

Asian Journal of **Plant Pathology**

ISSN 1819-1541



www.academicjournals.com

Asian Journal of Plant Pathology

ISSN 1819-1541 DOI: 10.3923/ajppaj.2017.35.47



Research Article Efficacy of Some Nanoparticles to Control Damping-off and Root Rot of Sugar Beet in El-Behiera Governorate

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Abstract

Background: This study was conducted to examine the potential antifungal activity and protective effect of some nanoparticles in controlling root rot disease of sugar beet. Magnesium oxide, titanium dioxide and zinc oxide nanoparticles (MgO, TiO₂ and ZnO NPs) were investigated against three pathogenic fungi isolated from sugar beet roots, Fusarium oxysporum f. sp., betae, Sclerotium rolfsii and *Rhizoctonia solani in vitro* and under greenhouse conditions. **Materials and Methods:** Solutions of particles of MgO, TiO₂ and ZnO nanoparticles (NPs) were used in different concentrations *in vitro* to study the inhibition of fungal radial growth and by spraying fungal culture of the tested fungi to test the effect of nanoparticles on hyphal and spore formation and Scanning Electron Microscopy (SEM) was used to visualize the effect of this application. Greenhouse experiment was conducted and sugar beet seeds (cv., Kawamera) were treated with the nanoparticles concentrations as seed dressing then seeds were planted in infested soil with the tested fungi. Disease assessment was evaluated, also the effect of nanoparticles on the vegetative and the chemical characteristics of sugar beet were examined. Results: Obtained results showed that the three NPs tested with concentrations investigated (25, 50 and 100 ppm) increased the in vitro fungal growth inhibition by reducing the radial fungal growth with the best effect was recorded with the highest concentration. Meanwhile, TiO₂ NP (100 ppm) showed the highest effect in decreasing mean radial growth by 77.25%. Also, TiO₂ showed 100% inhibition on sclerotia formation of Sclerotium rolfsii, while MgO was most effective and decreased sporulation (number of conidia) of *Fusariun oxysporum* f. sp., betae by 69.23%. The greenhouse experiment showed that the tested all tested NPs significantly decreased the developed root rot severity and TiO₂ was most effective and decreased it to 1.39% compared to 28.2% for the untreated. On the other hand, treatments with the tested NPs significantly increased root fresh weight (biomass) of the plants and also the dry weight compared to the untreated infested control. This was accompanied with an increase in sucrose and Total Soluble Solids (TSS) and also the total phenol content and activity of the defense related enzyme, polyphenol oxidase. **Conclusion:** Based on the obtained results the use of magnesium oxide, titanium dioxide and zinc oxide nanoparticles could be a good and environmentally safe alternative of fungicides in controlling damping-off and root rot disease of sugar beet.

Key words: Nanoparticles, sugar beet, damping-off disease, root rot disease, biological control, magnesium oxide, titanium dioxide, zinc oxide

Received: August 30, 2016

Accepted: November 18, 2016

Published: December 15, 2016

Citation: Eman El-Argawy, M.M.H. Rahhal, A. El-Korany, E.M. Elshabrawy and R.M. Eltahan, 2017. Efficacy of some nanoparticles to control damping-off and root rot of sugar beet in El-Behiera governorate. Asian J. Plant Pathol., 11: 35-47.

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Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Sugar beet (*Beta vulgaris* L.) as an industrial crop is cultivated in 48 countries in the world for a total land of over 9 million hectares and the second highest source of sugar providing after sugar-cane. Around 30% of the total annual sugar production in the world is from sugar beets¹. This plant is a good choice for crop rotation because it enriches soil with nutrients^{2,3}.

Sugar beet plants are often attacked by several pathogens such as fungi, bacteria and viruses which cause great losses in yield⁴. Root rot of sugar beet is considered the most effective disease that affects yield and quality as well as its sugar production. Several conventional methods have been used for the control of root rot of sugar beet. Some of these methods such the use of pesticides causes hazardous effects on the environment and human health. A great effort has been given to the development of safe non-traditional management methods that pose less danger to humans and animals^{5,6}. Thus, use of nanoparticles has been suggested by Kumar and Yadav⁷ as an alternative and effective approach which is eco-friendly and cost effective for the control of pathogenic microbes. In recent years, nanoparticle (NP) materials have received increasing attention due to their unique physical and chemical properties. The antimicrobial activity of the nanoparticles is known to be a function of the surface area in contact with the microorganisms. The small size and the high surface to volume ratio enhance their interaction with the microbes to carry out a broad range of probable antimicrobial activities^{8,9}. Some of the nanoparticles that have entered into the arena of controlling plant diseases are nano forms of silver¹⁰⁻¹², copper¹³, titanium dioxide¹⁴, zinc oxide¹⁵ and carbon¹⁶. Meanwhile, nanoparticles play unique role by contributing direct uptake and accumulation of silica, which leads to leaf erectness and enhances defense response to fungal pathogens¹⁷. The ZnO NPs at concentrations greater than 3 mmol L⁻¹ significantly inhibited the growth of Botrytis cinerea and Penicillium expansum¹⁸. Wani et al.¹⁹ reported that ZnO and MgO nanoparticles brought about significant inhibition to the germination of Alternaria alternate, Fusarium oxysporum, Rhizopus stolonifer and Mucor plumbeus spores. Meanwhile, nanoparticles of ZnO and TiO₂ showed antifungal effect against *Macrophomina* phaseolina. Also, Min et al.²⁰ showed that silver nanoparticles strongly inhibited the fungal growth and sclerotial germination of Rhizoctonia solani, Sclerotinia sclerotiorum and S. minor. In the present study, nanoparticles of MgO, ZnO and TiO₂ were tested for antimycotic efficacy to

control the root rot fungi affecting sugar beet in El-Behiera governorate and surrounding area as an alternative for the chemical fungicides.

MATERIALS AND METHODS

Tested sugar beet root rots fungi: Three good growing, highly virulent isolates of the sugar beet root rot fungi, *Fusarium oxysporum* f. sp., *betae, Sclerotium rolfsii* and *Rhizoctonia solani* were obtained from fungal collections of Plant Pathology Department, Faculty of Agriculture, Damnhour University. These isolates were previously isolated by the researcher from sugar beet fields, showed root rot symptoms in El-Behiera governorate during 2012-2013 growing season and the identification of the recovered fungi were done at the Department of Mycology and Plant Diseases Survey, Plant Pathology Research Institute, ARC, Giza. Confirmations were done to the identified fungi by comparison with those from the culture type collection of Maize and Sugar Crops Diseases Research Section of ARC, Giza.

