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Effectiveness of some Antagonistic Fungi and Botanicals against Fusarium solani and Fusarium oxysporum f. sp. lycopersici Infecting Brinjal and Tomato Plants

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ABSTRACT

In the present study, the pathogenic fusaria viz., Fusarium solani f. sp. melongena and F. oxysporum f. sp. lycopersici causing brinjal and tomato wilt were isolated from soil as well as from the infected plant parts. In vitro efficacy of three medicinal plants viz., Azadirachta indica (leaf extract), Psidium guajava (leaf extract), Eucalyptus camaldulensis (bark extract) and three fungal antagonists viz., Trichoderma harzianum, T. atroviride and T. longibrachiatum were tested at 25, 50 and 75% (v/v) by poisoned food technique against both the pathogens. The assessment of fungitoxicity was carried out in terms of percent mycelial growth inhibition against the test fungi. Among different medicinal plant extracts, Azadirachta indica (leaf) was found significantly superior to the rest in suppressing the growth of F. oxysporum f. sp. lycopersici as 100% inhibition was recorded at 50 and 75% concentration followed by Psidium guajava and Eucalyptus camaldulensis on 7th day of inoculation. On the other hand, among different microbial antagonists, T. longibrachiatum against both the test fungi was highly effective and there was 100% inhibition of mycelial growth at 50 and 75% concentration, while T. harzianum was effective against F. oxysporum f. sp. lycopersici followed by T. atroviride as it completely inhibited the mycelial growth at 75% concentration.

Key words: Wilt disease, F. solani, F. oxysporum f. sp. lycopersici, medicinal plants, fungal antagonists

INTRODUCTION

The Fusarium wilt of tomato (Lycopersicion esculantum Mill) caused by Fusarium oxysporum f. sp. lycopersici (Sacc.) Snyder and Hansen (FOL) and Brinjal wilt (Solanum melongena L.) caused by Fusarium solani f. sp. melongena is recognized as a devastating disease in tomato and brinjal growing regions worldwide (Beckman, 1987; Bondad-Reantaso et al., 2005) besides in different regions of India in severe to moderate (50-60%) form (Sherf and Macnab, 1986; Jiskani et al., 2007; Chakraborty and Chatterjee, 2009). In Kanpur (U. P.), the vegetable growers suffer more than 25.14-47.94% crop losses due to Fusarium wilt of tomato and 18.61-33.37% due to Fusarium wilt of brinjal in heavily infested fields. Kapoor (2008) has reported that most of the common varieties of tomato and brinjal are susceptible and fungicides are frequently used to control the disease.

However, the soil-borne disease is very difficult and uneconomical to control with chemicals alone. In this context, biological control is an alternative and eco-friendly strategy for disease management (Bowers and Locke, 2000; Momin and Nair, 2001; Eziashi *et al.*, 2007).

The extract of medicinal plants (Daferera et al., 2000; Manczinger et al., 2002; Sridhar et al., 2003; Dwivedi and Enespa, 2012) as well as antimicrobial agents (Haggag and Mohamed, 2007; Dwivedi and Enespa, 2013) have been found effective as antifungal agent. The leaf extract of Azadirachta indica showed in vitro antifungal effect against Fusarium solani and Fusarium oxysporum f. sp. lycopersici (Jarvis and Morgan, 2000; Dai et al., 2001; Verma and Kharwar, 2006; Agbenin and Marley, 2006). The antifungal effect (bark extract) of Eucalyptus camaldulensis (Pattanaik et al., 2002), leaf extract of Psidium guajava (Khayungarnnawee et al., 2004) against F. solani and F. oxysporum f. sp. lycopersici have also been studied.

