

ISSN 1996-3351

Asian Journal of
Biological
Sciences



Research Article

Absence of Typical Haversian System from the Compact Bone of Some Reptile and Bird Species

¹Yasser Ahmed, ²Mohammed Abdelsabour-Khalaf and ¹Fatma Khalil

¹Department of Histology, Faculty of Veterinary Medicine, South Valley University, Qena, Egypt

²Department of Anatomy and Embryology, Faculty of Veterinary Medicine, South Valley University, Qena, Egypt

Abstract

Background and Objective: Mammalian compact bone is composed mostly of Haversian system. Although there are many studies describing the typical Haversian system in mammals, there are few studies conducted on bones from non-mammalian species. The objective of the current study was to investigate the existence of the typical Haversian system in compact bones from reptiles and birds.

Materials and Methods: Femora were collected from geckos, Nile monitors, sparrows, ducks and geese. Samples were then, fixed in 10% paraformaldehyde and embedded in paraffin. Paraffin sections were stained with hematoxylin and eosin or Von Kossa and then examined using light microscopy. **Results:** The current study showed different histological structures of compact bones from studied species and an absence of the typical Haversian system seen in mammals. The compact bone of geckos was mostly avascular, while Nile monitor bone was highly vascular. Sparrow bone showed one side vascular and the other side avascular. The vascularity was represented by primary vascular canals surrounded by a few irregularly arranged osteocytes inside their lacunae forming primary osteons. Bone tissue from ducks and geese were similar and were composed mostly of dense Haversian tissue, while some areas had primary osteons. **Conclusion:** The compact bone microstructure in some reptile and bird species lacks the typical Haversian system described in mammals. This will be of a great importance in developing a better understanding of long bone anatomy and physiology in different species.

Key words: Haversian system, compact bone, geckos, Nile monitors, sparrows, ducks, geese, histology

Received: April 28, 2017

Accepted: May 30, 2017

Published: June 15, 2017

Citation: Yasser Ahmed, Mohammed Abdelsabour-Khalaf and Fatma Khalil, 2017. Absence of typical Haversian system from the compact bone of some reptile and bird species. Asian J. Biol. Sci., 10: 98-103.

Corresponding Author: Yasser Ahmed, Department of Histology, Faculty of Veterinary Medicine, South Valley University, Qena, Egypt
Tel: +20965211223/+201098782129 Fax: +20965211223

Copyright: © 2017 Yasser Ahmed *et al.* This is an open access article distributed under the terms of the creative commons attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

The shaft of compact bone in mammals is composed of similar units known as Haversian System (HS). An early study showed that although long bones may be used for the same purposes, their HS structure and arrangement are different from species to species, from bone to bone and even from side to side of the same bone¹. The typical mammalian HS is composed of vascular canals (Haversian canals) containing blood vessels and nerves and surrounded by different types of bone lamellae. Concentric lamellae are found around the Haversian canals, periosteal lamellae are close to periosteum, endosteal lamellae are close to the endosteum and circumferential lamellae between them². The Haversian canals with surrounding lamellae are commonly called secondary osteons (SO), which form on an earlier formed and smaller primary osteons (PO). The PO has no or a few lamellae smoothly merge with the surrounding bone tissue³. While bone lamellae of the SO are separated from surrounding bone tissue by a darkly stained cement line of bone matrix^{4,5}. According to its placement, the HS is subclassified into irregular, endosteal and dense types³. The irregular type consists of a few SO irregularly distributed through the bone tissue. The endosteal type has SO only close to the endosteal surface, while the dense type holds numerous SO distributed in all parts of the bone³.

The compact bone histological structure was recently used to differentiate between bone fragments from different animal species⁵⁻⁸ and from human and non-human sources⁹. Although there are large numbers of histological studies of compact bone of mammals²⁻¹³, there is very little literature on bone from reptiles and birds¹⁴. The current study aimed to investigate the existence of the typical HS in the compact bone of the femur from two non-mammalian groups; reptiles (geckos and Nile monitors) and birds (sparrows, ducks and geese).

MATERIALS AND METHODS

The current study undertaken in October, 2016 in the Histology laboratory, Faculty of Veterinary Medicine, South Valley University, Egypt.