Tested nanoparticle: Magnesium oxide (MgO), zinc oxid (ZnO) and titanium dioxide (TiO₂) nanoparticles (NPS) were obtained from MKnano, Canada and tested in the present study for their efficiency to control the root rot fungi affecting sugar beet cultivation in El-Behiera governorate. According to the source, the particles size ranged from $20-30\pm10$ nm and spherical in shape. Size and morphology of MgO, ZnO and TiO₂ nanoparticles were confirmed by Scanning Electron Microscope (SEM) at Faculty of Science, Alexandria University (Fig. 1a-c). The elemental compositions of bio-transformed product containing nanoparticles in the solution were confirmed by Transmission Electron Microscopy (TEM) equipped with energy dispersive x-ray spectroscopy (EDX). The TEM–EDX (Fig. 1d-f) clearly show that the Zn nanoparticles show the maximum intensity at 8.467-8.807 keV, whereas, Mg and Ti showed maximum intensity at 1.148-1.347 and 4.6, 4.367-4.648 keV, respectively. Results clearly exhibited the purity of the metal nanoparticles.

In vitro effect of nanoparticles on growth of the sugar beet root rots fungi: Three nanoparticle, MgO, ZnO and TiO_2 were tested in the present study. Different concentrations of nanoparticles (25, 50 and 100 ppm) were prepared by diluting the original stock solution (1000 ppm) using sterile deionized water. All solutions were stored at 4°C until use. Sterile deionized water was used as control.



Fig. 1(a-c): Scanning Electron Microscopy (SEM) images and Energy Dispersive X-ray (EDX) of (a) Magnesium oxide (MgO), (b) Titanium dioxide (TiO₂) and (c) Zinc oxide (ZnO) nanoparticles use in the present study

Effect on radial growth: The *in vitro* assay was performed by the agar dilution method described by Fraternale *et al.*²¹ with some modification. The autoclaved PDA media with concentrations of 25, 50 and 100 ppm for all nanoparticles tested, keeping one as control (PDA without nanoparticles) and NPs solution were poured into the petri dishes (9 cm diameter) before pouring the plates. Then, a disc of 0.5 cm diameter was taken from the edge of 7 day-old cultures of the tested fungi was placed in the center of each petri dish. The petri dish with the inoculum was then incubated at $25\pm2^{\circ}$ C. Then, 7 days after incubation or when the fungal growth in the control completely colonized the plate, radial growth, sporulation and sclerotia formation of sugar beet root rot fungi were investigated. All tests were performed in four replicate plates. **Effect on fungal radial growth:** Percentage of inhibition of mycelial radial growth was calculated according to Kaur *et al.*²² using the equation:

Inhibition (%) =
$$\frac{dc-dt}{dc} \times 100$$

where, dc is a mycelial growth in control and dt is a mycelial growth in the treatment.

Effect on sporulation and sclerotia formation: Sporulation (conidia number) of the sugar beet root rot fungus *Fusarium oxysporum* f. sp., *betae* was calculated using a hemaecytometer. Also number of sclerotia of *Sclerotium rolfsii* was recorded using 40X magnifying lens.

Effect on hyphal and spore morphology: Petri dishes containing healthy 10 day-old cultures on PDA of the sugar beet root rot fungi were sprayed with 1 mL of 100 ppm MgO, ZnO and TiO₂ nanoparticle solutions. At the same time cultures were sprayed with sterile deionized water and served as control. Twenty four hours later, the cultures were investigated under a Hitachi S-3500N scanning electron microscope at Faculty of Science, Alexandria University according to Elamawi and El-Shafey²³ to reveal the effect of nanoparticles on hyphae and spore morphology.

In vivo effect of nanoparticles on root rot and damping-off

on sugar beet: Greenhouse experiments were carried out in 2015 to study the efficacy of ZnO and MgO and TiO₂ nanoparticles to control root rot and damping-off of sugar beet. The pathogens were grown on medium of sand and corn (2:3 w/w) in glass bottles for 2 weeks at 27 °C. Plastic pots (25 cm diameter) were filled with soil sterilized with formalin (5%) as 3 kg soil pot⁻¹ and left for 1 week until complete formalin evaporation. Pot soil (1 clay:1 sand v/v) was infested by mixing the inoculum of fungi at the rate of 2% of soil weight according to Papavizas and Devey²⁴ and Marwa *et al.*²⁵.

The infested soil was watered and left for a week before planting to stimulate the fungal growth and ensure its establishment in the soil. On the other hand, nanoparticles solution were prepared (100 ppm+0.2% tween 80) and used as seed coating before planting and vitavax 200 fungicide (3 g kg⁻¹) were used as a control treatment. The pots were then planted with the sugar beet treated seeds, cv., Kawamera, (10 seeds pot⁻¹), watered and fertilized as usual. Each treatment had three pots and other pots planted with seeds in sterilized soil and served as control.

Disease assessments: Pre and post emergence damping off were calculated after 15 and 45 days of planting, respectively, according to El-Shafey *et al.*²⁶ as follows:

Pre-emergence damping-off (%) = $\frac{\text{No. of non-emerged seed}}{\text{Total No. of seeds sown}} \times 100$

Post-emergence (%) = $\frac{\text{No. of died plant}}{\text{Total No. of emerged plant}} \times 100$

Root rot severity was scored 150 days after planting based on Rowe²⁷ and Liu *et al.*²⁸ with the following ratings: 0: No internal or external browning, 1: No internal browning, with superficial lesions (\leq 25%) on tap root, 2: Slight internal browning with (<25 to \leq 50%) surface covered with cankers,

3: Moderate internal browning with <50 to \geq 75% cankers, 4: Severe internal and external (<75%) browning.

Disease severity =
$$\frac{\text{Sum } (n \times r0) + (n \times r1) + \dots + (n \times r4)}{4 \text{ N}} \times 100$$

where, n is No. of plants in each numerical rate (r0....r4) and N is the total No. of plants multiplied by the maximum numerical rate r4.

Effect of nanoparticles on the vegetative and the chemical characteristics of sugar beet: At the end of the pot experiment (150 days after planting), root yield was estimated as root fresh weight and dry weight, as well as the Total Soluble Solids (TSS), sucrose, total phenols and the defense related enzyme polyphenol oxidase in plant roots as follows.

Fresh and dry weight: After harvest, plant roots were thoroughly washed with tap water and dried at room temperature. Roots were then sliced and dried at 80°C for 72 h in air drying oven until constant weight was reached.

