

ISSN 1996-3351

Asian Journal of
Biological
Sciences



Research Article

Western Algerian Propolis Alcohol Extract Effects Against Disruption of Testes Function Induced by Cadmium-sulfate in Male Rats

¹Abdelkrim Berroukche, ¹Imen Denai, ²Mustapha Brahmi, ¹Hafsa Dellaoui, ¹Wassila Lansari and ¹Imen Zerarki

¹Research Laboratory of Water Resources and Environment, Department of Biology, Faculty of Science, Dr. Tahar-Moulay University, Saida 20000, Algeria

²Laboratory of Biotoxicology, Pharmacognosy and Biological Valorization of Plants, Department of Biology, Faculty of Science, Dr. Tahar-Moulay University, Saida 20000, Algeria

Abstract

Background and Objective: Propolis, a natural product derived from plant resins collected by honeybees has been used for thousands of years in traditional medicine all over the world. The composition of the propolis depends upon the vegetation of the area from where it was collected and on the bee species. This study aimed to assess the preventive effects of Algerian propolis against reproductive toxicity of Cadmium Sulfate (CdSO_4) in male rats. **Materials and Methods:** Animals were divided into four groups; group 1 was a control, group 2 received 15 mg CdSO_4 kg^{-1} b.wt., for consecutive 35 days, Group 3: exposed to CdSO_4 then treated with 200 mg propolis kg^{-1} , Group 4: treated with propolis then was administered CdSO_4 in the same conditions. **Results:** The results showed that CdSO_4 caused a decrease in body weight gain, testes weight and testosterone level. In the CdSO_4 -treated group, histopathologic examinations revealed apparent alterations in the testes, where it induced marked lesions in seminiferous tubules. Group 4, treated with propolis showed an increased plasma testosterone. **Conclusion:** Through these findings, propolis antagonized the harmful effects of CdSO_4 . This was proved histopathologically by the great improvement in testes. Propolis could be effective in the prevention against the reproductive toxicity of CdSO_4 .

Key words: Propolis, cadmium sulfate, testes, reproductive toxicity, male rats

Citation: Abdelkrim Berroukche, Imen Denai, Mustapha Brahmi, Hafsa Dellaoui, Wassila Lansari and Imen Zerarki, 2019. Western Algerian propolis alcohol extract effects against disruption of testes function induced by cadmium-sulfate in male rats. Asian J. Biol. Sci., 12: 24-30.

Corresponding Author: Abdelkrim Berroukche, Research Laboratory of Water Resources and Environment, Department of Biology, Faculty of Science, Dr. Tahar-Moulay University, Saida 20000, Algeria Tel: 00213555972162

Copyright: © 2019 Abdelkrim Berroukche *et al.* This is an open access article distributed under the terms of the creative commons attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Cadmium (Cd), toxic heavy metal at low dose, is widely present in soil and ecosystems with half-life about 15 years¹. Cd is used in industry to produce pesticides and batteries, also found in food (mollusks, crustacean and cocoa powder) and in tobacco smoke². About 25000 t of Cd was released every year in environment³. According to the International Agency of Research on Cancer (IARC), Cd is classified as a human carcinogen. Cd toxicity promotes the tumorigenesis through cell oxidative stress and inhibition of repair of oxidative DNA damage⁴. Cd toxicity impeded apoptosis and impelled cell proliferation leading to phenotype malignancy⁵. Studies suggested that Cd toxicity induced reactive oxygenated species (ROS) or free radical synthesis and consequently increased redox disruption⁶. Following an intestinal absorption, Cd bind to metalloproteins (MPP) and accumulate in different tissues such as lungs, liver, kidney, testis and prostate⁷. Cd disturb mitochondrial oxidative phosphorylation and inhibit cell respiration. Chronic occupational exposures to Cd reduce the fertility of workers^{7,8}. Cilenk *et al.*⁸ showed, in animals exposed to CdCl₂ at 1 mg kg⁻¹, significant decrease in sperm concentration, sperm motility, normal and live sperm⁸. The treatment of Cd toxicity is only symptomatic.