On the other hand, the antagonistic fungi especially *Trichoderma* spp. viz., *T. harzianum*, *T. atroviride* and *T. longibrachiatum* have been widely used against *F. solani* and *F. oxysporum* f. sp. lycopersici (Ahmed, 2011; Hend and Perveen, 2012). Christopher et al. (2010) have reported successful reduction of fusarial wilt in many crops with application of different species of *Trichoderma*. Rini and Sulochana (2007) and Kapoor (2008) have found *Trichoderma* spp. to be effective biocontrol agents against *F. solani* and *F. oxysporum*. However, it is also reported that all the isolates of *Trichoderma* spp. are not equally effective in the control of the pathogens in vitro and in vivo (Moussa et al., 2006; Morsy et al., 2009). Therefore, a specific effective native *Trichoderma* isolate has to be identified for successful control of a particular pathogen. The purpose of this study was to evaluate the efficacy of medicinal plants as well as fungal antagonists (*Trichoderma* spp.) against *F. solani* and *F. oxysporum* f. sp. lycopersici.

MATERIALS AND METHODS

Test fungal strains: The pathogenic fusaria viz., Fusarium solani and Fusarium oxysporum f. sp. lycopersici and fungal antagonists viz., Trichoderma harzianum, T. atroviride and T. longibrachiatum were isolated from the wilt affected (soil+plant parts) and healthy tomato and brinjal crop fields of Kanpur, Uttar Pradesh, India, using serial dilution technique as well as direct inoculation of plant parts. The cultures of the fungi were purified by single spore isolation technique on Czapek's Dox Agar medium (Chauhan et al., 2002). The isolated pathogenic and biocontrol mycoflora were identified on the basis of cultural and morphological characters (Gilman, 1957; Barnett and Hunter, 1972; Booth, 1985). The fungitoxicity was studied by poisoned food technique (Grover and Moore, 1962).

Preparation of crude plant extracts: Fresh plant materials of Azadirachta indica (leaves), Psidium guajava (leaves), Eucalyptus camaldulensis (bark) were washed thoroughly under running tap water followed by sterilized distilled water. The leaves were air dried and then grinded with the help of pestle and mortar. One mL of extract was added in 1ml distilled water (1:1 v/v) separately for each plant extract; filtered through muslin cloth and 100% plant extract solution was prepared. The extracts were poured in the flasks and further diluted to different concentrations (25, 50 and 75%) by adding sterile distilled water for further studies.

Determination of mycelial growth inhibition

Dry weight method: One hundred ml of broth amended with different filtrate of antagonistic fungi individually were inoculated with 6.0 mm agar discs in 250 mL Erlenmeyer flask. The flasks were then incubated at 25±2°C for 10 days. The fungal mats were removed by filtration (Whatman Filter Study 1 and then 42 Nos.), dried at 60±3°C for 24 h and dry weight (g) was recorded.

Linear growth method: The filtrates of the antagonistic Trichoderma spp. viz., T. harzianum, T. atroviride and T. longibrachiatum and extracts of medicinal plant viz., Azadirachta indica (leaves), Psidium guajava (leaves), Eucalyptus camaldulensis (bark) were added to autoclaved CZA medium to give fungal concentration of 25, 50 and 75% (v/v). The plates were inoculated with 6.0 mm disc of F. solani and F. oxysporum f. sp. lycopersici individually in the centre of each of the Petri plates and incubated at 25±2°C for 7 days. Three replicates were maintained for each treatment. The growth of fungus was measured on 7th day and mean of colony growth dia. (mm) was recorded and percentage of mycelial growth inhibition was calculated as compared to control (Gaspar et al., 2004).

Statistical analysis: The Data recorded during the course of investigation was subjected to two-way ANOVA without replication. The conclusion was drawn on the basis of analysis of variance. The calculated value of F was compared with Table value of F at 5% levels of significance for an appropriate degree of freedom.