Animals: The current study was carried out on apparently-healthy adult yellow-bellied Egyptian house geckos (*Hemidactylus flaviviridis*), Nile monitors (*Varanus niloticus*), Egyptian house sparrow (*Passer domesticus niloticus*), Muscovy ducks (*Cairina moschata*) and Egyptian geese (*Alopochen aegyptiacus*). Geckos (n = 4) were caught

alive from the office of the first author at the Faculty of Veterinary Medicine, South Valley University, Qena, Egypt. Nile monitor (n = 2) were collected from a lake in Qena City close to the Nile River. Sparrows (n = 4) were hunted with a rifle from the trees of Assiut City, Egypt. Ducks (n = 3) were obtained from a farm in Assiut city, Egypt. Geese (n = 3) were purchased from a village in Qena City, Egypt. Animals were euthanized and femora were rapidly dissected and immersed in 10% phosphate buffered formalin to avoid rapid postmortem changes. The femur was chosen because it is the longest and most loaded long bone¹⁰.

Sample processing: After fixation for 3 days at 4°C, samples were decalcified in 10% (v/v) analytical grade formic acid for 1-7 days depending on the species. During this time, the specimens were tested daily for proper decalcification. After, at least, 1 day for geckos, 3 days for sparrows, 5 days for ducks and geese and 7 days for Nile monitor, bone tissue became partially decalcified and ready for sectioning as a soft tissue. Decalcified bones were crosscut at the smallest breadth of their diaphysis (midshaft) to facilitate the easier study of compact bone histology. Cross but not longitudinal sections were taken because compact bone histology is different from one side to the other side of the same bone¹.

Histological examination: Decalcified specimens were embedded in analytical grade paraffin and processed for histological staining according to Ahmed *et al.*¹⁵. Paraffin sections (5 µm-thickness) were stained with hematoxylin and eosin (H and E) as a general stain or von Kossa's method for calcium detection in bone tissue according to Bancroft and Gamble¹¹. Stained sections were examined with a light microscope (Leica DMLS, Germany). Sections were photographed with Leica digital camera (Leica ICC50) using 4X, 10X, 40X and 100X objectives and pictures were captured in JPEG format.

RESULTS

No typical Haversian system, as described in mammals by other studies^{5,12}, could be seen in the compact bone from any of the examined species. Furthermore, the compact bones from the studied species were histologically different as explained below.

Histology of the compact bone from reptiles: Bone tissue from geckos appeared nearly avascular, except for 1 or 2 rounded vascular canals (Fig. 1a). The surrounding bone matrix contained a few number of osteocytes inside their