Propolis is natural resinous hive product. It is synthesized by honeybees, mixing their own waxes and salivated secretions with resins collected from the cracks in the bark of trees and leaf buds⁹. Propolis is consisted of 50% resin and balsam, 30% wax, 10% essential oils, 5% pollen and 5% other substances including organic compounds and minerals such as phenolic acids, flavonoids, terpenes, β -steroids, alcohols and sesquiterpenes¹⁰. Propolis, used in folk medicine, has biological activities like anti-microbial, anti-tumor and anti-oxidant properties¹⁰. Recent studies suggested that caffeic acid phenethyl ester (CAPE), bioactive compound found in propolis has anticancer properties¹¹. This molecule did not neutralize the prostate cancer but it slows its progression. Signaling transducer proteins (70S6 protein kinase and Akt), intermediates in steps of PI3K/mTOR signaling pathway, promote cell proliferation and angiogenesis¹². Propolis, through its magical molecule CAPE, inhibit Akt and 70S6 protein kinase activities and impelled cell death or apoptosis¹³. The present study aimed to assess the reproductive toxicity of Cadmium sulfate in adult male rats. Also, to evaluate the preventive effects of western Algerian propolis against the eventual testis dysfunction induced by Cadmium sulfate.

MATERIALS AND METHODS

Obtaining of propolis: Propolis samples were harvested by farmers at Rebahia area, located in the Saida region (western Algeria). The most abundant plant, as source of propolis, in this region was *Pinus halepensis*. Then propolis was purchased at local market, mainly at store specializing in sales of bee products.

Preparation of propolis alcohol extract: This experiment step was conducted according to Franchi's protocol¹⁴. Amount of 50 g of the propolis was cut into small pieces and extracted with 600 mL of 80% ethanol at 60°C for 30 min. Mixture was centrifuged and the supernatant was evaporated at 40°C. Dried residue was kept at 4°C. Aqueous suspension of propolis was prepared and orally administered to the animals for 35 days in a dose of 200 mg kg⁻¹ b.wt.¹⁵.

Experimental design: Twenty male adult rats (average weight 180-200 g) were used in the experiments. Animals were obtained from Pasteur Institute, Algiers, Algeria. The scientific committee of Research Laboratory of Water Resources and Environment, Faculty of Science, University of Saida, Algeria, approved the design of the experiments and the protocol conforms to the guidelines of the National Institutes of Health (NIH). Animals were caged in groups and given feed and water *ad libitum*. After one week of acclimation, animals were divided into four equal groups. The toxic dose of Cd used in the experiments was 15 mg kg⁻¹ b.wt.¹⁶:

Group 1: Normal animals used as controls

Group 2: Experiment control animals orally administrated with CdSO₄ (15 mg kg⁻¹ day⁻¹)

Group 3: Animals, initially exposed to CdSO₄ were orally treated with propolis extract (200 mg kg⁻¹ day⁻¹)

Group 4: Animals, initially treated with propolis were orally administered with CdSO₄ in the same experiment conditions

Biochemical study: During the experiment period, different blood biochemical parameters were determined. Serum cadmium, prostate specific antigen (PSA) and testosterone were measured using mini-VIDAS automate analyzer (Bio-Merieux, France). The enzyme-linked fluorescent assay (ELFA method) was used to assess serum PSA and testosterone levels. The activity of Glutathione S-transferase (GST) was measured according to Habig's

method¹⁷. The absorbance was measured at 310 nm using UV-spectrophotometer. The enzyme catalase (CAT) activity was measured spectrophotometrically at 240 nm¹⁸.

The blood samples were centrifuged at 2500 rpm for 10-15 min and the sera isolated were used for estimation of the serum glucose, triglycerides, total cholesterol, creatinine and urea levels. It was added to this study, a measurement of the blood count formula (BCF) that determine the count of all blood cells through controller Coulter STKS®.

