

## Endothelin-1-Induced Endoplasmic Reticulum Stress in Parkinson's Disease

<sup>1</sup>Arjun Jain, <sup>2</sup>Anna Migdalska-Richards and <sup>3</sup>Ashok Jain

<sup>1</sup>Department of Physiology, Development and Neuroscience, University of Cambridge, Cambridge, United Kingdom

<sup>2</sup>Department of Clinical Neurosciences, University College London Institute of Neurology, London, United Kingdom

<sup>3</sup>Hansraj College, Delhi University, Delhi, India

### ABSTRACT

**Background:** ET-1 is known to be implicated in various neurodegenerative diseases, including multiple sclerosis and Alzheimer's disease. However, till date, there has been little, if any, investigation into its possible involvement in Parkinson's Disease (PD). **Results:** Recent findings implicate ET-1 in the induction of Endoplasmic Reticulum (ER) stress in disease. ER stress has also been demonstrated to be an important factor in PD pathology and recent findings also indicate increased ET-1 levels in PD. **Conclusion:** The following review attempts to highlight the potential link between ET-1-induced ER stress and PD pathology.

**Key words:** Endothelin-1, endoplasmic reticulum stress, Parkinson's disease

Pharmacologia 5 (3): 84-90, 2014

### INTRODUCTION

ET-1 is a vasoactive peptide synthesized and secreted by a diverse range of cells. It acts via binding two G-protein coupled receptors, the endothelin A (ETA) and endothelin B (ETB) receptors, to initiate signaling events in a wide variety of target tissues. This includes key physiologic functions, such as maintenance of vascular tone, tissue differentiation and cell proliferation (Nelson *et al.*, 2003), as well as induction of pathological processes (under elevated, pathological levels of ET-1), which includes hypertension, inflammation and oxidative stress (Dong *et al.*, 2005; Fiore *et al.*, 2005; Kramer *et al.*, 1994; Simonson and Ismail-Beigi, 2011). A recent study by Jain *et al.* (2012) has demonstrated that ET-1 also induces Endoplasmic Reticulum (ER) stress (Jain *et al.*, 2012). This was shown in the context of pregnancy disorders, where ET-1 was found to induce placental ER stress by activating the PLC-IP<sub>3</sub> pathway. The broader implications of ET-1 induced ER stress and its role in pathology have since been reviewed (De Miguel and Pollock, 2013; Jain, 2013).

Previous studies have explored the link between ER stress and neurodegeneration. Disruption of ER functioning is associated with the accumulation of misfolded proteins, which are a characteristic occurrence

in many neurodegenerative diseases, including Parkinson's Disease (PD) (Gorman, 2008; Soto and Estrada, 2008; Winklhofer *et al.*, 2008). Furthermore, recent studies demonstrate increased ET-1 levels in PD (Calderon-Garciduenas *et al.*, 2013; Makarov *et al.*, 2013), which implicates ET-1 (and associated ER stress?) in the pathology of PD. The following sections review the evidence of ER stress and associated pathology in PD, as well as the potential contribution of ET-1 to PD pathology. It is hoped that this will attract future research on this topic, which might lead to new avenues of therapeutic intervention in PD. We first begin with an explanation of the role of the ER and associated ER stress pathways.

### ENDOPLASMIC RETICULUM STRESS

In addition to being involved in the synthesis, transport and modification of membrane and secretory proteins, the ER also serves as a reservoir of calcium ions (Ca<sup>2+</sup>) (Zhang and Kaufman, 2008). In the ER lumen, Ca<sup>2+</sup> is buffered by calcium-binding proteins. Many of these proteins also serve as molecular chaperones involved in folding and quality control of ER proteins (Michalak *et al.*, 1998), and their functional activity alters with changes in Ca<sup>2+</sup> concentration. Therefore, loss of ER Ca<sup>2+</sup> homeostasis leads to suppression of post-translational modifications of proteins in the ER, which leads to an accumulation of misfolded or unfolded

**Corresponding Author:** Arjun Jain, 42 Bumplizstrasse, 3027 Bern, Switzerland Tel: +41 0786796748

proteins. This induces activation of ER stress response pathways known collectively as the Unfolded Protein Response (UPR) (Brostrom and Brostrom, 2003). This is a multifaceted response, which includes suppression of non-essential protein synthesis via the PERK pathway, upregulation of ER chaperone proteins and folding enzymes, via the ATF6 (activating transcription factor 6) and IRE1 $\alpha$  (inositol-requiring 1 $\alpha$ ) pathways and activation of ER-associated protein degradation (ERAD) machinery in order to remove any remaining unfolded or misfolded proteins. Collectively, these pathways help restore ER homeostasis. However, if this fails, then apoptosis is induced in order to eliminate the stressed cell. Malfunction of ER stress responses due to aging, genetic mutations or environmental factors can result in various diseases such as diabetes, inflammation and neurodegenerative disorders (Yoshida, 2007).