Total Soluble Solids (TSS) and sucrose content: The TSS was measured in juice of fresh roots by using hand refractometer (Hycle groupe lifasa bio 21320 Pouuilly by Auxxois-Fransa). Sucrose percent was determined by using polarimetric on lead acetate extract of fresh macerated roots according to the method of Carruthers and Oldfield²⁹ and Fatouh³⁰.

Total phenol: Total phenolic content of fresh root was estimated by Folin Ciocalteu method of Zieslin and Ben-Zaken³¹ with some modification. One gram of sample was homogenized in 10 mL of 80% methanol and agitated for 15 min at 70°C, then 1 mL of methanolic extract was added to 5 mL of distilled water and 250 μ L of Folin Ciocalteu reagent and incubated at 25°C, after 3 min, 1 mL of the saturated solution of sodium carbonate and 1 mL of distilled water were added and the reaction mixtures were incubated further for 1 h at 25°C. The absorption of the developed blue color was measured using spectrophotometer at 725 nm. The total soluble phenol content was calculated according to a standard curve obtained from a Folin Ciocalteu reagent with a phenol solution and expressed as catechol equivalent per gram of fresh tissue.

Polyphenol oxidase assay: Enzyme extraction was conducted according to the method described by Maxwell and Bateman³² as follows. The root tissues were grounded in a mortar with 0.1 M sodium phosphate buffer at pH 7.1

(2 mL g⁻¹ fresh root tissues). The triturated tissues were strained through layer of cheesecloth and filtrates were centrifuged at 3000 rpm for 20 min at 6°C and the supernatant fluids were used for enzyme assays. The activity of polyphenol oxidase was measured using spectrophotometer (Spectronic 601). The control cuvette contained the buffer solution plus distilled water. The reaction mixture contained 0.5 mL enzyme extract, 0.5 mL sodium phosphate buffer at pH 7 and 0.5 mL of catechol brought to a final volume of 3 mL with distilled water. The activity of polyphenol oxidase was expressed as the change in absorbance per minute in 1.0 g fresh weight at 495 nm. The polyphenol oxidase activity was measured after 150 days of inoculation. All the experiments were repeated thrice.

Statistical analysis: Data were analyzed using one-way analysis of variance (ANOVA) and the least significant difference test to estimate statistical differences between means at p = 0.05.

RESULTS

In vitro effect of nanoparticles on growth of the sugar beet root rot fungi

Effect on fungal radial growth: Data presented in Table 1 showed that all the tested nanoparticles, MgO, TiO₂ and ZnO

at the different tested concentrations (25, 50 and 100 ppm) significantly inhibited the fungal radial growth of the tested sugar beet root rots fungi on PDA compared to the control (sterile deionized water). Meanwhile, TiO_2 at 100 and 50 ppm showed the highest mean inhibition effects being 77.52 and 69.72% for both concentrations, respectively. This was followed by 100 ppm MgO and 100 ppm ZnO, where mean inhibitions were 67.24 and 63.39% for both nanoparticles, respectively (Table 1). On the other hand, *F. oxysporum* f. sp., *betae* was best inhibited (55.71%) by 100 ppm MgO, while *R. solani* and *S. rolfsii* were best inhibited with the use of TiO_2 (100 ppm) being 78.8 and 100% of inhibition for both fungi, respectively.

Effect on number of conidia and sclerotia: Data illustrated in Fig. 2 showed that, all the tested nanoparticles (all at 100 ppm) significantly decreased the number of sclerotia of *S. rolfsii* and number of conidia of *F. oxysporum* f. sp., *betae* compared to control. The MgO NP was the most effective and posed 69.23% for the percentage of inhibition of number of *F. oxysporum* f. sp., *betae* conidia. This was followed by TiO₂ (61.45%) and ZnO (54.81%). On the other hand, TiO₂ posed 100% for the percentage of inhibition of number of *sclerotia* of *S. rolfsii* followed by MgO (55.1%) while, ZnO showed the lowest inhibition effect being 49.29%.



Fig. 2(a-b): *In vitro* percentage of inhibition of (a) No. of conidia of *Fusarium oxysporum* f. sp., *betae* and (b) No. of sclerotia of *Sclerotium rolfsii* on PDA amended with 100 ppm of the different nanoparticles

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Fungi	Cont.	Nanoparticles (ppm)										
		 MgO		TiO ₂				ZnO				
		25	50	100	25	50	100	25	50	100		
Fusarium oxysporum f. sp., betae	0.00	42.86	50.41	55.71	31.23	49.59	53.67	32.04	44.29	53.06		
Sclerotium rolfsii	0.00	70.33	77.00	78.56	75.89	83.67	100.00	72.56	73.67	77.44		
Rhizoctonia solani	0.00	52.22	62.22	67.44	51.11	75.89	78.89	49.22	50.00	59.67		
Mean	0.00 ^d	55.14°	63.21 ^b	67.24ª	52.74°	69.72 ^b	77.52ª	51.27°	55.99 ^b	63.39ª		

Table 1: *In vitro* percentage of inhibition of radial growth of sugar beet root rot pathogenic fungi on PDA by different nanoparticle concentrations

Means followed by different letter(s) are significantly different at p = 0.05 of probability, inhibition effects were determined based on five replicates for each treatment, Cont.: Untreated control

Table 2: In vivo effect of nanoparticles on damping-off and root rot disease severity of sugar beet (cv., Kawamera) in pot experiments infested with the root rot fungi of sugar beet

Fungi	Treatments (%)														
	Pre-emergence					Post-emergence				Root rot severity					
	Cont.	Vitavax	MgO	TiO ₂	ZnO	Cont.	Vitavax	MgO	TiO ₂	ZnO	Cont.	Vitavax	MgO	TiO ₂	ZnO
Fusarium oxysporum f. sp., betae	33.30	9.24	12.90	18.47	20.33	22.14	6.13	2.22	6.81	9.36	18.06	0.00	0.00	2.78	2.78
Sclerotium rolfsii	40.70	11.10	16.60	11.1	16.63	21.67	6.25	6.66	4.17	6.68	34.7	1.39	2.78	0.0	4.17
Rhizoctonia solani	42.57	18.50	22.20	20.3	24.03	22.73	6.66	7.14	6.96	7.32	31.92	4.17	4.17	1.39	2.78
Mean	38.86ª	12.95°	17.23 ^b	16.6 ^{bc}	20.33 ^b	22.18 ^b	6.35 ^b	5.34 ^b	5.98 ^b	7.79 ^b	28.23ª	1.85 ^b	2.32 ^b	1.39 ^b	3.24 ^b