Histopathological study: Testis were removed and kept in 10% formol saline for 24 h, dehydrated in ethanol and embedded in paraffin. Sections were cut at 5 micron thickness, mounted on slides and stained with hematoxylin and eosin.

Statistical analysis: Data were expressed as Mean \pm SD, with a value of $p < 0.05$ or $p < 0.01$ considered statistically significant. Statistical evaluation was performed by one way analysis of variance (ANOVA) followed by the Tukey's t-test for multiple comparisons. All analysis were made with the statistical software Sigma-Plot (version 11.0).

RESULTS

Results showed significant ($p < 0.01$) decrease in body weight and testis weight in animals administrated with CdSO₄ at the dose 15 mg kg⁻¹ (167.2 \pm 4.4 and 2.28 \pm 0.1 g, respectively) compared to controls (223.1 \pm 9.8 and 2.82 \pm 0.56 g, respectively) and animals treated with propolis at the dose 200 mg kg⁻¹; group 3 (190.3 \pm 2.8 and 2.64 \pm 2.01 g, respectively) and group 4 (176.6 \pm 2.7 and 2.4 \pm 0.96 g, respectively) (Table 1). Whereas experiments of this present

study indicated significant increase in blood glucose (1.83 \pm 0.1 g L⁻¹), lipid profile (Triglycerides: 1.76 \pm 0.1 g L⁻¹ and T-cholesterol: 4.11 \pm 0.18 g L⁻¹), kidney parameters (Urea: 0.84 \pm 0.07 g L⁻¹ and creatinine: 62.89 \pm 9.36 mg L⁻¹), blood PSA (9.15 \pm 3.62 ng mL⁻¹), testosterone (0.95 \pm 0.11 ng mL⁻¹) and serum cadmium (8.80 \pm 0.89 μ g L⁻¹) in animals treated with cadmium (group 2) compared to controls (1.03 \pm 0.11, 0.96 \pm 0.05, 0.74 \pm 0.06 and 0.17 \pm 0.02 g L⁻¹, 10.84 \pm 1.21 mg L⁻¹, 2.27 \pm 0.28 and 1.31 \pm 0.24 ng mL⁻¹, 0.03 \pm 0.001 μ g L⁻¹, respectively) and animals orally administrated with propolis as follow; group 3 (1.37 \pm 0.1, 1.37 \pm 0.04, 2.26 \pm 0.22, 0.55 \pm 0.06 g L⁻¹, 31.65 \pm 6.00 mg L⁻¹, 4.69 \pm 0.48, 1.06 \pm 0.10 ng mL⁻¹ and 1.40 \pm 0.90 μ g L⁻¹, respectively) and group 4 (1.36 \pm 0.06, 1.27 \pm 0.04, 1.62 \pm 0.09, 0.40 \pm 0.03 g L⁻¹, 19.44 \pm 2.14 mg L⁻¹, 3.94 \pm 0.24, 1.17 \pm 0.11 ng mL⁻¹ and 1.23 \pm 0.02 μ g L⁻¹, respectively) in the same experimental conditions such as (Table 1). Hematological results displayed low red cells and hemoglobin in animals administrated with cadmium (2.7 \pm 0.3 $\times 10^6$ IU and 5.1 \pm 0.8 g dL⁻¹, respectively) compared to controls (6.7 \pm 0.4 $\times 10^6$ IU and 15.6 \pm 1.8 g dL⁻¹, respectively) and animals treated with propolis in group 3 (4.8 \pm 2.2 $\times 10^6$ IU and 8.1 \pm 0.8 g dL⁻¹) and group 4 (5.2 \pm 0.0 $\times 10^6$ IU and 9.9 \pm 0.7 g dL⁻¹) (Table 1).

Treatment with CdSO₄ induced significant ($p < 0.01$) decrease in the activities of testes catalase (CAT) and glutathione S-transferase (GST) and reduced glutathione (GSH) compared to control animals. While, animals treated with propolis showed significant increase in CAT, GST and GSH ($p < 0.01$) (Table 1). In groups 3 and 4, propolis was able of recovering the activities of CAT and GST and the level of GSH compared to controls (Table 1).