### ENDOPLASMIC RETICULUM STRESS IN PARKINSON'S DISEASE

PD is a progressive neurodegenerative disease characterized by a loss of dopaminergic neurons in the Substantia Nigra Pars Compacta (SNPC) and the presence of intracytoplasmic inclusions known as Lewy bodies (Thomas and Beal, 2007). Within the Lewy bodies, diffuse deposits of misfolded  $\alpha$ -synuclein form the core in association with other proteins, including components of the ubiquitin-proteasome system (Gorman, 2008). Neurotoxins, such as 6-hydroxydopamine (6-OHDA) and N-methyl-4-

phenyl-1, 2,3,6-tetra-hydroxyridine are used as model compounds to mimic the disease process. Recent studies with cultured neuronal cells, including dopaminergic neurons, showed that these compounds trigger ER stress and induce a number of UPR target genes (Holtz and O'Malley, 2003; Ryu *et al.*, 2002). Gene profiling revealed that both ER chaperones (GRP78 and GRP94) and other components of the UPR, such as the transcription factor, CHOP/Gadd153 were upregulated in exposed cells, in addition to increased phosphorylation of the ER stress kinases, IRE1 $\alpha$  and PERK. Furthermore, neurons from PERK $^{-/-}$ embryos display increased sensitivity to 6-OHDA (Ryu *et al.*, 2002) suggesting that at least some neurotoxins of relevance to PD kill dopaminergic neurons through mechanisms modulated by or involving ER stress and UPR signalling components (Fig. 1).

Hereditary mutations in the ER-associated E3 ubiquitin ligase, Parkin, have also been associated with ER-stress-induced cell death and are found in patients with juvenile-onset PD (Dawson and Dawson, 2003; Takahashi *et al.*, 2003). Overexpression of wild-type Parkin suppresses ER stress-mediated cell death induced by  $\alpha$ -synuclein, the principal component of Lewy bodies that represent the hallmark lesions of PD (Imai *et al.*, 2000; Petrucelli *et al.*, 2002; Takahashi *et al.*, 2003).  $\alpha$ -synuclein is normally enriched in nerve terminals and is involved in synaptic function. There is an increase in the levels of natively folded  $\alpha$ -synuclein in the cytoplasm of substantia nigra neurons during normal aging and in PD (Chu and Kordower, 2007). In addition,