RESULTS AND DISCUSSION

Evaluation of medicinal plants against fungal pathogens: The mean fungal radial growth (mm) for each treatment is presented in (Table 1) indicate the inhibition zone of different extracts. All the medicinal plant extracts showed significant inhibition in the mycelial growth of the test pathogens. The A. indica leaf extract against F. oxysporum f. sp. lycopersici was most effective as it completely inhibited the mycelial growth at the concentration of 50 and 75%. A. indica leaf extract at 75% concentration was also most effective against F. solani on 7th day of inoculation. The growth of Fusarium solani was inhibited at 25% concentration of Azadirachta indica (leaves),

Table 1: Efficacy of medicinal plant extracts against Fusarium solani f. sp. melongena and Fusarium oxysporum f. sp. lycopersici at different concentration on 7th day of inoculation

	Concentration (%)	Pathogens/mean colony dia. (mm)	
Medicinal plants		FS	FOL
Azadirachta indica (leaf extract)	25	26.75±1.26	11.50±0.58 aII
	50	18.75 ± 1.50	0.00±0.00 aI
	75	7.25±0.96 aI	0.00±0.00 aI
Psidium guajava (leaf extract)	25	53.75±1.50	58.50±2.38
	50	47.00±2.94	47.75±1.50
	75	44.25±0.00	45.50±0.57
Eucalyptus camaldulensis (bark extract)	25	70.00 ± 1.41	71.75±1.50
	50	56.50±1.73	65.75±0.96
	75	15.50±1.00 aII	62.75±0.50
Control		73.25±0.95	73.00±1.41

Values shown are the Mean \pm SE of 3 replicates, significant at p \leq 0.05. Application of A. indica on F. oxysporum f. sp. lycopersici at 50 and 75% concentration had highest inhibiting effect followed by 75% conc. in A. indica on F. solani. These biopesticides checked fungal growth by 100, 100 and 90.10%, respectively

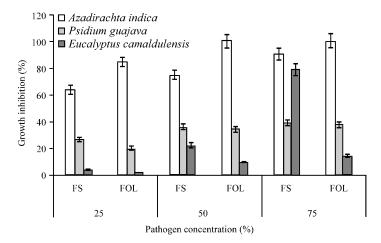


Fig. 1: Effect of medicinal plant extracts on radial growth of *F. solani* and *F. oxysporum* f. sp. *lycopersici* on 7th day of incubation

Psidium guajava (leaves) and Eucalyptus camaldulensis (bark) by 63.48, 26.62 and 4.44%, respectively. On the other hand, at 50% concentration of the foresaid medicinal plants inhibited the mycelial growth of Fusarium solani by 74.40, 35.84 and 22.87%, respectively. However, maximum inhibition i.e., 90.10, 39.59 and 78.84% was recorded at 75% concentration. The foresaid medicinal plants at 25% concentration inhibited the mycelia growth of Fusarium oxysporum f. sp. lycopersici by 84.25, 19.86 and 1.72%, respectively. On the other hand, at 50% concentration of these plants inhibited mycelial growth by 100.00, 34.59 and 9.93%, respectively. Maximum percentage inhibition i.e., 100.00, 37.67 and 14.04% was recorded at 75% concentration of Azadirachta indica (leaves) against F. oxysporum f. sp. lycopersici as indicated in Fig. 1.

Our findings are in conformity with Joseph and Priya (2010) with respect to efficacy of guava leaf extract against Aspergillus niger and Aspergillus aculeatus. Similar results were reported (Mossini et al., 2004; Niaz and Kazmi, 2005) with the increase in concentration of the medicinal plants, the greater inhibition of mycelial growth of Fusarium spp. and Aspergillus alternata at 0.15 and 0.1% concentration. Antifungal activities and chemical properties of Azadirachta indica oil and leaf extracts on the growth of Aspergillus, Rhizopus, Alternaria solani and Alternaria brassicae have been also reported (Mondali et al., 2009; Suleiman, 2010; Sasode et al., 2012). Bajwa and Ifftikhar (2005) evaluated Eucalyptus camaldulensis and Datura tetramera against fungal pathogens causing wilt in vegetable crops.