Table 1: Variation of biochemical parameters in rats treated with CdSO₄ and propolis

Variables	Group 1 (controls)	Group 2 (CdSO ₄)	Group 3 (CdSO ₄ /propolis)	Group 4 (propolis/CdSO ₄)
Body weight (g)	223.10 \pm 9.8	167.20 \pm 4.4	190.30 \pm 2.8	176.60 \pm 2.7
Gain body weight (g day ⁻¹)	1.09	-0.48*	0.17*	0.30*
Testis weight (g)	2.82 \pm 0.56	2.28 \pm 0.1	2.64 \pm 2.01	2.40 \pm 0.96
Blood glucose (g L ⁻¹)	1.03 \pm 0.11	1.83 \pm 0.1*	1.37 \pm 0.1*	1.36 \pm 0.06*
Triglyceride (g L ⁻¹)	0.96 \pm 0.05	1.76 \pm 0.1*	1.37 \pm 0.04*	1.27 \pm 0.04*
T-cholesterol (g L ⁻¹)	0.74 \pm 0.06	4.11 \pm 0.18	2.26 \pm 0.22	1.62 \pm 0.09
Urea (g L ⁻¹)	0.17 \pm 0.02	0.84 \pm 0.07	0.55 \pm 0.06	0.40 \pm 0.03
Creatinine (mg L ⁻¹)	10.84 \pm 1.21	62.89 \pm 9.36	31.65 \pm 6.00	19.44 \pm 2.14
Red cells ($\times 10^6$ IU)	6.70 \pm 0.4	2.70 \pm 0.3	4.80 \pm 2.2	5.20 \pm 0.0
Hemoglobin (g dL ⁻¹)	15.60 \pm 1.8	5.10 \pm 0.8	8.10 \pm 0.8	9.90 \pm 0.7
PSA (ng mL ⁻¹)	2.27 \pm 0.28	9.15 \pm 3.62*	4.69 \pm 0.48*	3.94 \pm 0.24*
Testosterone (ng mL ⁻¹)	1.31 \pm 0.24	0.95 \pm 0.11*	1.06 \pm 0.10	1.17 \pm 0.11*
Cadmium (μ g L ⁻¹)	0.03 \pm 0.001	8.80 \pm 0.89*	1.40 \pm 0.90*	1.23 \pm 0.02*
CAT (mol h ⁻¹ g ⁻¹ tissue)	6.50 \pm 1.2	3.20 \pm 0.60	5.70 \pm 0.8	8.12 \pm 0.92
GST (μ mol min ⁻¹ g ⁻¹ tissue)	1.09 \pm 0.10	0.60 \pm 0.15	0.97 \pm 0.12	1.46 \pm 0.30
GSH (mM g ⁻¹ tissue)	5.80 \pm 0.7	4.10 \pm 0.7	5.62 \pm 1.9	8.13 \pm 0.9

*Results significantly different ($p < 0.01$), T-cholesterol: Total cholesterol, CAT: Catalase, GST: Glutathione S-transferase, GSH: Reduced glutathione