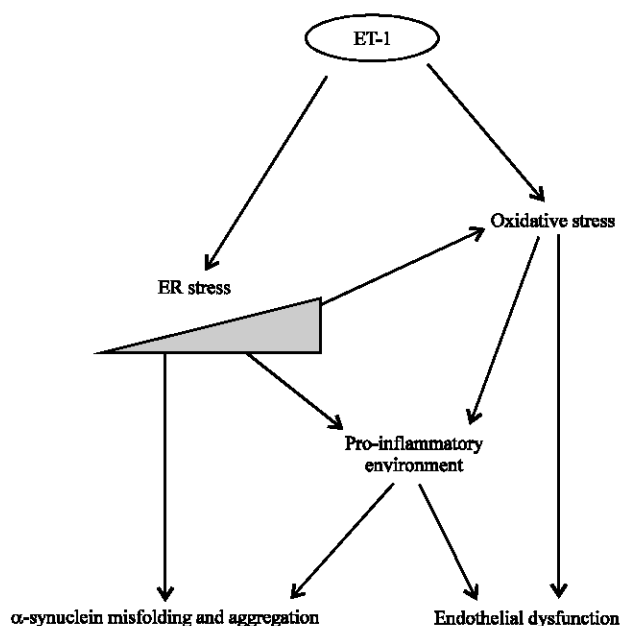


Fig. 1: ET-1 induced biochemical pathways of relevance to PD pathology

$\alpha$ -synuclein in Lewy bodies of PD is misfolded and aggregates accumulate (Uversky, 2007). The precise cause of the misfolding of  $\alpha$ -synuclein is unknown but various factors are associated with aggregation of  $\alpha$ -synuclein (Uversky, 2007). Indeed, there is also evidence of activation of the UPR with aggregation of  $\alpha$ -synuclein. This includes increased expression of GRP78 and CHOP, as well as increased p-eIF2 $\alpha$  levels (Doyle *et al.*, 2011; Smith *et al.*, 2005). ET-1-induced loss of ER Ca<sup>2+</sup> (Jain *et al.*, 2012) would be a cause of the  $\alpha$ -synuclein misfolding and induction of ER stress response pathways (Fig. 1). Furthermore, the Parkin-associated endothelin receptor-like receptor (Pael-R) has been shown to induce UPR-mediated cell death in models of PD (Imai *et al.*, 2001). Pael-R is a putative G protein-coupled transmembrane polypeptide, which was originally identified as an ETB receptor homologue (Donohue *et al.*, 1998). When over expressed in cells, this receptor tends to become unfolded, insoluble and ubiquitinated *in vivo* and insoluble Pael-R leads to UPR-mediated cell death. Parkin specifically ubiquitinates this receptor in the presence of ubiquitin-conjugating enzymes resident in the ER and promotes the degradation of insoluble Pael-R, resulting in suppression of cell death induced by Pael-R overexpression (Imai *et al.*, 2001). Although this endothelin receptor-like receptor (Pael-R) has been shown to induce UPR-mediated cell death in PD (Imai *et al.*, 2001), the detailed mechanisms underlying ET-1's neurodegenerative effects in PD remain to be elucidated. Previous work suggests that the ETB receptor is the key mediator of ET-1's neurodegenerative effects following intravitreal administration of ET-1 (Krishnamoorthy *et al.*, 2008). This finding, together with the fact that Pael-R is an ETB receptor homolog, further supports a role in neurodegeneration for ET-1 induced ER stress (via the ETB receptor), which has been recently demonstrated by Jain *et al.* (2012) for pregnancy disorders.

#### FURTHER PATHOLOGICAL PROCESSES INDUCED BY ET-1 AND RELATED ER STRESS

In addition to ER stress, both inflammation and oxidative stress are also closely associated with PD pathology (Bartels and Leenders, 2007; Block and Hong, 2005; Dauer and Przedborski, 2003; Gao and Hong, 2008; Jenner, 2003; Shukla *et al.*, 2011). It is well established that inflammation is ongoing in the PD brain. It is believed that the progressive nature of PD is characterized by chronic inflammation-induced neurodegeneration of dopamine-producing neurons within the substantia nigra and striatum (Bartels and Leenders, 2007; Block and Hong, 2005; Dauer and Przedborski, 2003; Gao and Hong, 2008). There is evidence for increased levels of

pro-inflammatory factors, such as tumour necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin-1 $\beta$  (IL-1 $\beta$ ) and IL-6 in the striatum and substantia nigra (Whitton, 2007). Inflammatory mediators can promote upregulation of  $\alpha$ -synuclein and  $\alpha$ -synuclein aggregation (Fig. 1). ET-1 has previously been shown to induce an inflammatory response in different pathologies, including pregnancy disorders and cardiovascular disease. A recent review by De Miguel and Pollock (2013) reviews the contribution of ET-1 induced ER stress to inflammation. Indeed, previous work has suggested that the ET-1 induced ER stress response can induce an inflammatory response (Jain, 2012; Jain *et al.*, 2012). ER stress can stimulate the activation of pro-inflammatory responses through a number of different pathways. For instance, the ER produces ROS as a by-product of protein folding, which would be increased under conditions of ER stress during repeated attempts to refold misfolded proteins, as would also be the case under the stress response induced by ET-1 (Jain *et al.*, 2012). ROS produced under conditions of ER stress initiates an inflammatory response through phosphorylation and subsequent degradation of the inhibitory subunit of NF- $\kappa$ B. NF- $\kappa$ B is a transcriptional regulator that plays a key role in the induction of inflammation (Rius *et al.*, 2008). A number of factors are released upon activation of the NF- $\kappa$ B pathway, which includes pro-inflammatory cytokines, such as TNF- $\alpha$  and IL-1 $\beta$ , which as described above are known to be implicated in PD pathology.

