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A Thermotolerant *Acetobacter* Strain Isolated from Iranian Peach Suitable for Industrial Microbiology

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ABSTRACT

The aims of this research were investigating natural origins for producing new vinegar types as well as isolating and identifying new AAB strains that could be used in large scale industrial procedures and those could resist formal stresses such as alcohol and acetic acid concentrations, temperature fluctuations and other conditional alterations. We isolated and identified a native Acetobacter strain with high acetic acid productivity and tolerance against high ethanol concentrations from Iranian peach as a summer delicious fruit that is very susceptible to food spoilage and decay. We used selective and specific laboratorial culture media such as Standard GYC, Frateur and Carr media. Also we used a new industrial culture medium and a miniature fermentor with a new aeration system innovated by Pars Yeema Biotechnologists Co., Isfahan Science and Technology Town, Isfahan, Iran. The isolated strain was successfully cultivated in modified Carr media with 2.5 and 5% ethanol simultaneously in high temperatures, 34-40°C after 96 h of incubation period. We showed that the increase of ethanol concentration resulted in rising of strain sensitivity to high temperature. In conclusion we isolated and characterized a new Acetobacter strain from Iranian peach that could be considered as a potential strain for production of a new vinegar type, peach vinegar, with a delicious taste and advantageous nutritional value in food biotechnology and industrial microbiology.

Key words: Acetobacter, acetic acid bacteria, vinegar, Iranian peach, thermotolerant, ethanol resistance

INTRODUCTION

Vinegar is a precious food additive and complement as well as effective preservative against food spoilage that is produced by Acetic Acid Bacteria (AAB) and contains essential nutrients such as amino acids regarding its fruit source (Kocher et al., 2006). FDA (Food and Drug Administration, USA) has explained the vinegar as a 4% acetic acid solution that is synthesized from sweet or sugary substances through alcoholic fermentation. The neoclassical fermentations resulted in several vinegar types with different tastes, fragrances and nutritional values because of applying various acetic acid bacteria in vinegar making procedures. Currently the vinegar manufacturers are seeking for new types of vinegar using different AAB as their starter and traditional vinegar production has been improved using various natural substrates and fruits

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(Beheshti Maal and Shafiee, 2010; Du Toit and Lambrechts, 2002; Gullo and Giudici, 2008). The AAB are classified into genera Acetobacter, Gluconobacter, Fratueria, Acidomonas and Gluconacetobacter (Kadere et al., 2008; Holt et al., 1994; Madigan et al., 2008; Ruiz et al., 2000). Among AAB, Acetobacter strains are the major bacteria that are dealing with vinegar production industrially (Sokollek et al., 1998; Kadere et al., 2008). Acetobacteria are living on certain fruits and flowers and responsible for beer and wine spoilage as well as fruit decay (Silva et al., 2007). They are motile or nonmotile gram negative rods that their catalase and oxidase reactions are positive and negative respectively. Acetobacteria grow at temperatures of 25-30°C optimally and oxidize ethanol to acetic acid and sequentially to carbon dioxide and water as over oxidizers (Beheshti Maal and Shafiee, 2010; Gullo et al., 2009). Acetobacter strains have been isolated from several resources such as palm tree and palm wine (Faparusi, 1973; Ogbona, 1993), apple, Jamaican cherry, longan, mango, pineapple and rambutan (Moryadee and Pathom-Aree, 2008; Sossou et al., 2009), Iranian white-red cherry (Beheshti Maal and Shafiee, 2010), grape, date and coconut (Joyeux et al., 1984; Drydale and Fleet, 1985; Kadere et al., 2008). Several vinegar types have been synthesized using AAB and various precursors such as balsam (Giudici and Rinaldi, 2007; Gullo et al., 2009; Falcone and Giudici, 2008), sugarcane (Kocher et al., 2006) and rice (Nanda et al., 2001). The AAB strains specially those of Acetobacter and Gluconobacter sp. could produce various polysaccharides e.g., cellulose, dextran and levan that affect the quality of vinegar produced. Some mutated Acetobacter strains could release heterogeneous exopolysaccharides combined from glucose, mannose, rhamnose and glucuronic acid (Kadere et al., 2008). The goals of this research were investigating natural origins for producing new vinegar types as well as isolating and identifying new AAB strains that could be used in large scale industrial procedures and those could resist formal stresses such as alcohol and acetic acid concentrations, temperature fluctuations and other conditional alterations.