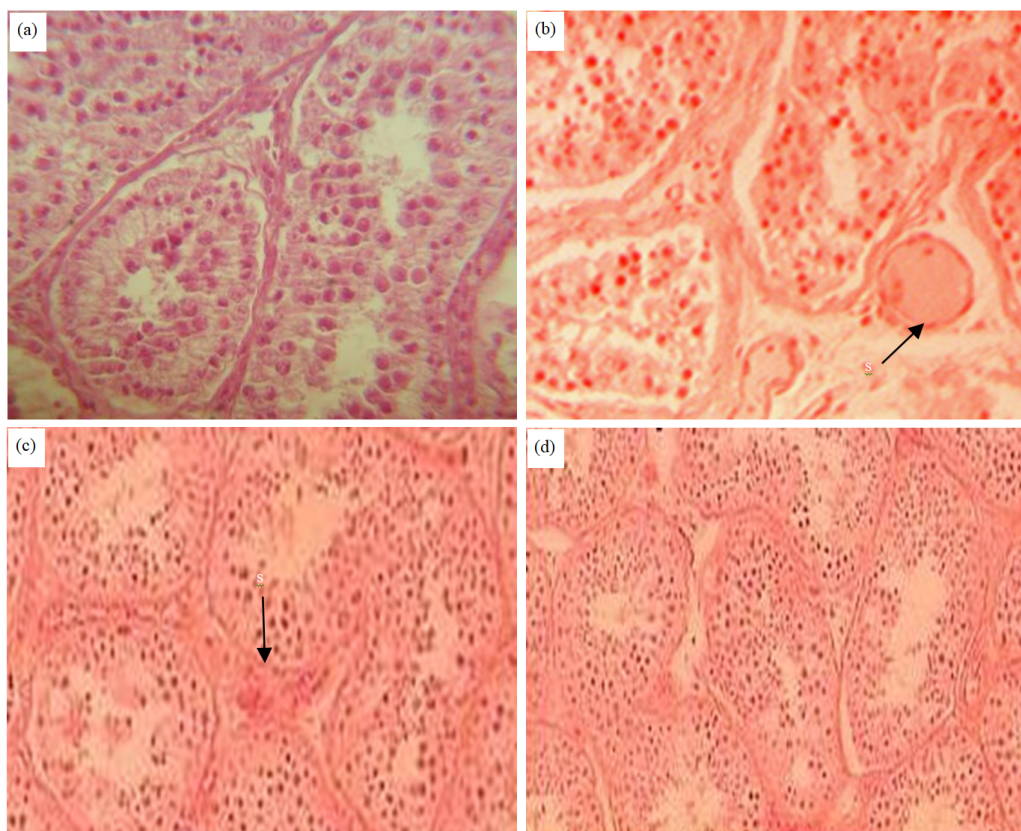


Fig. 1(a-d): Photomicrograph of testis exposed to CdSO₄ and treated with propolis extract
S with arrow indicates interstitial edema with congestion of blood vessels

The histological study showed, in control, testis are surrounded by a fibrous tissue. The testis are divided into lobules with several seminiferous tubules, which are surrounded by the interstitial tissue. Each tubule contained germ cells with various stages of spermatogonia, primary and secondary spermatocytes, spermatids and mature spermatozoa located in the centre of the tubule. In the interstitial tissue, Leydig cells were between the seminiferous tubules (Fig. 1a). Observation of treated testes with CdSO₄ showed necrosis and alteration. The architecture of seminiferous tubules was confused. Some germ cells had small and darkly stained nuclei. Dilation and congestion of blood vessels were observed in the interstitial spaces. Large Leydig cells were found in the interstitial tissue (Fig. 1b). Animals, initially treated with cadmium then with propolis, revealed that testis recovered more or less their normal structure reaching consistent seminiferous tubules (Fig. 1c). Rats, initially treated with propolis then administered with cadmium, displayed sections of testes less or more similar to the control sections. The germ cells appeared having regular shape with absence of cytoplasmic vacuolation (Fig. 1d).

DISCUSSION

The main objective of this present study was to explore whether the propolis effects enabled animals contaminated with cadmium-sulfate recovering their normal biochemical and histological status. Heavy metals are known as toxic and belonged to the class of the carcinogenic products. Several studies suggested that men exposed, at long time, to Cd toxicological environment developed infertility, a decreased sperm count and poor semen quality^{19,20}. Chronic exposure to cadmium induced harmful and deleterious effects on the human and animal health. The decrease in body weight gain of animals treated with cadmium were in concordance with recent studies²⁰. Cadmium slowed intestinal absorption of nutrients and strongly decreased the appetite of animals. No significant changes, in testes weight, were recorded after cadmium sulfate ingestion. This finding was in contrast with those of Haouem *et al.*²¹. The association between Cd toxicity and hyperglycemia, hypercholesterolemia and hyperlipidemia has been more extensively studied²². In this study, it was noticed in animals contaminated with Cd increased blood