Evaluation of Trichoderma spp. against fungal pathogens: The percentage reduction of F. solani and F. oxysporum at different concentration of culture filtrates of T. harzianum, T. atroviride and T. longibrachiatum is presented in Table 2. The data revealed a significance increase (p<0.001) in colony growth reduction of F. solani and F. oxysporum f. sp. lycopersici with increasing the concentration of both the fungal filtrates. T. harzianum, T. atroviride and T. longibrachiatum filtrates inhibited the growth of F. solani by 84.01, 48.91 and 100% and F. oxysporum f. sp. lycopersici by 100.00, 73.67 and 100.00%, respectively at 75% concentration. At 50% concentration, the foresaid antagonists inhibited the growth of F. solani by 74.88, 44.06 and 86.87% and F. oxysporum f. sp. lycopersici by 92.35, 65.74 and 100.00%, respectively. At lower concentration (25%), T. harzianum, T. atroviride and T. longibrachiatum were least effective and

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Table 2: Effect of *Trichoderma* spp. against *Fusarium solani* (FS) and *Fusarium oxysporum* f. sp. *lycopersici* (FOL) at different concentration on 7th day of inoculation

		Pathogens/mean colony diameter (mm)		Mycelial dry weight (g) of
Fungal antagonists	Concentration (%)			
		FS	FOL	fungal antagonists
T. richoderma harzianum	25	46.25±0.96	19.75±2.50	2.70
	50	22.00 ± 1.82	6.75±0.95 aII	
	75	14.00±1.82 aII	0.00±0.00 aI	
T. richoderma atroviride	25	50.75±0.96	37.75±1.50	1.48
	50	49.00±0.81	30.25±0.50	
	75	44.75 ± 0.50	23.25±1.26	
T. richoderma longibrachiatum	25	21.75±1.26	18.75±0.96	2.16
	50	11.50±0.58 aII	0.00±0.00 aI	
	75	0.00±0.00 aI	0.00±0.00 aI	
Control		87.60±0.14	88.30±0.41	

Values shown are the Mean \pm SD of 3 replicates, significant at p \leq 0.05. Application of *T. longibrachiatum* at 50 and 75% concentration had highest inhibiting effect on both the pathogens followed by *T. harzianum* at 75% concentration on *Fusarium oxysporum*. These fungal antagonists checked fungal growth by 100, 100 and 100%, respectively

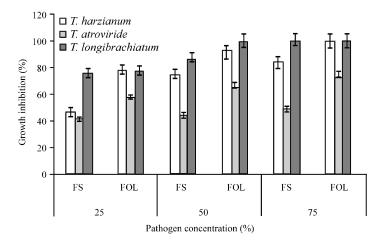


Fig. 2: Percentage inhibition of *F. solani* f. sp. *melongena* and *F. oxysporum* f. sp. *lycopersici* at different concentration of *Trichoderma* spp. on 7th day of inoculation

inhibited the growth of *F. solani* by 47.20, 42.06 and 75.17% and *F. oxysporum* f. sp. *lycopersici* by 77.63, 57.25 and 78.76%, respectively compared to control on 7th day of inoculation (Fig. 2)

Our findings are in conformity with Devi and Singh (2012) with respect to in vitro efficacy of Trichoderma spp. against Fusarium oxysporum f. sp. lycopersici causing tomato wilt. Antagonistic activity of three isolates of Trichoderma spp. against F. solani and F. oxysporum f. sp. lycopersici due to parasitism, competition and antibiosis was reported by Elad (2000); Freeman et al. (2004); Dubey et al. (2007); Anwar et al. (2008); coiling and penetration of antagonistic hyphae of T. virens and T. harzianum around the hyphae of F. solani and their lysis (Kumar and Dubey, 2001; Zaldivar et al., 2001) and production of organic metabolites (Haggag and Mohamed, 2007). It was observed that with the increase in concentration of the culture filtrates of Trichoderma spp. there was greater inhibition of the mycelial growth of Rhizoctonia solani, Sclerotinia, Fusarium, Pythium (Ashrafizadeh et al., 2005; Yigit and Dikilitas, 2007; Khan and Sinha, 2007).

CONCLUSION

In the present study, the medicinal plant (Azadirachta indica) leaf extract and fungal antagonists viz., T. longibrachiatum and T. harzianum were most effective against both the pathogens. Therefore, these bio-pesticides (botanicals and bio-agents) can be used as an alternative to pesticides to minimize the wilt disease of these crops besides improving the yield as they are environmental safe.