Pre-emergence and post-emergence damping-off were recorded 15 and 45 days after planting, respectively, while root rot severity was recorded 150 days after planting, all nanoparticles were applied as 100 ppm, while vitavax was applied as 3 g kg⁻¹

Effect on hyphal and spore morphology: In order to reveal the effect of the tested nanoparticles on hyphal and spore morphology, healthy fungal cultures on PDA of the sugar beet root rot fungi were sprayed with 100 ppm MgO, TiO₂ and ZnO solutions and other plates were also sprayed with water as control and investigated by Scanning Electron Microscope (SEM). The microscopic observation showed the images of conidia and mycelia in the control for *F. oxysporum* f. sp., *betae* with typical net structure and smooth surface (Fig. 3) while, MgO, TiO₂ and ZnO nanoparticles clearly damaged the hyphae and conidia of *F. oxysporum* f. sp., betae as fungal mycelia and conidia were sunken, wrinkled and damaged after 24 h (Fig. 3). On the other hand, Fig. 4 showed the image of *R. solani* and *S. rolfsii* mycelia of the control with typical net structure and smooth surface. In contrast MgO NPs formed unusual bulges on the surface of fungal hyphae, while TiO₂ caused severe hyphal distortion. The ZnO NPs, however, caused deformation and lysis for fungal hyphae of both fungi (Fig. 4).

In vivo effect of nanoparticles on root rot and damping-off

on sugar beet: In pot experiment the effect of MgO, TiO₂ and ZnO NPs and vitavax on damping-off and root rot of sugar beet (cv., Kawamera) caused by *F. oxysporum* f. sp., *betae*, *S. rolfsii* and *R. solani* is shown in Table 2. Treatment with TiO₂ was the most effective and decreased mean percentage of pre-emergence damping-off to 16.60% compared to 38.86% for the untreated in tested control and this was not

significantly different form the vitavax effect and also was not significantly different form MgO and ZnO NPs effect. On the other hand, all the tested NPs significantly decreased post emergence damping-off and MgO was most effective as decreased post emergence to 5.34% compared to 22.18% for the untreated control. Meanwhile, this effect was not significantly different form the vitavax fungicide effect and the other two NPs, TiO₂ and ZnO (Table 2). Concerning to the root rot severity, all NPs significantly deceased it where TiO₂ was the most effective and decreased disease severity to 1.39% compared to 28.2% for the untreated control. Meantime, TiO₂ effect was not significantly different vitavax and also the MgO and ZnO effect (Table 2).

Effect of nanoparticles on the vegetative and the chemical characteristics of sugar beet

Root fresh (biomass) and dry weight: Data in Table 3 showed the effect of MgO, TiO₂ and ZnO NPs on some agronomic parameters of sugar beet plants grown in soil infested with root rots fungi. Treatment with TiO₂ proved to be the best treatment as significantly increased the mean root fresh weight (biomass) to 193.4 g plant⁻¹ compared to 104.5 g for the untreated infected control. This effect was even significantly higher than vitavax which showed 175.5 g mean root fresh weight and was higher than MgO (167.5 g) and ZnO (151.5 g).

A similar trend was obtained for the dry weight of roots where TiO_2 showed the highest effect and increased dry



Fig. 3(a-d): Scanning Electron Microscopy (SEM) of conidia and hyphae of *F. oxysporum* f. sp., *betae* grown on PDA sprayed with either water as a (a) Control or 100 ppm of (b) MgO, (c) TiO₂ and (d) ZnO nanoparticle solutions, 24 h after treatment

Table 3: In vivo effect of nanoparticles on root fresh weight (biomass) and dry weight of sugar beet grown in soil infested with sugar beet root rot fungi in pot experiment, 150 days after planting

Fungi	Treatments (g plant ⁻¹)												
	Root fres	h weight				Root dry weight							
	Cont.	Vitavax	MgO	TiO ₂	ZnO	Cont.	Vitavax	MgO	TiO ₂	ZnO			
Fusarium oxysporum f. sp., betae	108.02	192.65	217.25	177.05	161.55	29.98	48.69	51.03	47.58	46.32			
Sclerotium rolfsii	105.32	170.1	142.8	207.26	139.45	28.77	45.27	45.77	50.28	45.73			
Rhizoctonia solani	100.22	163.95	143.28	195.85	153.7	27.5	46.07	44.87	48.82	45.81			
Mean	104.52 ^d	175.57 ^b	167.78 ^{bc}	193.4ª	151.57°	28.75 ^d	46.68 ^{abc}	47.22 ^{ab}	48.89ª	45.95 ^{abc}			



Fig. 4(a-d): Scanning Electron Microscopy (SEM) of *R. solani* and *S. rolfsii* hyphae grown on PDA sprayed with either water as a (a) Control or 100 ppm of (b) MgO, (c) TiO₂ and (d) ZnO nanoparticle solutions, 24 h after treatment

weight of roots to 48.8 g plant⁻¹ compared to 28.7 g for the untreated infected control. Meanwhile, TiO₂ effect was not significantly different from the other NPs treatments and the vitavax (Table 3).

Total Soluble Solids (TSS) and sucrose: For TSS and sucrose content, data of Table 4 revealed a similar trend for both parameters. All the NPs tested and vitavax increased TSS (%) and sucrose (%) compared to the untreated infected control.

However, TiO_2 showed the highest effect and increased TSS to 22.9% and sucrose to 18.3% compared to 16.7 and 13.8% for both parameter, respectively in the untreated control (Table 4).

