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## Antioxidant Potential and Nutritional Values of Vegetables: A Review

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#### ABSTRACT

More than 70% people in developing countries depend on vegetables and fruits further regular dietary needs. Vegetables prevents human from several sever and chronic diseases. It is necessary to consume daily at least 400 g of vegetables including pulses, nuts and seeds. Since vegetables have antioxidant properties they prevents human from cancer, cardio vascular diseases, diabetes, hyper tension, leprosy, rheumatism, epilepsy, liver and urinary disorder, stroke, paralysis etc. The vegetables contain several phytochemicals possessing antioxidant activity. The major groups of phytochemicals include vitamin A, C, E and K, carotenpoid, terpenoid, flavonoids, polyphenols, saponins, enzymes and minerals. The present paper describes the relationship between phytochemicals and antioxidant activity. More than 50 vegetables and leafy vegetables are identified for their antioxidant activity in terms of DPPH, FRAR, IC<sub>50</sub>, ORAC values. Correlation between antioxidant activities was both positive and negative in vegetables. The present review emphasizes on vegetables as a strong source of antioxidants and a power house of nutrition. The many uncommon vegetables used by the tribals of under developed countries need to be studied for their antioxidant activity and medicinal properties.

Key words: Vegetables, antioxidant properties, phytochemicals, minerals

#### INTRODUCTION

Recent researches have identified a vast majority of antioxidants from vegetables and fruits. Antioxidants like vitamin A, C, E, \( \beta\)-carotene, glutathione precursors like selenium, vitamin B2 (Riboflavin), B3 (Niacin), B6 (Pyridoxin), B9 (Folic acid), B12 (Cynacobalmin), Bioflavanoids which are very rich all seasonal fruits and vegetables with vivid colors stand as prophylactic (Godber, 1990). Selective intake of the foods containing the above antioxidants can prevent the onset of the degenerative diseases like particularly cardiovascular diseases cancers and diabetes. Some very important micronutrients like Chromium and Venadium which improve insulin sensitivity, magnesium preventing retinopathy and vitamin E improving antioxidant defence and insulin sensitivity have become deficient in diabetics. A number of dietary antioxidants exist beyond the traditional vitamins collectively known as phytonutrients or phytochemicals which are being increasingly appreciated for their antioxidant activity. Regular consumption of fruit and vegetables is associated with reduced risks of cancer, cardiovascular disease, stroke, alzheimer disease, cataracts and some of the functional declines associated with aging. Prevention is a more effective strategy than is treatment of chronic diseases. Functional foods that contain significant amounts of bioactive components may provide desirable health benefits beyond basic nutrition and play important roles in the prevention of chronic diseases.

#### ETHNO-MEDICINAL USES OF VEGETABLES

It is well known that many plants and foods have been and continue to be ingested because of their perceived medicinal and health-benefiting characteristics. Providing modern healthcare to rural people in India is still a far reaching goal due to economic constraints (Grover et al., 2002). Hence, people mainly depend on the locally available plant materials to cure various health disorders (Chopra et al., 1956; Grover and Vats, 2001). Plants possess components, which render beneficial properties (Tanabe et al., 2002). Hence, currently attention is being drawn towards exploring plant sources for substances that provide nutritional and pharmaceutical advantages to humans. Green leafy vegetables (GLVs) are a good source of minerals and vitamins. The ethnobotanical reports offer information on medicinal properties of GLVs like antidiabetic (Kesari et al., 2005) anti-histaminic (Yamamura et al., 1998), anti-carcinogenic (Rajeshkumar et al., 2002), hypolipidemic (Khanna et al., 2002) and anti-bacterial activity (Kubo et al., 2004) (Table 1).