REFERENCES

- Agbenin, O.N. and P.S. Marley, 2006. *In-vitro* assay of some plant extracts against *Fusarium oxysporum* f. sp. *lycopersici* casual agent of tomato wilt. J. Plant Prot. Res., 46: 215-220.
- Ahmed, M., 2011. Management of Fusarium wilt of tomato by soil amendment with *Trichoderma koningii* and a white sterile fungus. Indian J. of Res., 5: 35-38.
- Anwar, A., G.N. Bhat, K.A. Bhat, 2008. Mycoparasitic behaviour of certain bioagents against sheath blight pathogen (*Rhizoctonia solani*) of rice. J. Mycol. Plant Pathol., 38: 135-137.
- Ashrafizadeh, A., H.R. Etebarian and H.R. Zamanizadeh, 2005. Evaluation of *Trichoderma* isolates for biocontrol of fusarium wilt of melon. Iranian J. Plant Pathol., 41: 39-57.
- Bajwa, R. and S. Iftikhar, 2005. Antifungal activity of allelopathic plant extracts Vis: *In vitro* control of fungal pathogens by aqueous leaf extracts of *Eucalyptus*. Mycopathology, 3: 7-12.
- Barnett, H.L. and B.B. Hunter, 1972. Illustrated Genera of Imperfect Fungi. 2nd Edn., Minneapolis Burgress Publishing Co., Minneapolis, pp. 241.
- Beckman, C.H., 1987. The Nature of Wilt Diseases of Plants. The American Phytopathologycal Society Press, USA., ISBN-13: 978-0890540749, pp: 175.
- Bondad-Reantaso, M.G., R.P. Subasinghe, J.R. Arthur, K. Ogawa and S. Chinabut *et al.*, 2005. Disease and health management in Asian aquaculture. Vet. Parasitol., 132: 249-272.
- Booth, C., 1985. The Genus *Fusarium*. 2nd Edn., Commonwealth Mycological Institute, Kew, Surrey, England, Pages: 237.
- Bowers, J.H. and J.C. Locke, 2000. Effect of botanical extracts on the population density of *Fusarium oxysporum* in soil and control of *Fusarium* wilt in the greenhouse. Plant Dis., 84: 300-305.
- Chakraborty, M.R., N.C. Chatterjee and T.H. Quimio, 2009. Integrated management of fusarial wilt of eggplant (*Solanum melongena*) with soil solarization. Micol. Apll. Int., 21: 25-36.
- Chauhan, J.B., R.B. Subramanian and P.K. Sanyal, 2002. Influence of heavy metals and a fungicide on growth profiles of nematophagous fungus Arthrobotrys musiformis, a potential biocontrol agent against animal parasitic nematodes. Indian J. Environ. Toxicol., 12: 22-25.
- Christopher, D.J., T.S. Raj, S.U. Rani and R. Udhayakumar, 2010. Role of defense enzymes activity in tomato as induced by *Trichoderma virens* against *Fusarium* wilt caused by *Fusarium oxysporum* f. sp. *lycopersici*. J. Biopesticides, 3: 158-162.
- Daferera, D.J., B.N. Ziogas and M.G. Polissiou, 2000. GC-MS analysis of essential oils from some Greek aromatic plants and their fungitoxicity on *Penicillium digitatum*. J. Agric. Food Chem., 48: 2576-2581.
- Dai, J., V.A. Yaylayan, G.S.V. Raghavan, J.R. Pare and Z. Liu, 2001. Multivariate calibration for the determination of total azadirachtin-related limonoids and simple terpenoids in neem extracts using vanillin assay. J. Agric. Food Chem., 49: 1169-1174.