Total phenols: Data of Table 5 revealed that all the nanoparticles tested increased the defensive reaction related to total phenols compared to the untreated infected control. However, TiO_2 was of the highest effect and increased mean

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150 days arter planting													
Fungi	Treatments (%)												
	TSS					Sucrose							
	Cont.	Vitavax	MgO	TiO ₂	ZnO	Cont.	Vitavax	MgO	TiO ₂	ZnO			
Fusarium oxysporum f. sp., betae	17.17	24	24	23.67	20.33	13.97	18.78	18.63	18.56	17.25			
Sclerotium rolfsii	16	22	21.67	23	21.67	13.43	17.52	17.91	18.48	17.51			
Rhizoctonia solani	17	21	21.00	22	21.33	13.96	17.14	17.34	17.87	17.37			
Mean	16.72 ^c	22.33ª	22.22 ^{ab}	22.89ª	21.1 ^b	13.79 ^d	17.8 ^{abc}	17.96 ^{ab}	18.31ª	17.38 ^{bc}			

Table 4: In vivo effect of nanoparticles on TSS and sucrose content of sugar beet roots grown in soil infested with sugar beet root rot fungi in pot experiments, 150 days after planting

Table 5: Effect of MgO, TiO₂ and ZnO on total phenol content and activity of polyphenol oxidase of sugar beet roots grown in soil infested with sugar beet root rots fungi in pot experiments, 150 days after planting

Fungi	Treatments											
	Total phe	nols (mg g $^{-1}$ fre	esh tissue)			Polyphenol oxidase activity (Absorbance min g^{-1} fresh tissue						
	Cont.	Vitavax	MgO	TiO ₂	ZnO	Cont.	Vitavax	MgO	TiO ₂	ZnO		
Fusarium oxysporum f. sp., betae	4.60	8.83	9.58	7.87	7.83	0.025	0.059	0.058	0.056	0.051		
Sclerotium rolfsii	3.83	8.75	8.10	9.41	7.75	0.026	0.062	0.059	0.084	0.052		
Rhizoctonia solani	4.73	8.49	7.45	9.26	8.68	0.030	0.059	0.057	0.112	0.073		
Mean	4.39 ^d	8.69 ^{ab}	8.38 ^{bc}	8.85ª	8.09 ^c	0.027 ^d	0.060 ^b	0.058°	0.084ª	0.059 ^{bc}		

of the total phenols to 8.85 mg g^{-1} fresh weight compared to 4.39 in the untreated control (Table 5). Meanwhile, this effect was not significantly different from MgO nanoparticle and fungicide vitavax.

Polyphenol oxidase activity: Data of Table 5 revealed that all the nanoparticles tested increased of the defensive reaction related to enzyme activity (polyphenol oxidase) compared to the untreated infected control. The TiO₂ nanoparticles showed highest effect and increased mean of the polyphenol oxidase activity to 0.084 (absorbance min g⁻¹ fresh tissue) compared to 0.027 in the untreated control (Table 5). The effect was significantly higher than vitavax (0.060), ZnO (0.059) and MgO (0.058).

DISCUSSION

Control of plant diseases is the most challenging aspect in crop production. In recent years, resistance to commercially available fungicides by phytopathogenic fungi has been increasing and has become a serious problem³³.

A greater effort has been given for the development of safe management methods that pose less danger to humans and animals. Thus, use of nanoparticles has been suggested as an alternative and effective approach with antifungal potential which is eco-friendly and cost effective for the control of pathogenic microbes^{7,34-37}.

In the present study, all the tested nanoparticles i.e., magnesium oxide (MgO), zinc oxid (ZnO) and titanium dioxide (TiO₂) at the tested concentrations (25, 50 and 100 ppm)

significantly inhibited the in vitro fungal radial growth, sporulation and sclerotia formation of the tested damping-off and root rot fungi of sugar beet, Fusarium oxysporum f. sp., betae, Sclerotium rolfsii and Rhizoctonia solani on PDA compared to control (sterile deionized water). The TiO₂ at 100 ppm was of the highest effect as inhibited mean radial growth of the tested fungi by 77.52%, followed by MgO with 67.2% inhibition. However, MgO at 100 ppm was most effective and decreased number of conidia of Fusarium oxysporum f. sp., betae by 69.33% which was not significantly different than TiO₂ which decreased number of conidia by 61.54%. However, TiO₂ at 100 ppm was most effective to inhibit sclerotia formation of Sclerotium rolfsii by 100%, while MgO was the second in this respect being 55.1%. The ZnO, however, showed the least effect compared to the other two NPs but still significantly effective to decrease radial growth, sporulation and sclerotia formation compared to the control. These effects were accompanied with deformation and lysis of the fungal hyphae and spores. The results were in harmony with the findings of Sharma et al.³⁸ as reported that ZnO NPs shown antifungal activity against Pythium debarynum and Sclerotium rolfsii, causing a significant decrease in the fungal growth that corresponds to increase in the concentration of ZnO NPs. Besides, Kumar et al.³⁹ reported that Ag NPs treated plates containing *S. rolfsii* showed sclerotia formation either lacking or abnormal, if formed. There result were in contrast with the findings of Kasprowicz et al.40 who reported that the number of Fusarium culmorum spores formed by mycelia increased in the culture after contact with silver nanoparticles (Ag NPs), especially on the nutrient-poor PDA medium but in agreement with the findings of Al-Othman *et al.*⁴¹ who reported that Ag NPs reduced *A. flavus* spores number. Also, Yehia and Ahmed⁴² mentioned that ZnO NPs may disrupt and damage the conidia of *F. oxysporum* and deformate the structure of fungal hyphae and consequently the growth was deeply inhibited. The generation of active free hydroxyl radicals (-OH) by photoexcited TiO₂ particles is probably responsible for the antibacterial activity⁴³.

Meanwhile, the inhibitory effect of nanoparticles may be due to release of extracellular enzymes and metabolites⁴⁴. Also, some studies proposed that ZnO NPs may cause structural changes of microbial cell membrane, causing cytoplasm leakage and eventually the death of bacterial cells^{45,46}.

The *in vivo* pot experiment conducted supported the *in vitro* results, treatment with TiO_2 was the most effective and decreased percentage of damping-off to 16.60% compared to 38.8% for the untreated control. This TiO_2 effect was not significantly different from the vitavax fungicide effect. For the post-emergence damping-off, MgO was most effective and reduced post-emergence to 5.3% compared to 22.1% for the untreated control. This effect was not significantly different from TiO_2 and the fungicide vitavax where post-emergence damping-off was 5.9 and 6.3%, respectively. All the tested NPs significantly decreased root rot severity to 1.39% compared to 28.2% for the untreated control and was even not significantly different form the fungicide vitavax effect.

These findings were in agreement with Elamawi and Al-Harbi⁴⁷ who reported that *Fusarium* disease incidence on tomato was reduced by silver nanoparticles to 5% compared to 100% for the untreated control also, Jo *et al.*⁴⁸ found that silver nanoparticles, reduced the disease severity by *Bipolaris sorokiniana* which causes seedling blight, root rot, crown rot and leaf spot blotch on various gramineous species and *Magnaporthe grisea* which causes blast on rice. Servin *et al.*⁴⁹ showed that ZnO, TiO₂ and CuO nanoparticles may have significant use in pathogen control program by directly inhibiting disease causing organisms or by affecting the systemic acquired resistance.