MATERIALS AND METHODS

The presented study was conducted since June 2007, at The Department of Microbiology, Islamic Azad University, Falavarjan Branch, The Department of Biology, University of Isfahan and The Laboratory of Biotechnology, Isfahan Science and Technology Town (ISTT), Isfahan, Iran.

Iranian peach extract preparation: The heterogeneous samples of both intact and spoilaged summer fruit, Iranian peach, were gathered from various fruit stores in Isfahan, Iran using sterile containers and incubated in a good ventilated cabinet at room temperature, 25-30°C, for 2 weeks. After rising sour smelling from incubated container that suggested the activity of AAB, the Iranian peach spoilages were pressed, squeezed, homogenized and passed through a sterile basket with recognized pore size. The cores of peaches were isolated and extracts were transferred to sterile 2 L bottles, capped and incubated at 30°C for 7 day anaerobically. For inhibiting bottle's explosion due to alcoholic fermentation, a few tiny punctures were made on top of the bottles for extra gas drainage.

Microbial culture media and instruments: The laboratorial culture media that we used were included GYC standard medium (yeast extract, 10 g L⁻¹; D-glucose, 50 g L⁻¹; CaCO₃, 30 g L⁻¹; agar, 25 g L⁻¹; distilled water, 1000 mL), Frateur medium (yeast extract, 10 g L⁻¹; CaCO₃, 20 g L⁻¹; ethanol, 20 g L⁻¹; agar, 20 g L⁻¹; distilled water, 1000 mL), Carr medium (yeast extract, 3%; agar, 2%; bromocresol green, 0.002%; ethanol, 2% (v/v); distilled water, 1000 mL) and modified Carr media with 3, 4, 5, 6, 7, 8, 9 and 10% ethanol concentrations. The industrial culture medium

(ethanol, 2%; acetic acid, 2%; yeast extract, 1%) was applied for initial enrichment and isolation of AAB from Iranian peach. All the chemicals were from the Merck. The main instruments that we used were incubator (Heraus, Germany), shaker incubator (Jahl, JSH 20L, Iran), autoclave (Iran Tolid, Iran), miniature glass fermentor (Pars Yeema Biotechnologists Co., ISTT, Isfahan, Iran), Spectrophotometer (Milton Roy, USA), high speed refrigerated centrifuge (Hitachi, 20PR, Japan), microscope (Nikon, Japan) and stereomicroscope (Wild, Germany).

AAB enrichment and isolation from Iranian peach: Fifty milliliters of homogenized Iranian peach extract after 7 days incubation at 30°C were collected and centrifuged at 10000 rpm for 20 min. The supernatants were discarded and the pellets were added to miniature glass fermentors containing of 400 mL industrial broth medium. The fermentors then were placed in a 30°C incubator while aeration was carrying out through an appropriate sparging system for 7 days. Each 24 h the acetic acid elevations were examined through titration assay. The containing of fermentors with acetic acid percentage more than 4% were cultured on AAB selective culture media using streak plate method and incubated at 30°C for 4 days.

Macroscopic, microscopic and biochemical properties of isolated strains: The macroscopic traits of isolated colonies from Iranian peach, morphological characteristics of colonies were investigated using stereomicroscopy. Microscopic and biochemical examinations of pure individual colonies were carried out using gram staining, catalase and oxidase reactions, ability of overoxidizing in Carr medium and ability of consuming calcium carbonate (transparenting around of colonies) in Frateur medium.

Investigation of strain's tolerance to various ethanol concentrations: The isolated strains of AAB from Iranian peach were cultured on modified Carr media with 3, 4, 5, 6, 7, 8, 9 and 10% ethanol concentrations using streak plate method and incubated at 30°C for 96 h.