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- Devi, T.N. and M.S. Singh, 2012. Evaluation of suitable fungicide for integration with *Trichoderma* isolates for the control of tomato wilt. J. Mycopathol. Res., 50: 223-228.
- Dubey, S.C., M. Suresh and B. Singh, 2007. Evaluation of *Trichoderma* species against *Fusarium oxysporum* f. sp. *ciceris* for integrated management of Chickpea wilt. Biol. Control J., 40: 118-127.
- Dwivedi, S.K. and Enespa, 2012. Effectiveness of extract of some medicinal plants against soil borne fusaria causing diseases on *Lycopersicon esculantum* and *Solanum melongena* plants. Int. J. Pharma. Biol. Sci., 3: 1171-1180.
- Dwivedi, S.K. and Enespa, 2013. *In vitro* efficacy of some fungal antagonists against *Fusarium solani* and *Fusarium oxysporum* f. sp. *lycopersici* causing brinjal and tomato wilt. Int. J. Biol. Pharm.. Res., 4: 46-52.
- Elad, Y., 2000. Biological control of foliar pathogens by means of *Trichoderma harzianum* and potential modes of action. Crop Prot., 19: 709-714.
- Eziashi, E.I., I.B. Omamor and E.E. Odigie, 2007. Antagonism of *Trichoderma viride* and effects of extracted water soluble compounds from *Trichoderma* species and benlate solution on *Ceratocystis paradoxa*. Afr. J. Biotechnol., 6: 388-392.
- Freeman, S., D. Minz, I. Kolesnik, O. Barbul and A. Zveibil *et al.*, 2004. *Trichoderma* biocontrol of *Colletotrichum acutatum* and *Botrytis cinerea* and survival in strawberry. Eur. J. Plant Pathol., 110: 361-370.
- Gaspar, H., S.S. Feio, A.I. Rodrigues and R. Soest, 2004. Antifungal activity of (+)-curcuphenol, a metabolite from the marine sponge didiscus oxeata. Mar. Drugs, 2: 8-13.
- Gilman, J.D., 1957. A Manual of Soil Fungi. Iowa State University Press, Ames, Iowa, USA., Pages: 450.
- Grover, R.K. and J.D. Moore, 1962. Toximetric studies of fungicides against brown rot organisms Sclerotina flucticola and S. laxa. Phytopathology, 52: 876-880.
- Haggag, W.M. and H.A.A. Mohamed, 2007. Biotechnological aspects of microorganisms used in plant biological control. American-Eurasian J. Sustainable Agric., 1: 7-12.
- Hend, A. and K. Perveen, 2012. Biological control of *Fusarium* wilt of tomato by antagonist fungiand cyanobacteria. Afri. J. of Biotechnol., 11: 1100-1105.
- Jarvis, A.P.E. and D. Morgan, 2000. Analysis of small samples of limonoids of neem (*Azadirachta indica*) using solid phase extraction from tissue culture. Photochem. Anal., 11: 184-189.
- Jiskani, M.M., M.A. Pathan, K. Wagan, H.M. Imran and H. Abro, 2007. Studies on the control of tomato damping-off disease caused by *Rhizoctonia solani* Kuhn. Pak. J. Bot., 39: 2749-2754.
- Joseph, B. and R.M. Priya, 2010. *Invitro* Antimicrobial activity of *Psidium guajava* L. leaf essential oil and extracts using agar well diffusion method. Int. J. Curr. Pharma. Res., 2: 28-32.
- Kapoor, A.S., 2008. Biocontrol potential of *Trichoderma* spp. against important soil borne diseases of vegetable crops. Indian Phytopathol., 61: 492-498.
- Khan, A.A. and A.P. Sinha, 2007. Screening of *Trichoderma* spp. against *Rhizoctonia solani* the causal agent of rice sheath blight. Indian Phytopath., 60: 450-456.
- Khayungarnnawee, A., W. Phatvet, T. Sematong, S. Thubthimthed and T. Suntorntanasat, 2004. P12 Preminary Study on sedative effect of Guava (*Psidium guajava* L.) Leaf oil. Thai J. Pharmacol., 29: 76-77.
- Kumar, D. and S.C. Dubey, 2001. Management of collar rot of pea by the integration of biological and chemical methods. Indian Phytopathol., 54: 62-66.