On the other hand, the present study showed that MgO, TiO₂ and ZnO NPs significantly enhanced agronomic parameters of sugar beet plants grown in soil infested with root rot fungi of sugar beet. Treatment with TiO₂ proved to be the best treatment as increased the mean root fresh weight (biomass) to 193.4 g compared to 104.5 g for the untreated infected control. This effect was even significantly higher than the fungicide vitavax which showed 175.5 g mean root fresh weight and was higher than MgO (167.5 g) and ZnO (151.5 g). A similar trend was obtained for the dry weight of roots. Their findings were in harmony with that of Mahajan *et al.*⁵⁰ as mentioned that seedling roots of *Vigna radiate* and *Cicer arietinum* absorbed ZnO NPs and promoted the root and shoot length and root and shoot biomass. Also, De la Rosa *et al.*⁵¹ applied different concentrations of ZnO NPs on cucumber and found that cucumber seed germination was enhanced. Mahmoodzadeh *et al.*⁵² reported that TiO₂ NPs enhanced seed germination, promoted radicle and plumule growth of canola seedlings. Also, Jaberzadeh *et al.*⁵³ reported that TiO₂ augmented wheat plant growth and yielded components under water deficit stress condition.

Meanwhile, the present study also showed that the NPs tested increased TSS and sucrose (%) of the roots compared to the untreated infected control where TiO₂ was the most effective and increased TSS and sucrose to 22.9 and 18.3% compared to 16.7 and 13.7% for both parameters, respectively and this was not significantly different from the vitavax effect. Application of TiO₂ NPs was found to improve plant-photosynthesis efficacy, plant-enzyme activity and provide plants with more nitrogen nutrient by chemical fixation of nitrogen in the air⁵⁴⁻⁵⁷. Also, all the NPs tested in the present study and vitavax enhanced the plant defense and related the enzyme activity i.e., polyphenol oxidase and the total phenols compared to the untreated infected control. Phenol is one of the most stress-responsive plant compounds according to Rodrigues et al.57. In general, the toxicity of NPs is determined by their particle size, shape and biodegradability⁵⁸⁻⁶⁰.

There are five theories, which have been proposed about the mechanisms of nano-metal toxicity: (1) Release of toxic ions (Cd²⁺, Zn²⁺, Ag⁺) that can bind to sulphur-containing proteins, this accumulation prevents the proteins from properly functioning in the membrane and interfere in cell permeability, (2) They can be genotoxic_ions that can destroy DNA which leads to cell death, (3) Interruption of electron transport, protein oxidation and membrane potential collapse due to its contact with CeO₂ or nC60, (4) Generation of Reactive Oxygen Species (ROS) mediated cellular damage and different metal-catalyzed oxidation reactions could underlie specific types of protein, membrane or DNA damage and (5) Interference with nutrient uptake. These mechanisms may not operate separately suggesting that more than one mechanism can occur simultaneously^{61,62}.

CONCLUSION

In vitro and in planta experiments clearly showed that the magnesium oxide, titanium dioxide and zinc oxide

nanoparticles (MgO, TiO₂ and ZnO NPs) increased the *in vitro* fungal growth inhibition. Meanwhile, TiO₂ NP (100 ppm) showed the highest inhibition effect of fungal radial growth and the formation of Sclerotium rolfsii, while MgO was most effective in decreasing the sporulation (number of conidia) of Fusariun oxysporum f. sp., betae. Application of all tested NPs significantly decreased the developed root rot severity and TiO₂ was most effective. On the other hand, treatments with the tested NPs significantly increased both root fresh weight (biomass) and dry weight of the treated plants. Also, the treated plants showed increased levels of sucrose, Total Soluble Solids (TSS), phenol content and activity of the defense related enzyme, polyphenol oxidase. The use of nanoparticles as an environmentally safe substitute of the traditional synthetic fungicide will provide a green method of controlling plant diseases. More attention should be given to develop strong delivery method of nanoparticles compounds in different agriculture systems.

SIGNIFICANCE STATEMENTS

Yearly average production of sugar beets in Egypt is about 10 million tons this makes it the world number 9 producer of sugar beet and sugar industry is almost relying on sugar beet plants. Our data presented in the study will help sugar beet growers to control the damping-off and root rot disease in a safe way that will reduce the risk of using harmful synthetic fungicides and to maintain more eco-friendly sugar beet cultivation system.

REFERENCES

- Jafarnia, B., R. Ghorbani, A.Z. Feizabady and A.R. Ghaemi, 2013. Impact of crop density and soil fertilization on sugar beet. Int. J. Agric. Crop Sci., 5: 2991-2999.
- 2. Koocheki, A. and A. Soltani, 1996. The Sugar Beet Crop. Jihad of Mashhad Publication, Iran.
- 3. Koocheki, A., 1994. Agriculture and Energy (An Ecological View). Ferdowsi University Press, Tehran, Iran.
- 4. Esfahani, M.N., 2006. Present status of *Fusarium* dry rot of potato tuber in Isfhan (Iran). Indian Phytopathol., 59: 142-147.
- Lamsal, K., S.W. Kim, J.H. Jung, Y.S. Kim, K.S. Kim and Y.S. Lee, 2011. Inhibition effects of silver nanoparticles against powdery mildews on cucumber and pumpkin. Mycobiology, 39: 26-32.
- Kim, S.W., J.H. Jung, K. Lamsal, Y.S. Kim, J.S. Min and Y.S. Lee, 2012. Antifungal effects of silver nanoparticles (AgNPs) against various plant pathogenic fungi. Mycobiology, 40: 53-58.