Apart from stimulating an inflammatory response, ROS produced during ER stress can also contribute to oxidative stress. A large body of evidence from post-mortem studies implicates oxidative stress in the pathology of the PD brain (Jenner, 2003; Shukla *et al.*, 2011). Oxidative stress can promote protein modifications, such as nitrosylation, which can cause  $\alpha$ -synuclein misfolding and aggregation (Uversky, 2007). It has also been found that neurons exposed to such insults can develop increased  $\alpha$ -synuclein levels (Takahashi *et al.*, 2007; Vila *et al.*, 2000). In addition to via ER stress pathways, ET-1 can directly induce oxidative stress, which has been demonstrated in different diseases, including neurodegenerative disorders (Briyal *et al.*, 2011; Fiore *et al.*, 2005; Kalani, 2008; Mi *et al.*, 2012; Pollock, 2006; Potenza *et al.*, 2009). The study by Briyal *et al.* (2011) demonstrated that ETA receptor antagonists block oxidative stress and associated cognitive impairment in Alzheimer's disease. Another study found that in the brain, as part of the oxidative stress and inflammatory response, activated microglia secrete pro-inflammatory factors, such as nitric oxide (NO) that induce astrocytosis and exacerbate neuronal damage (Barbeito *et al.*, 2004). Several other studies have also

found excessive generation of NO and NO-derived reactive nitrogen species is implicated in the pathogenesis of neurodegenerative disorders, including PD (Calabrese *et al.*, 2000; Dehmer *et al.*, 2000; Uehara *et al.*, 2006; Wang *et al.*, 2011). Glial cells, including astrocytes, are considered the major sources of NO production from iNOS in inflammatory neurodegeneration (Bal-Price and Brown, 2001; Weldon *et al.*, 1998). ET-1 is actually found to induce iNOS expression in rat brain astrocytes via the ETB receptor-dependent activation of PI3K, MAPK and NF- $\kappa$ B pathways (Wang *et al.*, 2011). As described earlier, NF- $\kappa$ B would also be stimulated under the ET-1 induced ER stress response. It has also previously been found that ET-1 significantly enhances the synthesis of NO in glial cells through activation of PLC (Filipovich and Fleisher-Berkovich, 2008). The study by Jain *et al.* (2012) demonstrated that ET-1 induces ER stress via activation of the PLC-IP3 pathway. Therefore, there are several overlaps in the signaling pathways already known to be activated by ET-1 that can contribute to neurodegeneration with those demonstrated to be stimulated by the ET-1-induced ER stress response and it would be worth investigating to what extent ET-1 triggers ER stress response pathways that could contribute to PD pathology.

Further to the above, oxidative stress can also induce a plethora of different factors that cause endothelial cell dysfunction (Haorah *et al.*, 2007; Heitzer *et al.*, 2001; Yachie *et al.*, 1999). The pro-inflammatory factors stimulated under conditions of ER stress would further compound this effect and contribute to endothelial cell injury (Fig. 1). Endothelial dysfunction is a systemic pathological state of the endothelium that is broadly defined as an imbalance between vasodilating and vasoconstricting substances produced by (or acting on) the endothelium (Deanfield *et al.*, 2005). Increasing evidence implicates dysfunction of brain endothelial cells in the pathogenesis of neurodegenerative diseases, including PD (Lauwers *et al.*, 2003). Furthermore, a recent study by Makarov *et al.* (2013) has identified markers of endothelial damage in PD that include increased ET-1 levels which further implicates ET-1 (and associated ER stress) in the pathology of PD. The potential importance of ET-1 in PD is also suggested by another recent study that found flavonol-rich cocoa intervention significantly decreases plasma ET-1 in test subjects, which is correlated to improved cognition and reduced development of some of the neuropathological hallmarks of PD, such as endothelial dysfunction and neuroinflammation (Calderon-Garciduenas *et al.*, 2013).

