	27	33	50	
Tetracycline	5	8	11	16	17	26	29	25	

NI*: No Inhibition

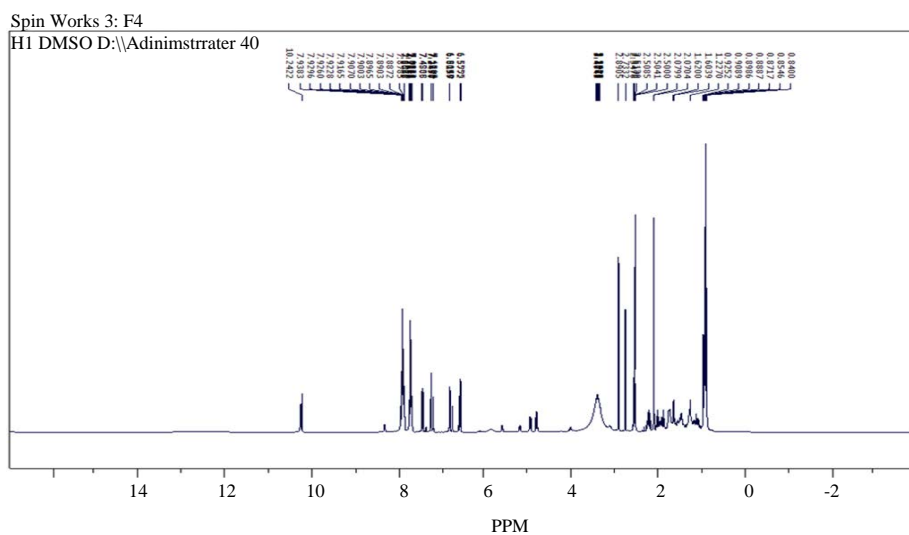


Fig. 7: ¹H NMR spectrum of F4 compound

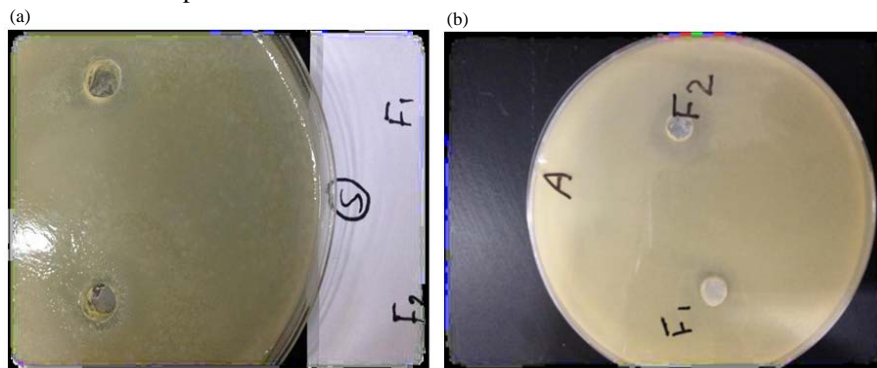


Fig. 8(a, b): The antibacterial activity of compounds F1 and F2

peptidolycans long chain of carbohydrates cross-linked by stumpy peptides and therefore, if any component abnormal which created or synthesized can inserted into these peptides will affect and the fobidding these bacteria from multiply and breakdown the formation of the cross-links thus, leading to shredding that wall (Yabe *et al.*, 2011).

CONCLUSION

In this study, emphasis was placed on the utilization of amino acid derivatives such as phthalyl DL-leucine and sulfa drugs to synthesized new amides by condensation reaction. This reaction can be done by nucleophilic acyl substitution reaction by using N, N-Dicyclohexylcarbodiimide (DCCI) as condensation reagent in order to accelerated the process of condensation. These new synthesized amides have also been characterized based on elemental analysis, FT-IR and HNMR spectra. The analysis confirmed the composition and structures of these newly amides. Also the antibacterial activity have been examined for these newly compounds by using two types of bacteria, negative gram stain (*Aermpans veronil*) and against positive gram (*Staphlococcus aureus*) by using different concentration of these compounds.

From the biological results it has been observed that these compounds show slightly active against Gram positive and have abetter activity toward the Gram negative depend on the instillation of a cell wall of microorganism. In fact these newly amides showed excellent effectiveness at high concentrations (300 mg L⁻¹) especially, against Gram-negative stain. Also the MIC was done by compare these newly amides with tetracycline as standard to demonstrate the capacity of these newly amide against some types of bacteria.

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