glucose, cholesterol triglycerides, urea and creatinine levels. These outcomes were endorsed by the recent studies performed by Lei *et al.*²³ and Son *et al.*²⁴. Interesting results were obtained in Cd-exposed workers during 20 years. It was suggested they were characterized by high blood glucose levels and decreased serum insulin. In the same way, studies revealed an association between urinary cadmium levels and the prevalence of diabetes²². This present study displayed low red cells count and a decreased hemoglobin rate. This finding also showed an induced anemia cadmium toxicity. Cd induced damage to the erythrocyte membrane resulting in hemolysis which may be the cause of decreased of haemoglobin level³. This study also revealed an inverse association between prostate specific antigen (PSA) and blood cadmium level. In prostate cancer diagnosis, blood PSA was used as a tumoral marker for the early detection of prostate cancer²⁵. In the present study, accumulation of Cd in testes decreased serum testosterone. This result was in disagreement with the finding of Zeng *et al.*²⁶ and Haouem *et al.*²¹. CdSO₄ intoxicated animals showed an oxidative stress including in CAT, GST and GSH. These findings were similar to that found in the previous studies, which revealed cadmium induced oxidative stress inducing free radical generation²¹.

Propolis, natural product elaborated by honeybee, has pharmacological properties such as antibiotic activities, anticancer, antioxidant and anti-inflammatory²⁷. Propolis contained bioactive compounds including terpenes, sterols, polyphenols, phenolic acid and flavonoid²⁸. Propolis antioxidant activities were illustrated by scavenging reactive oxygen species (ROS) or free radicals and prevention against lipid peroxidation¹⁰. Propolis also stimulated antioxidant enzymes such as catalase and glutathione S-transferase against oxidative stress caused by ROS²⁹. The treatment animals with propolis significantly increased serum testosterone and also the anti-oxidative enzymatic activities of CAT and GST compared to controls and animals intoxicated with Cd. Furthermore, these results revealed treatment of rats with propolis plus CdSO₄ decreased glutathione (GSH) level at nearly normal. These results joined other studies that had already shown propolis protected against cadmium chloride toxicity³⁰. Several scientific research supported that treatment with propolis enhances damaged testes tissues and limited functional disruption induced by cadmium in testis tissue³¹. The histological changes in testes of animals treated with CdSO₄ were in agreement with the study conducted by Cilenk *et al.*⁸. The treatment animals with propolis improved

less or more the histopathological profile of testis. The propolis protective effects against Cd-induced testicular injuries was reported by Messaoudi *et al.*³².

These findings are debatable for different reasons; to change the extraction method and to use different organic solvent instead of ethanol, to explore the toxicity of the propolis, the alcohol propolis extract could be insufficient to promote relevant preventive effects as expected and finally to investigate synergy molecular reactions of bioactive compounds elucidating how do magical molecules acting in cells and fighting oxidative stress resulting from Cadmium's toxicity.

CONCLUSION

This study, first of its kind in Algeria, showed that exposure to Cadmium sulfate induced necrosis in testicular tissue, increased serum PSA and decreased blood testosterone. This study confirmed obvious results in the literature. The treatment with propolis had beneficial effects improving all the biochemical parameters. Therefore, the present study endorsed the preventive effects of propolis to minimize the Cadmium toxicity.

SIGNIFICANCE STATEMENT

This study discovers the high preventive effects of local Algerian propolis against the toxicity of cadmium sulfate in male rats. This study will help researchers to uncover the critical areas of the honeybee product therapy that many researchers were not able to explore. Thus, the new theory on honeybee derived products and food supplements may be arrived at the replacement, the synthetic chemical drugs by natural substances.

ACKNOWLEDGMENT

The authors wish to thank Dr. Z. Haddi for helping biochemical analysis and for his assistance in histological techniques.

REFERENCES

1. Gunnarsson, D., G. Nordberg, P. Lundgren and G. Selstam, 2003. Cadmium-induced decrement of the LH receptor expression and cAMP levels in the testis of rats. *Toxicology*, 183: 57-63.