## THERAPEUTIC INTERVENTION BASED AROUND ET-1 IN PARKINSON'S DISEASE

Given the evidence at hand, it would be useful to investigate the potential role of ET-1 in the pathology of PD. If implicated, future research can then investigate the efficacy of blocking ET-1 induced pathology as a therapeutic tool. Recent work has described the use of ET-1 traps as a potential therapeutic tool (Jain, 2013). The application of such a tool has the advantage of blocking the action of ET-1 through both the ETA and ETB receptors. This would have the benefit of potentially targeting all different pathological processes induced by ET-1, including ER stress, oxidative stress and inflammation that might be induced from ET-1 binding the ETA or ETB receptor. The design of ET-1 traps would require constructing a fusion molecule encompassing the different domains of interaction between ET-1 and each of its receptors. As previously described, this would then need to be fused to the Fc portion of human IgG1, which would direct formation of disulfide-linked dimers (Economides *et al.*, 2003; Jain, 2013). Transfection of these expression constructs into mammalian cells would result in secretion of the desired dimeric inline trap, which would act as a soluble receptor that binds ET-1 and therefore prevents it binding the target cell surface receptors. Another approach would be to design therapeutic antibodies against ET-1. There have been recent advances in the design of therapeutic antibodies (Chames *et al.*, 2009), which may be exploited in the development of therapeutic antibodies against ET-1. These would again have the advantage of potentially blocking the pathological actions of ET-1 through both receptors.

## CONCLUSION

It is hoped that this review has highlighted the potential pathological role of ET-1 in PD. ET-1 could be a key mediator of ER stress, inflammation, as well as oxidative stress, which are all important pathological processes in PD. Needless to say, the primary objective would be to first verify the role of ET-1 in PD pathology. Subsequent investigation into potential therapeutic strategies for circumventing potential ET-1-induced pathology might lead to the development of new strategies for therapeutic intervention that ameliorate PD pathology.

## ACKNOWLEDGMENTS

Arjun Jain would like to thank his family for all their love and support. Special thanks also to Vidhi Mehrotra for everything. Ashok Jain would like to thank Santosh, Leela, Kirti and Ira. This paper is dedicated to Prema Hiremath.