Investigation of strain's resistance to high temperatures: The isolated strains of AAB from Iranian peach were cultivated on Carr medium and modified Carr media with 2.5 and 5% ethanol concentrations and incubated at high temperatures of 34, 36, 38 and 40°C for 96 h.

Acetic acid titration assay: In all industrial broth media, both before and after inoculation with isolated strains from Iranian peach, the titration assay of acetic acid percentage were took placed as follow: five milliliters of the broth medium were added to 20 mL of distilled water in a 250 mL flask and mixed thoroughly with 5 drops of phenol phetalein (phenol phetalein, 0.1 g; ethanol, 60 g; distilled water, 40 g) and then 0.5 normal sodium hydroxide (NaOH, 20 g L⁻¹; distilled water, 1000 mL) were added using 200 mL burette to mentioned mixture until appearance of pale pink color in the flask. The volume of consumed NaOH was measured and the acetic acid percentage in each medium was computed.

RESULTS

Incubation of industrial culture media inoculated with Iranian peach extracts at 30°C and for 4 days resulted in turbidity and acetic acid enhancement (data not shown). Streak plate method cultivation of turbid broth on Carr medium and Frateur medium resulted in emerging of pure, individual colonies that were similar to *Acetobacter* genus. The differential macroscopic, microscopic

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and biochemical examinations revealed that the isolated strain was related to AAB. The capability of strains in converting the blue color of Carr medium to yellow color and growth in 2% ethanol after 48 h suggested that they were related to *Gluconobacter* or *Acetobacter* sp. but color returning of Carr medium from yellow to blue after 96 h incubation at 30°C showed that the isolated bacterium was an overoxidizer strain and confirmed that related to *Acetobacter* sp. Growth in Frateur culture medium and transparenting the zones around the individual colonies after 96 h incubation at 30°C showed that the same strain has utilized calcium carbonate in Frateur medium, the property that is exclusively related to genus *Acetobacter*, so reconfirmed that the isolated strain was *Acetobacter*. A summary of main experiments and their results that led to identification of *Acetobacter* strain isolated from Iranian peach were shown in Table 1.

The cultivation of *Acetobacter* strain isolated from Iranian peach on modified Carr media with 3-10% ethanol concentrations at 30°C for 4 days showed that it could tolerate against high ethanol concentrations. Table 2-5 show the results of experiments accessed the strain tolerance in various ethanol concentrations on modified Carr media with 3, 4, 5, 6, 7, 8, 9 and 10% ethanol after 24-96 h incubation at 30°C. The growth rates of isolated *Acetobacter* strain in modified Carr media with 3, 4, 5, 6, 7, 8, 9 and 10% ethanol after 24 h incubation at 30°C were 4+, 2+, 1+, -, -, - and -, respectively (Table 2).

The isolated *Acetobacter* strain after 48 h incubation at 30°C in modified Carr media with 3, 4, 5, 6, 7, 8, 9 and 10% ethanol showed the growth rates of 4+, 3+, 2+, 1+, -, -, - and -, respectively as were shown in Table 3.

The same strain on modified Carr media with 3, 4, 5, 6, 7, 8, 9 and 10% ethanol at 30°C after 72 h indicated the growth rates of 4+, 3+, 3+, 2+, 1+, 1+, - and -, respectively (Table 4).

Table 1: Macroscopic, microscopic and biochemical tests of isolated Acetobacter strain from Iranian peach after 24-96 h incubation at 30°C

Culture media examinations		Carr medium agar	Frateur medium agar	
Macroscopic examinations	Colony	Blue-yellowish, needle-tip to small with	Pale gray-greenish, small	
	characterizations	<3 mm diameter, circular, convex and	colonies with ≤3 mm	
		regular shape with brownish center,	diameter, circular, convex and	
		vinegar smelling	regular shape, clearance zone	
			around the individual colonies	
Microscopic examinations	Gram reaction	Gram negative	Gram negative	
	Morphology	Short rods/coccobacilli	Short rods/coccobacilli	
	Arrangement	Mono, diplo and a few streptobacilli	Mono and diplobacilli	
Biochemical characterizations	Aerobic Growth	Positive	Positive	
	Anaerobic Growth	Negative	Negative	
	Catalase Reaction	Positive	Positive	
	Oxidase Reaction	Negative	Negative	
	Overoxidation capability	Positive	Negative	
	CaCO ₃ utilization	Negative	Positive	