2. Ji, Y.L., H. Wang, P. Liu, X.F. Zhao and Y. Zhang *et al*, 2011. Effects of maternal cadmium exposure during late pregnant period on testicular steroidogenesis in male offspring. *Toxicol. Lett.*, 205: 69-78.
3. Simsek, N., A. Karadeniz, Y. Kalkan, O.N. Keles and B. Unal, 2009. *Spirulina platensis* feeding inhibited the anemia- and leucopenia-induced lead and cadmium in rats. *J. Hazard. Mater.*, 164: 1304-1309.
4. Kim, J. and J. Soh, 2009. Cadmium-induced apoptosis is mediated by the translocation of AIF to the nucleus in rat testes. *Toxicol. Lett.*, 188: 45-51.
5. Ozawa, N., N. Goda, N. Makino, T. Yamaguchi, Y. Yoshimura and M. Suematsu, 2002. Leydig cell-derived heme oxygenase-1 regulates apoptosis of premeiotic germ cells in response to stress. *J. Clin. Invest.*, 109: 457-467.
6. Xu, L.C., H. Sun, S. Wang, L. Song, H.C. Chang and X.R. Wang, 2005. The roles of metallothionein on cadmium-induced testes damages in sprague-dawley rats. *Environ. Toxicol. Pharmacol.*, 20: 83-87.
7. Patrick, L., 2003. Toxic Metals and antioxidants: Part II. The role of antioxidants in arsenic and cadmium toxicity. *Altern. Med. Rev.*, 8: 106-128.
8. Cilenk, K.T., I. Ozturk and M.F. Sonmez, 2016. Ameliorative effect of propolis on the cadmium-induced reproductive toxicity in male albino rats. *Exp. Mol. Pathol.*, 101: 207-213.
9. Nakajima, Y., K. Tsuruma, M. Shimazawa, S. Mishima and H. Hara, 2009. Comparison of bee products based on assays of antioxidant capacities. *BMC Complement. Altern. Med.*, Vol. 9. 10.1186/1472-6882-9-4.
10. Yousef, M.I. and A.F. Salama, 2009. Propolis protection from reproductive toxicity caused by aluminium chloride in male rats. *Food Chem. Toxicol.*, 47: 1168-1175.
11. Bueno-Silva, B., S.M. Alencar, H. Koo, M. Ikegaki, G.V.J. Silva, M.H. Napimoga and P.L. Rosalen, 2013. Anti-inflammatory and antimicrobial evaluation of neovestitol and vestitol isolated from Brazilian red propolis. *J. Agric. Food Chem.*, 61: 4546-4550.
12. Liu, C.C., J.M. Hsu, L.K. Kuo and C.P. Chuu, 2013. Caffeic acid phenethyl ester as an adjuvant therapy for advanced prostate cancer. *Med. Hypotheses*, 80: 617-619.
13. Sarker, D., A.H.M. Reid, T.A. Yap and J.S. de Bono, 2009. Targeting the PI3K/AKT pathway for the treatment of prostate cancer. *Clin. Cancer Res.*, 15: 4799-4805.
14. Franchi, G.C., C.S. Moraes, V.C. Toreti, A. Dausch, A.E. Nowill and Y.K. Park, 2012. Comparison of effects of the ethanolic extracts of Brazilian propolis on human leukemic cells as assessed with the MTT assay. *Evid. Based Complement. Alternat. Med.*, Vol. 2012. 10.1155/2012/918956.
15. Bhadauria, M. and S.K. Nirala, 2009. Reversal of acetaminophen induced subchronic hepatorenal injury by propolis extract in rats. *Environ. Toxicol. Pharmacol.*, 27: 17-25.
16. Alvarez, S.M., N.N. Gomez, L. Scardapane, F. Zirulnik, D. Martinez and M.S. Gimenez, 2004. Morphological changes and oxidative stress in rat prostate exposed to a non-carcinogenic dose of cadmium. *Toxicol. Lett.*, 153: 365-376.