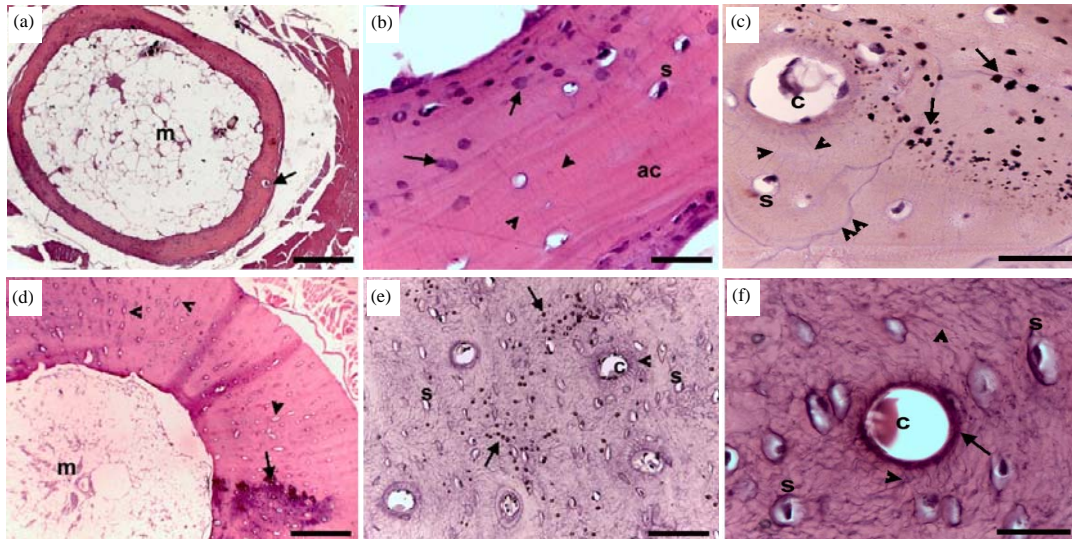


Fig. 1 (a-f): Histology of the compact bone of reptilian femur, Paraffin sections from the midshaft of the femur of geckos (a-c) and Nile monitors (d-f) stained by H and E (a, b, d, f) and von Kossa (c, e), (a) Bone marrow (m) and vascular canal (arrow), (b) Osteocytes in lacunae (s), canaliculi (arrowheads), dark-stained granules (arrows) and areas of acellular bone tissue (ac), (c) Vascular canal (c), osteocytes (s), canaliculi (arrowheads), cement line (double arrowhead) of the SO and von Kossa-positive granules (arrows), (d) Bone marrow (m), vascular canals (arrowheads) and patches of dark-stained areas (arrow), (e) Vascular canal (c), canaliculi (arrowhead), osteocytes (s) and von Kossa-positive granules (arrows) and (f) Vascular canal (c), acidophilic area (arrow), canaliculi (arrowheads) and osteocytes (s)
Scale bar = 250 μ m (a), 25 μ m (b), 25 μ m (c), 250 μ m (d), 62.5 μ m (e) and 25 μ m (f)

lacunae communicating with each other through canaliculi, which appeared perpendicular to the long axis of the bone. Large areas of avascular bone tissue were characterized the gecko' compact bone. Many large deeply stained granules were seen in the bone matrix (Fig. 1b). These granules stained positive for calcium with von Kossa (Fig. 1c). One or two SO were observed in some specimens and consisted of vascular canals surrounded by one or two layers of bone lamellae containing about 4-5 osteocytes embedded in lacunae and separated from the surrounding tissue by irregular cement lines (Fig. 1c). Unlike the gecko, compact bone from Nile monitors appeared highly vascular and cellular (Fig. 1d). The bone tissue contained many rounded, oval or elongated vascular canals surrounded by osteocytes with no demarcation lines separating them from the bone matrix and forming PO (Fig. 1e, f). One of the characteristic features was the presence of dense acidophilic areas around the vascular canals surrounded by a well-developed and long branched network of canaliculi (Fig. 1f). Similar to the bone matrix of geckos, many large deeply stained granules were observed and stained positive for calcium by von Kossa (Fig. 1e).

Histology of the compact bone from birds: Compact bone from sparrows showed a small number of rounded vascular

canals on one side, while the other side was avascular (Fig. 2a). The vascular canals were surrounded by a few osteocytes merged with the surrounding tissue forming PO (Fig. 2b). Two or three small irregular SO were surrounded by one or two layers of bone lamellae and separated from the surrounding tissues by cement lines (Fig. 2b). Canaliculi were not clear with the light microscopy. Von Kossa-positive granules were observed in some areas of the bone tissue, especially around vascular canals (Fig. 2c). In ducks and geese the compact bone was histologically of a similar structure. It was composed mostly of dense Haversian tissue morphologically different from typical HS of mammals. The Haversian tissue in ducks and geese had a large number of small SO clustered close to each other through the bone matrix (Fig. 2d). The SO were rounded with rounded or elliptical vascular canals surrounded by 1-3 layers of bone lamellae with a small number of osteocytes inside their lacunae and were separated from bone tissue by relatively thick and darkly stained areas (Fig. 2e). These areas stained positive by von Kossa (Fig. 2f). Transfers canals were seen in some areas (Fig. 2f). Many PO were seen and some appeared to have two vascular canals (Fig. 2g). The PO had rounded central canals surrounded by irregularly distributed osteocytes and a network of thin canaliculi (Fig. 2h). Close to the