- Kumar, V. and S.K. Yadav, 2009. Plant-mediated synthesis of silver and gold nanoparticles and their applications. J. Chem. Technol. Biotechnol., 84: 151-157.
- 8. Morones, J.R., J.L. Elechiguerra, A. Camacho, K. Holt, J.B. Kouri, J.T. Ramirez and M.J. Yacaman, 2005. The bactericidal effect of silver nanoparticles. Nanotechnology, 16: 2346-2353.
- 9. Martinez-Gutierrez, F., P.L. Olive, A. Banuelos, E. Orrantia and N. Nino *et al.*, 2010. Synthesis, characterization and evaluation of antimicrobial and cytotoxic effect of silver and titanium nanoparticles. Nanomed.: Nanotechnol. Biol. Med., 6:681-688.
- 10. Kim, K.J., W.S. Sung, S.K. Moon, J.S. Choi, J.G. Kim and D.G. Lee, 2008. Antifungal effect of silver nanoparticles on dermatophytes. J. Microbiol. Biotechnol., 18: 1482-1484.
- 11. Kim, K.J., W.S. Sung, B.K. Suh, S.K. Moon, J.S. Choi, J.G. Kim and D.G. Lee, 2009. Antifungal activity and mode of action of silver Nano-particles on *Candida albicans*. BioMetals, 22: 235-242.
- 12. Kumar, A., P.K. Vemula, P.M. Ajayan and G. John, 2008. Silver-nanoparticle-embedded antimicrobial paints based on vegetable oil. Nat. Mater., 7: 236-241.
- 13. Cioffi, N., L. Torsi, N. Ditaranto, G. Tantillo and L. Ghibelli *et al.*, 2005. Copper nanoparticle/polymer composites with antifungal and bacteriostatic properties. Chem. Mater., 17: 5255-5262.
- Kwak, S.Y., S.H. Kim and S.S. Kim, 2001. Hybrid organic/inorganic Reverse Osmosis (RO) membrane for bactericidal anti-fouling. 1. preparation and characterization of TiO₂ nanoparticle self-assembled aromatic polyamide Thin-Film-Composite (TFC) membrane. Environ. Sci. Technol., 35: 2388-2394.
- 15. Liu, Y., L. He, A. Mustapha, H. Li, Z.Q. Hu and M. Lin, 2009. Antibacterial activities of zinc oxide nanoparticles against *Escherichia coli* O157:H7. J. Applied Microbiol., 107: 1193-1201.
- Khodakovsky, A., P. Schroder and W. Sweldens, 2000. Progressive geometry compression. Proceedings of the 27th Annual Conference on Computer Graphics and Interactive Techniques, July 23-28, 2000, New Orleans, LA., USA., pp: 271-278.
- Suriyaprabha, R., G. Karunakaran, R. Yuvakkumar, P. Prabu, V. Rajendran and N. Kannan, 2012. Growth and physiological responses of maize (*Zea mays* L.) to porous silica nanoparticles in soil. J. Nanopart. Res., 14: 1294-1296.
- 18. He, L., Y. Liu, A. Mustapha and M. Lin, 2011. Antifungal activity of zinc oxide nanoparticles against *Botrytis cinerea* and *Penicillium expansum*. Microbiol. Res., 166: 207-215.
- 19. Wani, A.H., M. Amin, M. Shahnaz and M.A. Shah, 2012. Antimycotic activity of nanoparticles of Mgo, FeO and ZnO on some pathogenic fungi. Int. J. Manuf. Mater. Mech. Eng., 2: 59-70.
- Min, J.S., K.S. Kim, S.W. Kim, J.H. Jung and K. Lamsal *et al.*, 2009. Effects of colloidal silver nanoparticles on Sclerotium-forming phytopathogenic fungi. Plant Pathol J., 25: 376-380.

- Fraternale, D., L. Giamperi and D. Ricci, 2003. Chemical composition and antifungal activity of essential oil obtained from *in vitro* plants of *Thymus mastichina* L. J. Essent. Oil Res., 15: 278-281.
- 22. Kaur, P., R. Thakur and A. Choudhary, 2012. An *in vitro* study of the antifungal activity of silver/chitosan nanoformulations against important seed borne pathogens. Int. J. Sci. Technol. Res., 1: 83-86.
- 23. Elamawi, R.M.A. and R.A.S. El-Shafey, 2013. Inhibition effects of silver nanoparticles against rice blast disease caused by *Magnaporthe grisea*. Egypt. J. Agric. Res., 91: 1271-1283.
- 24. Papavizas, G.C. and C.B. Davey, 1962. Isolation and pathogenicity of Rhizoctonia saprophytically existing in soil. J. Phytopathol., 52: 834-840.
- 25. Marwa, A.M., S. Atwa, T. Shehata and M.M.H. Rahhal, 2014. Induction of resistance against soybean damping-off caused by *Rhizoctonia solani*. Egypt. J. Phytopathol., 42: 137-158.
- El-Shafey, H.A., F.A. El-Shorbagy, I.I. Khalil and E.M. El-Assiuty, 1988. Additional sources of resistance to the late-wilt disease of maize caused by *Cephalosporium maydis*. Agric. Res. Rev., 66: 221-230.
- 27. Rowe, R.C., 1980. Comparative pathogenicity and host ranges of *Fusarium oxysporum* isolates causing crown and root rot of greenhouse and field-grown tomatoes in North America and Japan. Phytopathology, 70: 1143-1148.
- 28. Liu, L., J.W. Kloepper and S. Tuzun, 1995. Induction of systemic resistance in cucumber against bacterial angular leaf spot by plant growth-promoting rhizobacteria. Phytopathology, 85: 843-847.
- 29. Carruthers, A. and J.F.T. Oldfield, 1961. Methods for the assessment of beet quality. Int. Sugar J., 63: 72-74.
- 30. Fatouh, H.M.M., 2012. Pathological studies on sugar beet rot. M.Sc. Thesis, Banha University, Egypt.
- 31. Zieslin, N. and R. Ben-Zaken, 1993. Peroxidase activity and presence of phenolic substances in peduncles of rose flowers. Plant Physiol. Biochem., 31: 333-339.
- 32. Maxwell, D.P. and D.F. Bateman, 1967. Changes in the activity of some oxidases in extracts of *Rhizoctonia* infected bean hypocotyls in relation to lesion maturation. Phytopathology, 57: 132-136.
- Goffeau, A., 2008. Drug resistance: The fight against fungi. Nature, 452: 541-542.
- 34. Gupta, A.K. and M. Gupta, 2005. Synthesis and surface engineering of iron oxide nanoparticles for biomedical applications. Biomaterials, 26: 3995-4021.
- Long, S.P., E.A. Ainsworth, A.D.B. Leakey, J. Nosberger and D.R. Ort, 2006. Food for thought: Lower-than-expected crop yield stimulation with rising CO₂ concentrations. Science, 312: 1918-1921.
- Magrez, A., S. Kasas, V. Salicio, N. Pasquier and J.W. Seo *et al.*, 2006. Cellular toxicity of carbon-based nanomaterials. Nano Lett., 6: 1121-1125.