Table 2: The tolerance of isolated *Acetobacter* strain from Iranian peach against various ethanol concentrations on modified Carr media at 30°C after 24 h

	Percentage							
Ethanol concentrations	3	4	5	6	7	8	9	10
Growth rates	4+	2+	1+	Negative	Negative	Negative	Negative	Negative
Acetic acid production	4+	2+	1+	Negative	Negative	Negative	Negative	Negative

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Table 3: The tolerance of isolated *Acetobacter* strain from Iranian peach against various ethanol concentrations on modified Carr media at 30°C after 48 h

	Percentage								
Ethanol concentrations	3	4	5	6	7	8	9	10	
Growth rates	4+	3+	2+	1+	Negative	Negative	Negative	Negative	
Acetic acid production	4+	3+	2+	1+	Negative	Negative	Negative	Negative	

Table 4: The tolerance of isolated *Acetobacter* strain from Iranian peach against various ethanol concentrations on modified Carr media at 30°C after 72 h

	Percent	tage						
Ethanol concentrations	3	4	5	6	7	8	9	10
Growth rates	4+	3+	3+	2+	1+	1+	Negative	Negative
Acetic acid production	4+	3+	3+	2+	1+	1+	Negative	Negative

Table 5: The tolerance of isolated *Acetobacter* strain from Iranian peach against various ethanol concentrations on modified Carr media at 30°C after 96 h

	Percentage							
Ethanol concentrations	3	4	5	6	7	8	9	10
Growth rates	4+	4+	4+	4+	4+	3+	2+	2+
Acetic acid production	4+	4+	4+	4+	4+	3+	2+	2+

Table 6: The tolerance of isolated Acetobacter strain from Iranian peach after 24-96 h incubation in 2.5% ethanol at high temperatures

Temperature (°C)	Growth rates after different incubation time (h)					
	24	48	72	96		
34	3+	4+	4+	4+		
36	-	1+	1+	1+		
38	2+	2+	2+	2+		
40	2+	2+	2+	2+		

The *Acetobacter* strain isolated from Iranian peach after incubation at 30°C for 96 h and on different modified Carr media with 3, 4, 5, 6, 7, 8, 9 and 10% ethanol showed the growth rates of 4+, 4+, 4+, 4+, 3+, 2+ and 2+, respectively as have been shown in Table 5.

The comparison of growth rates of isolated *Acetobacter* strain from Iranian peach in increasingly ethanol concentrations from 3 to 10% at 30°C and in different incubation times, 24-96 h, were shown in Fig. 1.

The tolerance examinations of *Acetobacter* strain isolated from Iranian peach against high temperatures such as 34, 36, 38 and 40°C indicated that this strain could grow on 2.5 and 5% ethanol concentrations in these high temperatures.

Table 6 shows the growth of isolated *Acetobacter* strain on modified Carr mediums with 2.5% ethanol concentrations at high temperatures of 34, 36, 38 and 40°C after 24-96 h incubation period.

The next tolerance examinations showed that the *Acetobacter* strain isolated from Iranian peach grew in modified Carr media with 5% ethanol concentrations after 24-96 h at high temperatures of 34, 36, 38 and 40°C as shown in Table 7.

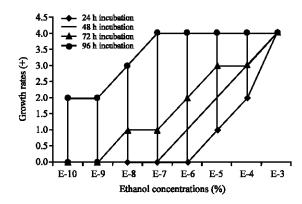


Fig. 1: The comparison of growth rates of *Acetobacter* strain isolated from Iranian peach and it's tolerance against various ethanol concentrations in different incubation periods, 24-96 h, at 30°C on modified Carr media with 3-10% ethanol

Table 7: The tolerance of isolated Acetobacter strain from Iranian peach after 24-96 h incubation in 5% ethanol at high temperatures