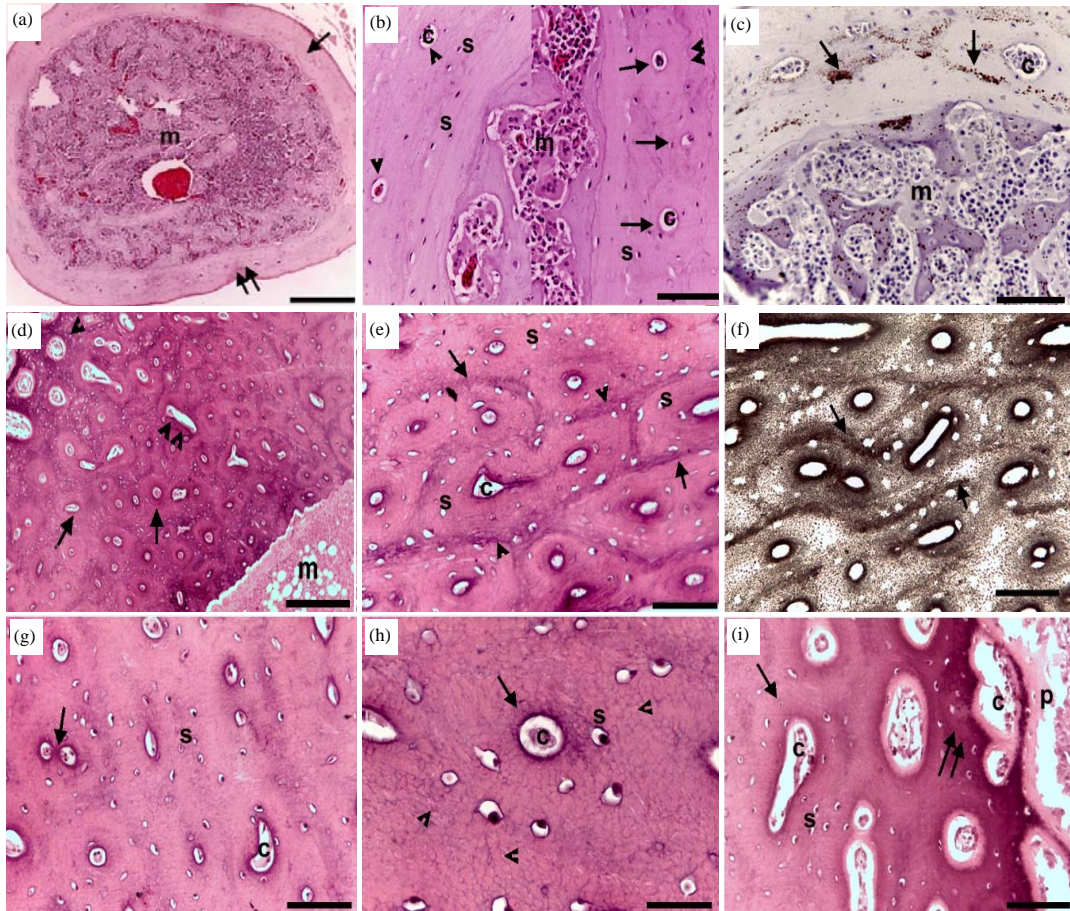


Fig. 2(a-i): Histology of the compact bone of avian femur, Paraffin sections from the midshaft of the femur of sparrows (a-c), ducks (d, g, h) and geese (e, f, i) stained by H and E (a, b, d, e, g, h, i) and von Kossa (c, f), (a) Bone marrow (m), avascular side (arrow) and vascular side (double arrows) of bone tissue, (b) Bone marrow (m), SO (arrows), PO (arrowheads), vascular canals (c), cement line (double arrowheads) and osteocytes inside lacunae (s), (c) Bone marrow (m), vascular canals (c), von Kossa-positive granules (arrows), (d) Bone marrow (m), SO (arrows), SO close to periosteum (arrowhead) and transverse canals (double arrowheads), (e) SO (arrows), cement lines (arrowheads), vascular canals (c) and osteocytes inside lacunae (s), (f) Von Kossa-positive cement line (arrows), (g) Two vascular canals (arrow) of the same PO and osteocytes (s), (h) PO (arrow), vascular canal (c), canaliculi (arrowheads) and osteocytes (s) inside lacunae and (i) SO (arrow) close to the periosteum (p), vascular canals (c), osteocytes inside lacunae (s) and dark-stained bone tissue (double arrows)

Scale bars = 250 μm (a), 62.5 μm (b), 62.5 μm (c), 250 μm (d), 62.5 μm (e), 62.5 μm (f), 250 μm (g), 25 μm (h) and 25 μm (i)

periosteum, most osteons were of the secondary type and showed comparatively large and irregular vascular canals surrounded by a darkly stained bone tissue with many osteocytes (Fig. 2i).