- 37. Nel, A.E., T. Xia, L. Madler and N. Li, 2006. Toxic potential of materials at the nanolevel. Science, 311: 622-627.
- Sharma, D., S. Sharma, B.S. Kaith, J. Rajput and M. Kaur, 2011. Synthesis of ZnO nanoparticles using surfactant free in-air and microwave method. Applied Surf. Sci., 257: 9661-9672.
- Kumar, G.D., N. Natarajan and S. Nakkeeran, 2015. Synthesis and characterization of silver (Ag) nanoparticles and its antifungal activity against *Sclerotium rolfsii* in chilli (*Capsicum annum* L.). Int. J. Agric. Sci. Res., 5: 211-218.
- 40. Kasprowicz, M.J., M. Koziol and A. Gorczyca, 2010. The effect of silver nanoparticles on phytopathogenic spores of *Fusarium culmorum*. Can. J. Microbiol., 56: 247-253.
- Al-Othman, M.R., A.R.M. Abd El-Aziz, M.A. Mahmoud, S.A. Eifan, M.M. El-Shikh and M. Majrashi, 2014. Application of silver nanoparticles as antifungal and antiaflatoxin B1 produced by *Aspergillus flavus*. Dig. J. Nanomater. Biostruct., 9: 151-157.
- Yehia, R.S. and O.F. Ahmed, 2013. *In vitro* study of the antifungal efficacy of zinc oxide nanoparticles against *Fusarium oxysporum* and *Penicilium expansum*. Afr. J. Microbiol. Res., 7: 1917-1923.
- Wei, C., W.Y. Lin, Z. Zainal, N.E. Williams and K. Zhu *et al.*, 1994. Bactericidal activity of TiO₂ photocatalyst in aqueous media: Toward a solar-assisted water disinfection system. Environ. Sci. Technol., 28: 934-938.
- 44. Perez-de-Luque, A. and D. Rubiales, 2009. Nanotechnology for parasitic plant control. Pest Manage. Sci., 65: 540-545.
- 45. Sawai, J. and T. Yoshikawa, 2004. Quantitative evaluation of antifungal activity of metallic oxide powders (MgO, CaO and ZnO) by an indirect conductimetric assay. J. Applied Microbiol., 96: 803-809.
- 46. Brayner, R., R. Ferrari-Iliou, N. Brivois, S. Djediat, M.F. Benedetti and F. Fievet, 2006. Toxicological impact studies based on *Escherichia coli* bacteria in ultrafine ZnO nanoparticles colloidal medium. Nano Lett., 6: 866-870.
- Elamawi, R.M. and R.E. Al-Harbi, 2014. Effect of biosynthesized silver nanoparticles on *Fusarium oxysporum* fungus the cause of seed rot disease of faba bean, tomato and barley. J. Plant Protect. Pathol. Mansoura Univ., 5: 225-237.
- Jo, Y.K., B.H. Kim and G. Jung, 2009. Antifungal activity of silverions and nanoparticles on phytopathogenic fungi. Plant Dis., 93: 1037-1043.
- Servin, A., W. Elmer, A. Mukherjee, R. de la Torre-Roche and H. Hamdi *et al.*, 2015. A review of the use of engineered nanomaterials to suppress plant disease and enhance crop yield. J. Nanopart. Res., Vol. 17. 10.1007/s11051-015-2907-7.
- Mahajan, P., S.K. Dhoke and A.S. Khanna, 2011. Effect of nano-ZnO particle suspension on growth of mung (*Vigna radiate*) and gram (*Cicer arietinum*) seedlings using plant agar method. J. Nanotechnol. 10.1155/2011/696535.

- 51. De la Rosa, G., M.L. Lopez-Moreno, D. de Haro, C.E. Botez, J.R. Peralta-Videa and J.L. Gardea-Torresdey, 2013. Effects of ZnO nanoparticles in alfalfa, tomato and cucumber at the germination stage: Root development and X-ray absorption spectroscopy studies. Pure Applied Chem., 85: 2161-2174.
- 52. Mahmoodzadeh, H., M. Nabavi and H. Kashefi, 2013. Effect of nanoscale titanium dioxide particles on the germination and growth of canola (*Brassica napus*). J. Ornamental Hortic. Plants, 3: 25-32.
- 53. Jaberzadeh, A., P. Moaveni, H.R.T. Moghadam and H. Zahedi, 2013. Influence of bulk and nanoparticles titanium foliar application on some agronomic traits, seed gluten and starch contents of wheat subjected to water deficit stress. Notulae Botanicae Horti Agrobotanici Cluj-Napoca, 41: 201-207.
- Yang, F., F. Hong, W. You, C. Liu, F. Gao, C. Wu and P. Yang, 2006. Influence of nano-anatase TiO₂ on the nitrogen metabolism of growing spinach. Biol. Trace Elem. Res., 110: 179-190.
- 55. Yang, F., C. Liu, F. Gao, M. Su and X. Wu *et al.*, 2007. The improvement of spinach growth by nano-anatase TiO₂ treatment is related to nitrogen photoreduction. Biol. Trace Elem. Res., 119: 77-88.
- Liu, R. and R. Lal, 2015. Potentials of engineered nanoparticles as fertilizers for increasing agronomic productions. Sci. Total Environ., 514: 131-139.

- Rodrigues, F.A., F.X.R. Vale, G.H. Korndorfer, A.S. Prabhu, L.E. Datnoff, A.M.A Oliveira and L. Zambolim, 2003. Influence of silicon on sheath blight of rice in Brazil. Crop Protect., 22: 23-29.
- 58. Das, S.K., A.R. Das and A.K. Guha, 2009. Gold nanoparticles: Microbial synthesis and application in water hygiene management. Langmuir, 25: 8192-8199.
- Keck, C.M. and R.H. Muller, 2013. Nanotoxicological Classification System (NCS)-A guide for the risk-benefit assessment of nanoparticulate drug delivery systems. Eur. J. Pharmaceut. Biopharmaceut., 84: 445-448.
- Ahamed, M., M.J. Akhtar, H.A. Alhadlaq, M.A.M. Khan and S.A. Alrokayan, 2015. Comparative cytotoxic response of nickel ferrite nanoparticles in human liver HepG2 and breast MFC-7 cancer cells. Chemosphere, 135: 278-288.
- Zeng, F., C. Hou, S.Z. Wu, X.X. Liu, Z. Tong and S.N. Yu, 2007. Silver nanoparticles directly formed on natural macroporous matrix and their anti-microbial activities. Nanotechnology, Vol. 18, No. 5. 10.1088/0957-4484/18/5/055605
- 62. Lemire, J.A., J.J. Harrison and R.J. Turner, 2013. Antimicrobial activity of metals: Mechanisms, molecular targets and applications. Nat. Rev. Microbiol., 11: 371-384.