## REFERENCES

- Bal-Price, A. and G.C. Brown, 2001. Inflammatory neurodegeneration mediated by nitric oxide from activated glia-inhibiting neuronal respiration, causing glutamate release and excitotoxicity. *J. Neurosci.*, 21: 6480-6491.
- Barbeito, L.H., M. Pehar, P. Cassina, M.R. Vargas and H. Peluffo *et al.*, 2004. A role for astrocytes in motor neuron loss in amyotrophic lateral sclerosis. *Brain Res. Rev.*, 47: 263-274.
- Bartels, A.L. and K.L. Leenders, 2007. Neuroinflammation in the pathophysiology of Parkinson's disease: Evidence from animal models to human *in vivo* studies with [<sup>11</sup>C]-PK11195 PET. *Movement Disorders*, 22: 1852-1856.
- Block, M.L. and J. Hong, 2005. Microglia and inflammation-mediated neurodegeneration: Multiple triggers with a common mechanism. *Prog. Neurobiol.*, 76: 77-98.
- Briyal, S., T. Philip and A. Gulati, 2011. Endothelin-A receptor antagonists prevent amyloid- $\beta$ -induced increase in ETA receptor expression, oxidative stress and cognitive impairment. *J. Alzheimer's Dis.*, 23: 491-503.
- Brostrom, M.A. and C.O. Brostrom, 2003. Calcium dynamics and endoplasmic reticular function in the regulation of protein synthesis: Implications for cell growth and adaptability. *Cell Calcium*, 34: 345-363.
- Calabrese, V., T.E. Bates and A.M. Stella, 2000. NO synthase and NO-dependent signal pathways in brain aging and neurodegenerative disorders: The role of oxidant/antioxidant balance. *Neurochem. Res.*, 25: 1315-1341.
- Calderon-Garciduenas, L., A. Mora-Tiscareno, M. Franco-Lira, J.V. Cross and R. Engle *et al.*, 2013. Flavonol-rich dark cocoa significantly decreases plasma endothelin-1 and improves cognition in urban children. *Front. Pharmacol.*, Vol. 4. 10.3389/fphar.2013.00104
- Chames, P., M. Van Regenmortel, E. Weiss and D. Baty, 2009. Therapeutic antibodies: Successes, limitations and hopes for the future. *Br. J. Pharmacol.*, 157: 220-233.
- Chu, Y. and J.H. Kordower, 2007. Age-associated increases of  $\alpha$ -synuclein in monkeys and humans are associated with nigrostriatal dopamine depletion: Is this the target for Parkinson's disease? *Neurobiol. Dis.*, 25: 134-149.
- Dauer, W. and S. Przedborski, 2003. Parkinson's disease: Mechanisms and models. *Neuron*, 39: 889-909.
- Dawson, T.M. and V.L. Dawson, 2003. Rare genetic mutations shed light on the pathogenesis of Parkinson disease. *J. Clin. Invest.*, 111: 145-151.
- De Miguel, C. and J.S. Pollock, 2013. Does endoplasmic reticulum stress mediate endothelin-1-induced renal inflammation? *Am. J. Physiol.-Regul. Integr. Comp. Physiol.*, 305: R107-R109.
- Deanfield, J., A. Donald, C. Ferri, C. Giannattasio and J. Halcox *et al.*, 2005. Endothelial function and dysfunction. Part I: Methodological issues for assessment in the different vascular beds: A statement by the Working Group on Endothelin and Endothelial Factors of the European Society of Hypertension. *J. Hypertens.*, 23: 7-17.
- Dehmer, T., J. Lindenau, S. Haid, J. Dichgans and J.B. Schulz, 2000. Deficiency of inducible nitric oxide synthase protects against MPTP toxicity *in vivo*. *J. Neurochem.*, 74: 2213-2216.
- Dong, F., X. Zhang, L.E. Wold, Q. Ren, Z. Zhang and J. Ren, 2005. Endothelin-1 enhances oxidative stress, cell proliferation and reduces apoptosis in human umbilical vein endothelial cells: Role of ETM<sub>B</sub> receptor, NADPH oxidase and caveolin-1. *Br. J. Pharmacol.*, 145: 323-333.
- Donohue, P.J., H. Shapira, S.A. Mantey, L.L. Hampton, R.T. Jensen and J.F. Battey, 1998. A human gene encodes a putative G protein-coupled receptor highly expressed in the central nervous system. *Mol. Brain Res.*, 54: 152-160.
- Doyle, K.M., D. Kennedy, A.M. Gorman, S. Gupta, S.J. Healy and A. Samali, 2011. Unfolded proteins and endoplasmic reticulum stress in neurodegenerative disorders. *J. Cell. Mol. Med.*, 15: 2025-2039.
- Economides, A.N., L.R. Carpenter, J.S. Rudge, V. Wong and E.M. Koehler-Stec *et al.*, 2003. Cytokine traps: Multi-component, high-affinity blockers of cytokine action. *Nat. Med.*, 9: 47-52.
- Filipovich, T. and S. Fleisher-Berkovich, 2008. Regulation of glial inflammatory mediators synthesis: Possible role of endothelins. *Peptides*, 29: 2250-2256.
- Fiore, G., P. Florio, L. Micheli, C. Nencini and M. Rossi *et al.*, 2005. Endothelin-1 triggers placental oxidative stress pathways: Putative role in preeclampsia. *J. Clin. Endocrinol. Metab.*, 90: 4205-4210.
- Gao, H.M. and J.S. Hong, 2008. Why neurodegenerative diseases are progressive: Uncontrolled inflammation drives disease progression. *Trends Immunol.*, 29: 357-365.
- Gorman, A.M., 2008. Neuronal cell death in neurodegenerative diseases: Recurring themes around protein handling. *J. Cell. Mol. Med.*, 12: 2263-2280.