DISCUSSION

In the current study, compact bone from reptilian and avian species were examined with the aim of determining the presence of typical HS as previously described in mammalian

compact bone^{5,8,12,16}. No typical HS could be found in any bone specimen. Bone from small (geckos) and large (Nile monitors) reptiles showed different histology. In geckos, the compact bone was mostly avascular except for PO. The PO appeared as one or two vascular canals surrounded by a small number of osteocytes and only one SO could occasionally be seen in some sections. Similarly, in the femur of a Turkish lizard (*Stellagama stellio*), only two PO were seen and incorrectly described as SO¹⁴. Nile monitor bone was highly vascular and showed many rounded, oval or elongated vascular canals

surrounded by randomly distributed osteocytes forming PO. A similar observation was described early in the femurs of alligators¹. The difference in vascularity of the bone tissue from both reptile species is related to the bone size and cortical bone growth rate¹⁷. The PO seen in the examined species could remain during the whole animal's life and never been replaced by De Buffrenil *et al.*¹⁷ as the remodeling in such species is a physiologically delayed process¹⁸. Avian compact bone showed histological variations in sparrows versus ducks and geese. The bone tissue from the sparrow was vascular on one side and avascular on the other side. Such variation in vascularity of the same bone may be related to the mechanical stress difference applied on different sides of the bone. Only two or three small irregular SO could be observed in some sections. The low vascularity and SO appearance is related to the low-bone remodeling of birds such as sparrows. On the other hand, ducks and geese compact bone tissue was similar and consisted of dense Haversian tissue of many rounded clusters of SO separated from each other by thick dark areas. In another study, it was reported that compact bone from *Galliformes* had many small osteons with small number of bone lamellae⁸. The SO close to the periosteum had large irregular vascular canals and denser osteocytes. The PO was distributed to many areas and 2 vascular canals could be seen in the same PO. The morphological variations seen in sparrows versus ducks and geese in relation to osteonal morphology and density could be explained by the fact that these species apply biomechanical stress to the bone differently.

The histological variations seen in the compact bone of studied species may be related to the mechanical stress applied to long bones and position of the bones in the body and all could be closely correlated with the different lifestyles of these animals¹⁰. Even in mammals, there are variations in osteonal density according to the animal size and then mechanical load applied to bone tissue. For example, unlike large mammals, rat has very few SO and short osteonal canals¹³. Furthermore, the wild sheep long bone has a higher number and diameter of SO than domestic sheep¹⁰.

To the best of our knowledge, this is the first histological study that provides a descriptive comparative histology of compact bone from non-mammalian species (reptiles and birds).

CONCLUSION AND FUTURE RECOMMENDATION

It can be concluded that the typical HS described in mammalian compact bone is absent from the compact bone of some reptiles and birds. Furthermore, the histological structure of compact bone in the studied animals was widely different. Thus, the histological analysis of the long bones

structure may be functionally correlated to the habits of animals, the muscular stress applied to long bones and the relative position of the bone to the animal body.

A future study will focus on the histological differences between compact bone from different animal species to investigate the possibility of using the structure of compact bone to differentiate between different species, which will be of a great importance to veterinary forensic medicine.

SIGNIFICANCE STATEMENT

This study discovered the absence of the typical HS in the compact bone of some reptilian and avian species. This is of significant importance to veterinary forensic medicine. This study will help the researchers to uncover the critical areas of long bone morphology in different species that many researchers were not previously able to explore. Thus, a new theory on long bone development may be developed.

ACKNOWLEDGMENTS

The authors would like to thank Dr. Brad Morrison, Science Department at the Toronto District School Board, Canada and Prof. Antar Abdallah, Professor of English education at Taibah University, Saudi Arabia, for editing of the manuscript. As well, thank Dr Fatma Abass and Mrs. Fatma Abdel Hakim, Department of Anatomy, Faculty of Veterinary Medicine, South Valley University, Qena, Egypt for kindly providing the goose and duck specimens.