	Growth rates after different incubation time (h)					
Temperature (°C)	24	48	72	96		
34	2+	3+	3+	3+		
36	-	1+	1+	1+		
38	1+	1+	1+	1+		
40	1+	1+	2+	2+		

DISCUSSION

Sossou et al. (2009) have isolated Acetobacter sp. (ASVO3) from pineapple juice and produced vinegar after 23-25 days at 30°C. They studied their strain tolerance against glucose but reported that the concentration of ethanol had no effect on growth of AAB while we showed that the increase of ethanol concentration resulted in a greater lag phase in growth and rising of strain sensitivity to high temperatures. Beheshti Maal and Shafiee (2010) have reported the isolation and characterization of an Acetobacter strain from Iranian white red cherry. They have specified that the mentioned isolated strain had growth at 34-36°C in high ethanol, 5-9%, concentrations and after 4 day incubation duration that confirm our results but in this research we indicated the adaptation of isolated strain in higher temperatures. Moryadee and Pathom-Aree (2008) have isolated thermotolerant Acetobacter sp. from apple, Jamaican cherry, longan, mango, pineapple and rambutan using a culture medium containing of 4% ethanol. They reported an Acetobacter strain isolated from rambutan that produced the highest acetic acid amount at 37°C after 7 day incubation period that is not comparable to our strain. Iiha et al. (2000) have utilized honey for vinegar production using a mixed AAB and in room temperature but didn't report any suggestion for tolerance of their responsible strains against high temperature or ethanol concentrations. Kocher et al. (2006) have studied the production of vinegar by sugarcane using corn cobs, bagasse and wood shavings as immobilizers and Acetobacter aceti NRRL746. They showed that the fermentation time was reduced to 13 days by proposed recycling method that is much more of our production period. According to Kadere et al. (2008) the coconut toddy (mnazi) could be applied for production of vinegar using an Acetobacter strain that they isolated from coconut. They reported that the isolated strain had grew at 25, 30 and 40°C. Ndoye et al. (2007) have presented a new species of Acetobater, A. senegalensis, a thermotolerant Acetobacter that they isolated from mango (Mangifera indica L.). They have reported that their strain had exhibited growth at 35°C as an optimum temperature. While the previous studies have shown the isolation of Acetobacteria from different resources and with different characteristics, we isolated a thermotolerant Acetobacter strain from Iranian peach that we obtained from Isfahan, Iran. We showed that this strain could grow at high ethanol concentrations, 7-10%, at 30°C after 96 h on modified Carr media. We indicated that the isolated Acetobacter has grew on 2.5 and 5% ethanol concentrations at 34°C after 24 h but had no growth on the same ethanol concentrations at 36°C after 24 h, instead had growth after 48 h at 36°C. Also this strain had an acceptable growth on 2.5 and 5% ethanol concentrations at 38 and 40°C, unusual temperatures for AAB industrially, after 24 h. We suggested that the elevation of ethanol concentration resulted in extending lag phase of growth curve in isolated Acetobacter strain from Iranian peach but after 96 h in Carr medium with more than 6% ethanol concentration, the problem was overcome. These observations showed that the isolated Acetobacter stain from Iranian peach could rapidly adapt itself to extreme conditions such as high temperatures of 38-40°C as well as high concentrations of 5% ethanol simultaneously. We suggested this strain could be used in pilot-plant and large-scale fermentation of vinegar specially a new type of vinegar, peach vinegar. While, the peach is a very delicious summer fruit with nutritional advantages, but it is a very susceptible fruit against microbial spoilage. We showed that this strain could use mentioned spoilages as precursor for production of peach vinegar as a superior food additive. These properties, parallel and rapid adaptations to high temperatures and ethanol concentrations, could be considered as very dramatic characteristics for industrial strains that are qualified for large-scale commercial fermentations in vinegar industry and industrial microbiology. In conclusion, this is the first report of isolation and identification of an Acetobacter strain from the Iranian peach. The isolated thermotolerant Acetobacter strain could grow at 5% ethanol concentrations and 38-40°C that suggests a very suitable strain for large-scale vinegar fermentation in microbial biotechnology as well as a new amenable strain for bioremediation uses in environmental microbiology.

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