- Haorah, J., S.H. Ramirez, K. Schall, D. Smith, R. Pandya and Y. Persidsky, 2007. Oxidative stress activates protein tyrosine kinase and matrix metalloproteinases leading to blood-brain barrier dysfunction. *J. Neurochem.*, 101: 566-576.
- Heitzer, T., T. Schlinzig, K. Krohn, T. Meinertz and T. Munzel, 2001. Endothelial dysfunction, oxidative stress and risk of cardiovascular events in patients with coronary artery disease. *Circulation*, 104: 2673-2678.
- Holtz, W.A. and K.L. O'Malley, 2003. Parkinsonian mimetics induce aspects of unfolded protein response in death of dopaminergic neurons. *J. Biol. Chem.*, 278: 19367-19377.
- Imai, Y., M. Soda and R. Takahashi, 2000. Parkin suppresses unfolded protein stress-induced cell death through its E3 ubiquitin-protein ligase activity. *J. Biol. Chem.*, 275: 35661-35664.
- Imai, Y., M. Soda, H. Inoue, N. Hattori, Y. Mizuno and R. Takahashi, 2001. An unfolded putative transmembrane polypeptide, which can lead to endoplasmic reticulum stress, is a substrate of Parkin. *Cell*, 105: 891-902.
- Jain, A., 2012. Endothelin-1: A key pathological factor in pre-eclampsia? *Reprod. BioMed. Online*, 25: 443-449.
- Jain, A., M. Olovsson, G.J. Burton and H.W. Yung, 2012. Endothelin-1 induces endoplasmic reticulum stress by activating the PLC-IP<sub>3</sub> pathway: Implications for placental pathophysiology in preeclampsia. *Am. J. Pathol.*, 180: 2309-2320.
- Jain, A., 2013. Endothelin-1-induced endoplasmic reticulum stress in disease. *J. Pharmacol. Exp. Ther.*, 346: 163-172.
- Jenner, P., 2003. Oxidative stress in Parkinson's disease. *Ann. Neurol.*, 3: S26-S38.
- Kalani, M., 2008. The importance of endothelin-1 for microvascular dysfunction in diabetes. *Vasc. Health Risk Manage.*, 4: 1061-1068.
- Kramer, B.K., M. Ackermann, S.M. Kohler and G.A.J. Riegger, 1994. Role of endothelin in hypertension. *Clin. Invest.*, 72: 88-93.
- Krishnamoorthy, R.R., V.R. Rao, R. Dauphin, G. Prasanna, C. Johnson and T. Yorio, 2008. Role of the ET<sub>B</sub> receptor in retinal ganglion cell death in glaucoma. *Can. J. Physiol. Pharmacol.*, 86: 380-393.
- Lauwers, E., Z. Debyser, J.V. Dorpe, B. De Strooper, B. Nuttin and V. Baekelandt, 2003. Neuropathology and neurodegeneration in rodent brain induced by lentiviral vectormediated overexpression of  $\alpha$ -synuclein. *Brain Pathol.*, 13: 364-372.
- Makarov, N.S., S.V. Spiridonova, V.V. Nikitina, O.N. Voskresenskaia and N.B. Zakharova, 2013. Molecular markers of endothelial damage in patients with Parkinson's disease. *Zhurnal Nevrologii i Psikiatrii Imeni SS Korsakova*, 113: 61-64 (In Russian).
- Mi, X.S., X. Zhang, Q. Feng, A.C. Lo, S.K. Chung and K.F. So, 2012. Progressive retinal degeneration in transgenic mice with overexpression of endothelin-1 in vascular endothelial cells. *Invest. Ophthalmol. Visual Sci.*, 53: 4842-4851.
- Michalak, M., P. Mariani and M. Opas, 1998. Calreticulin, a multifunctional Ca<sup>2+</sup> binding chaperone of the endoplasmic reticulum. *Biochem. Cell Biol.*, 76: 779-785.
- Nelson, J., A. Bagnato, B. Battistini and P. Nisen, 2003. The endothelin axis: Emerging role in cancer. *Nat. Rev. Cancer*, 3: 110-116.
- Petrucelli, L., C. O'Farrell, P.J. Lockhart, M. Baptista and K. Kehoe *et al.*, 2002. Parkin protects against the toxicity associated with mutant  $\alpha$ -synuclein: Proteasome dysfunction selectively affects catecholaminergic neurons. *Neuron*, 36: 1007-1019.
- Pollock, D.M., 2006. How does endothelin induce vascular oxidative stress in mineralocorticoid hypertension? *Clin. Sci.*, 110: 205-206.
- Potenza, M.A., S. Gagliardi, C. Nacci, M.R. Carratu and M. Montagnani, 2009. Endothelial dysfunction in diabetes: From mechanisms to therapeutic targets. *Curr. Med. Chem.*, 16: 94-112.
- Rius, J., M. Guma, C. Schachtrup, K. Akassoglou and A. Zinkernagel *et al.*, 2008. NF- $\kappa$ B links innate immunity to the hypoxic response through transcriptional regulation of HIF-1 $\alpha$ . *Nature*, 453: 807-811.
- Ryu, E.J., H.P. Harding, J.M. Angelastro, O.V. Vitolo, D. Ron and L.A. Greene, 2002. Endoplasmic reticulum stress and the unfolded protein response in cellular models of Parkinson's disease. *J. Neurosci.*, 22: 10690-10698.
- Shukla, V., S.K. Mishra and H.C. Pant, 2011. Oxidative stress in neurodegeneration. *Adv. Pharmacol. Sci.* 10.1155/2011/572634
- Simonson, M.S. and F. Ismail-Beigi, 2011. Endothelin-1 increases collagen accumulation in renal mesangial cells by stimulating a chemokine and cytokine autocrine signaling loop. *J. Biol. Chem.*, 286: 11003-11008.
- Smith, W.W., H. Jiang, Z. Pei, Y. Tanaka and H. Morita *et al.*, 2005. Endoplasmic reticulum stress and mitochondrial cell death pathways mediate A53T mutant  $\alpha$ -synuclein-induced toxicity. *Hum. Mol. Genet.*, 14: 3801-3811.