REFERENCES

1. Foote, J.S., 1911. The comparative histology of femoral bones. *Trans. Am. Microscop. Soc.*, 30: 87-140.
2. Cvetkovic, V.J., S.J. Najman, J.S. Rajkovic, A. Zabar, P.J. Vasiljevic, L.B. Djordjevic and M.D. Trajanovic, 2013. A comparison of the microarchitecture of lower limb long bones between some animal models and humans: A review. *Vet. Med.*, 58: 339-351.
3. Hillier, M.L. and L.S. Bell, 2007. Differentiating human bone from animal bone: A review of histological methods. *J. Forensic Sci.*, 52: 249-263.
4. Martiniakova, M., B. Grosskopf, R. Omelka, K. Dammers, M. Vondrakova and M. Bauerova, 2007. Histological study of compact bone tissue in some mammals: A method for species determination. *Int. J. Osteoarchaeol.*, 17: 82-90.
5. Zedda, M., G. Lepore, P. Manca, V. Chisu and V. Farina, 2008. Comparative bone histology of adult horses (*Equus caballus*) and cows (*Bos taurus*). *Anatomia Histol. Embryol.*, 37: 442-445.

6. Martiniakova, M., B. Grosskopf, R. Omelka, M. Vondrakova and M. Bauerova, 2006. Differences among species in compact bone tissue microstructure of mammalian skeleton: Use of a discriminant function analysis for species identification. *J. Forensic Sci.*, 51: 1235-1239.
7. Martiniakova, M., B. Grosskopf, M. Vondrakova, R. Omelka and M. Fabis, 2006. Differences in femoral compact bone tissue microscopic structure between adult cows (*Bos taurus*) and pigs (*Sus scrofa domestica*). *Anatomia Histol. Embryol.*, 35: 167-170.
8. Castrogiovanni, P., R. Imbesi, M. Fisichella and V. Mazzone, 2011. Osteonic organization of limb bones in mammals, including humans and birds: A preliminary study. *Italian J. Anat. Embryol.*, 116: 30-37.
9. Dominguez, V.M. and C.M. Crowder, 2012. The utility of osteon shape and circularity for differentiating human and non human Haversian bone. *Am. J. Phys. Anthropol.*, 149: 84-91.
10. Giua, S., V. Farina, A. Cacchioli, F. Ravanetti, M. Carcupino, M.M. Novas and M. Zedda, 2014. Comparative histology of the femur between mouflon (*Ovis aries musimon*) and sheep (*Ovis aries aries*). *J. Biol. Res.*, 87: 74-77.
11. Bancroft, J.D. and M. Gamble, 2008. *Theory and Practice of Histological Techniques*. 6th Edn., Elsevier Health Sciences, Philadelphia, PA., ISBN-13: 9780443102790, Pages: 725.
12. Zedda, M., G. Lepore, G.P. Biggio, S. Gadau, E. Mura and V. Farina, 2015. Morphology, morphometry and spatial distribution of secondary osteons in equine femur. *Anat. Histol. Embryol.*, 44: 328-332.
13. Martiniakova, M., R. Omelka, B. Grosskopf, Z. Mokosova and R. Toman, 2009. Histological analysis of compact bone tissue in adult laboratory rats. *Slovak J. Anim. Sci.*, 42: 56-59.
14. Kumas, M. and D. Ayaz, 2015. An observation of haversian systems in *Stellagama stellio* (Linnaeus, 1758)(Squamata: Sauria: Agamidae) in Barla/Isparta, Turkey. *Turk. J. Zool.*, 39: 1160-1161.
15. Ahmed, Y.A., S.A. Soliman and E.A. Abdel-Hafez, 2013. Ossification of the femur and tibia of the post-hatching Japanese quail. *Pak. J. Biol. Sci.*, 16: 859-864.
16. Gudea, A. and A. Stefan, 2015. Histomorphometric and fractal analysis of femoral, tibial, and metatarsal compact bone samples in sheep (*Ovis aries*), goat (*Capra hircus*) and roe deer (*Capreolus capreolus*). *Turk. J. Vet. Anim. Sci.*, 39: 528-536.
17. De Buffrenil, V., A. Houssaye and W. Bohme, 2008. Bone vascular supply in monitor lizards (Squamata: Varanidae): Influence of size, growth and phylogeny. *J. Morphol.*, 269: 533-543.
18. Smirina, E.M. and N.B. Ananjeva, 2007. Growth layers in bones and acrodont teeth of the agamid lizard *Laudakia stoliczkana* (Blanford, 1875)(Agamidae, Sauria). *Amphibia-Reptilia*, 28: 193-204.