Asian J. Plant Pathol., 8 (1): 18-25, 2014

- Manczinger, L., Z. Antal and L. Kredics, 2002. Ecophysiology and breeding of mycoparacitic *Trichoderma strains* (a review). Acta Microbiol. Immunol. Hungarica, 49: 1-14.
- Momin, R.A. and G. Nair, 2001. Mosquitocidal, nematicidal and antifungal compounds from *Apium graveolens* L. Seeds. J. Agric. Food Chem., 49: 142-145.
- Mondali, N.K., A. Mojumdar, S.K. Chatterje, A. Banerjee, J.K. Datta and S. Gupta, 2009. Antifungal activities and chemical characterization of Neem leaf extracts on the growth of some selected fungal species *in vitro* culture medium. J. Applied Sci. Environ. Manage., 13: 49-53.
- Morsy, E.M., K.A. Abdel-Kawi and M.N.A. Khalil, 2009. Efficiency of *Trichoderma viride* and *Bacillus subtilis* as biocontrol agents against *Fusarium solani* on tomato plants. Egrupt. J. Phytopathol., 31: 47-57.
- Mossini, S.A., K.P. de Oliveira and C. Kemmelmeier, 2004. Inhibition of patulin production by *Penicillium expansum* cultured with neem (*Azadirachta indica*) leaf extracts. Basic Microbiol., 44: 106-113.
- Moussa, A.L., S.S. Fahmy and M.A.A. Shaltout, 2006. Evaluation of some bacterial isolates and compost tea for Bio-controlling *Macrophomina phaseolina* and *Sclertium roefsii* infected sunflower. Egypt. J. Agric. Res., 84: 1331-1344.
- Niaz, I. and S.A.R. Kazmi, 2005. Neem seed coal oil fractions on stored on stored grain fungi. Int. J. Biol. Biotechnol., 2: 705-706.
- Pattanaik, S., V.R. Subramanyam and C. Kole, 2002. Antibacterial and antifungal activity of the essential oils *in vitro*. Microbiosis, 86: 237-246.
- Rini, C.R. and K.K. Sulochana, 2007. Usefulness of *Trichoderma* and *Pseudomonas* against *Rhizoctonia solani* and *Fusarium oxysporum* infecting tomato. J. Tropical. Agric., 45: 21-28.
- Sasode, R.S., S. Prakash, A. Gupta, R.K. Pandya and A. Yadav, 2012. *In vitro* study of some plant extracts against *Alternaria brassicae* and *Alternaria brassiciola*. J. Phytol., 4: 44-46.
- Sherf, A.F., A.A. Macnab, 1986. Vegetable diseases and their control. A Wiley-interscience Publication, New York, pp. 381-396.
- Sridhar, S.R., R.V. Rajagopal, R. Rajavel, S. Masilamani and S. Narasimhan, 2003. Antifungal activity of some essential oils. J. Agric. Food Chem., 51: 7596-7599.
- Suleiman, M.N., 2010. Fungitoxic activity of neem and pawpaw leaves extracts on *Alternaria solani*, causal organism of yam rots. Adv. Environ. Biol., 4: 159-161.
- Verma, V.C. and R.N. Kharwar, 2006. Efficacy of neem leaf extract against it's own fungal endophytic *Curvularia lunata*. J. Agric. Technol., 2: 329-335.
- Yigit, F. and M. Dikilitas, 2007. Control of fusarium wilt of tomato by combination of fluorescent Pseudomonas, non-pathogen Fusarium and Trichoderma harzianum T-22 in greenhouse conditions. Plant Pathol. J., 6: 159-163.
- Zaldivar, M., J.C. Velasquez, I. Contreras and L.M. Perez, 2001. *Trichoderma aureoviride* 7-121, a mutant with enhanced production of lytic enzymes: Its potential use in waste cellulose degradation and/or biocontrol. Electron. J. Biotechnol., 4: 1-9.