- Soto, C. and L.D. Estrada, 2008. Protein misfolding and neurodegeneration. *Arch. Neurol.*, 65: 184-189.
- Takahashi, R., Y. Imai, N. Hattori and Y. Mizuno, 2003. Parkin and endoplasmic reticulum stress. *Ann. N. Y. Acad. Sci.*, 991: 101-106.
- Takahashi, M., L.W. Ko, J. Kulathingal, P.Z. Jiang, D. Sevlever and S.H.C. Yen, 2007. Oxidative stress-induced phosphorylation, degradation and aggregation of  $\alpha$ -synuclein are linked to upregulated CK2 and cathepsin D. *Eur. J. Neurosci.*, 26: 863-874.
- Thomas, B. and M.F. Beal, 2007. Parkinson's disease. *Hum. Mol. Genet.*, 16: R183-R194.
- Uehara, T., T. Nakamura, D. Yao, Z.Q. Shi and Z. Gu *et al.*, 2006. S-nitrosylated protein-disulphide isomerase links protein misfolding to neurodegeneration. *Nature*, 441: 513-517.
- Uversky, V.N., 2007. Neuropathology, biochemistry and biophysics of  $\alpha$ -synuclein aggregation. *J. Neurochem.*, 103: 17-37.
- Vila, M., S. Vukosavic, V. Jackson-Lewis, M. Neystat, M. Jakowec and S. Przedborski, 2000.  $\alpha$ -synuclein up-regulation in substantia nigra dopaminergic neurons following administration of the parkinsonian toxin MPTP. *J. Neurochem.*, 74: 721-729.
- Wang, H.H., H.L. Hsieh and C.M. Yang, 2011. Nitric oxide production by endothelin-1 enhances astrocytic migration via the tyrosine nitration of matrix metalloproteinase-9. *J. Cell. Physiol.*, 226: 2244-2256.
- Weldon, D.T., S.D. Rogers, J.R. Ghilardi, M.P. Finke and J.P. Cleary *et al.*, 1998. Fibrillar  $\beta$ -amyloid induces microglial phagocytosis, expression of inducible nitric oxide synthase and loss of a select population of neurons in the rat CNS *in vivo*. *J. Neurosci.*, 18: 2161-2173.
- Whitton, P.S., 2007. Inflammation as a causative factor in the aetiology of Parkinson's disease. *Br. J. Pharmacol.*, 150: 963-976.
- Winklhofer, K.F., J. Tatzelt and C. Haass, 2008. The two faces of protein misfolding: Gain- and loss-of-function in neurodegenerative diseases. *EMBO J.*, 27: 336-349.
- Yachie, A., Y. Niida, T. Wada, N. Igarashi and H. Kaneda *et al.*, 1999. Oxidative stress causes enhanced endothelial cell injury in human heme oxygenase-1 deficiency. *J. Clin. Invest.*, 103: 129-135.
- Yoshida, H., 2007. ER stress and diseases. *FEBS J.*, 274: 630-658.
- Zhang, K. and R.J. Kaufman, 2008. From endoplasmic-reticulum stress to the inflammatory response. *Nature*, 454: 455-462.