17. Habig, W.H., M.J. Pabst and W.B. Jakoby, 1974. Glutathione S-transferases: The first enzymatic step in mercapturic acid formation. *J. Biol. Chem.*, 249: 7130-7139.
18. Xu, J.B., X.F. Yuan and P.Z. Lang, 1997. Determination of catalase activity and catalase inhibition by ultraviolet spectrophotometry. *Chin. Environ. Chem.*, 16: 73-76.
19. Benoff, S., A. Jacob and I.R. Hurley, 2000. Male infertility and environmental exposure to lead and cadmium. *Hum. Reprod. Update*, 6: 107-121.
20. Siu, E.R., D.D. Mruk, C.S. Porto and C.Y. Cheng, 2009. Cadmium-induced testicular injury. *Toxicol. Applied Pharmacol.*, 238: 240-249.
21. Haouem, S., M.F. Najjar, A. El Hani and R. Sakly, 2008. Accumulation of cadmium and its effects on testis function in rats given diet containing cadmium-polluted radish bulb. *Exp. Toxicol. Pathol.*, 59: 307-311.
22. Tinkov, A.A., T. Filippini, O.P. Ajsuvakova, J. Aaseth and Y.G. Gluhcheva *et al*, 2017. The role of cadmium in obesity and diabetes. *Sci. Total Environ.*, 601: 741-755.
23. Lei, L.J., T.Y. Jin and Y.F. Zhou, 2005. Effects of cadmium on levels of insulin in rats. *J. Hyg. Res.*, 34: 394-396.
24. Son, H.S., S.G. Kim, B.S. Suh, D.U. Park and D.S. Kim *et al*, 2015. Association of cadmium with diabetes in middle aged residents of abandoned metal mines: The first health effect surveillance for residents in abandoned metal mines. *Ann. Occup. Environ. Med.*, Vol. 27. 10.1186/s40557-015-0071-2.
25. Andriole, G.L., E.D. Crawford, R.L. Grubb III, S.S. Buys and D. Chia *et al*, 2009. Mortality results from a randomized prostate-cancer screening trial. *N. Engl. J. Med.*, 360: 1310-1319.
26. Zeng, X., T. Jin, Y. Zhou and G.F. Nordberg, 2003. Changes of serum sex hormone levels and MT mRNA expression in rats orally exposed to cadmium. *Toxicology*, 186: 109-118.
27. Banskota, A.H., Y. Tezuka, K. Midorikawa, K. Matsushige and S. Kadota, 2000. Two novel cytotoxic benzofuran derivatives from Brazilian Propolis. *J. Nat. Prod.*, 63: 1277-1279.
28. Khalil, M.L., 2006. Biological activity of bee propolis in health and disease. *Asian Pac. J. Cancer Prev.*, 7: 22-31.

29. Jasprica, I., A. Mornar, Z. Debeljak, A. Smolic-Bubalo and M. Medic-Saric *et al.*, 2007. *In vivo* study of propolis supplementation effects on antioxidative status and red blood cells. *J. Ethnopharmacol.*, 110: 548-554.
30. Tohamy, A.A., E.M. Abdella, R.R. Ahmed and Y.K. Ahmed, 2014. Assessment of anti-mutagenic, anti-histopathologic and antioxidant capacities of Egyptian bee pollen and propolis extracts. *Cytotechnology*, 66: 283-297.
31. Kamiya, T., M. Izumi, H. Hara and T. Adachi, 2012. Propolis suppresses CdCl₂-induced cytotoxicity of COS7 cells through the prevention of intracellular reactive oxygen species accumulation. *Biol. Pharm. Bull.*, 35: 1126-1131.
32. Messaoudi, I., M. Banni, L. Said, K. Said and A. Kerkeni, 2010. Evaluation of involvement of testicular metallothionein gene expression in the protective effect of zinc against cadmium-induced testicular pathophysiology in rat. *Reprod. Toxicol.*, 29: 339-345.