Table 1: Traditional uses of some vegetables

Family names	Botanical names (common name)	Traditional uses
Amaryllidaceae	Allium sativum	Heart diseases (including atherosclerosis, high cholesterol,
	Garlie	high blood pressure) and cancer
Araceae	Colocasia esculenta Schott.	Atrophy, bronchial disorders, cough, cuts, injuries as haemost
	Kachalu	burns, stings of bees, wounds
Zingiberaceae	Zinger officinale	Dyspepsia, gastroparesis, rheumatoid arthritis, osteoarthritis, or joint,
	Ginger	muscle injury, short-term relief of pregnancy-related nausea and vomiting
Solanaceae	$Ly copersicon\ esculentum$	Cervix, stomach, mouth, colon and even rectal cancer, heart diseases,
	Tomato	premature aging protective against neurodegenerative diseases
Apiaceae	Daucus carota	Dropsy, retention of urin, gravel, bladder problems, flatulence, nephritic
	Carrot	complaints, ulcer, amenorrhea, eczema, itching, liver disorders, cancer, painful
		urination (strangury), dysmenorrheal, abcesses, bad wounds and colic
Solanaceae	Capsicum annuum	Atonic gout, dyspepsia, tympanitis, paralysis etc
	Green pepper	
Cucurbitaceae	Cucumis sativus	Anticancer (breast cancer, ovarian cancer, uterine cancer) and prostate
	Cucumber	cancer also for weight loss, aiding digestion and chronic constipation
Polygonaceae	Rumex vesicarius Linn.	Purgative, colic, to relieve headaches and alleviate body pain
	Bladder-Dock	
Cucurbitaceae	Lagenaria siceraria	Insomnia, insanity, epilepsy, stomach acidity, indigestion and ulcers,
	Bottle Gourd	constipation and other nervous diseases
Malvaceae	Hibiscus sabdariffa Linn.	Antihypertensive, stomachic, emollient diuretic, mild laxative, treatment
	Roselle	for cardiac, nerve diseases and cancer
Labiatae	Ocimum sanctum	Controlling blood glucose levels and cholesterol levels, headaches, common
	Tulsi	colds, stomach disorders, inflammation, heart diseases, various poisoning,
		malaria and even for periods of women
Meliaceae	Azadirachta indica	Anti-bacterial, anti-viral, anti-fungal, anti-helmintic, anti-diabetic, measles,
	Neem	chicken-pox, malaria, diabetes. Contraceptive and sedative
Cucurbitaceae	$Momordica\ charantia\ { m L}.$	Hypoglycemic, blood purifier, preventing constipation, for weight loss, for eye
	Bitter gourd	caring, useful for medication in early stages of cholera, strengthening immune
		systems and treating HIV infection
Cucurbitaceae	Luffa cylindrical L.	Bitter tonic, emetic, diuretic and purgative and useful in asthma, skin
	Vietnamese gourd	diseases, splenic enlargement, rheumatism, backache, internal hemorrhage,
		anthelmentic, carminative, laxative, depurative, emollient, expectorant, tonic,
		chest pains as well as hemorrhoids

Table 1: Continue

Family names	Botanical names (common name)	Traditional uses
Cucurbitaceae	Luffa acutangula (L.) Roxb. Silk Gourd	Fever, restlessness and thirst during the course of febrile diseases, dyspnea, cough profuse leukorrhea, strangury (a slow and painful spasmodic discharge of urine drop by drop) with blood, furuncle, galactostasis, carbuncle and swelling
Cucurbitaceae	Trichosanthes dioica Roxb.	Antipyretic, diuretic, cardiotonic, laxative, antiulcer, etc.
	Pointed gourd	
Cucurbitaceae	Luffa cylindrical (L.) Roem.	Diuresis, promoting blood circulation, dredge main and collateral channels,
	Towel gourd/ Sponge guard	detoxification effect, hemostasis, diminish inflammation, regulate menstruation,
		eliminating phlegm cough effect, skin aging effects
Solanaceae	Solanum tuberosom L.	Appetite, antibacterial, arthritis, infections, boils, burns and sore eyes
	Potato	
Solanaceae	Solanum melongena L.	Blood circulation, preventing arteriosclerosis, wound healing, preventing cough
	Egg plant	and rheumatism
Musaceae	Musa paradisica L. (unripe)	Bronchitis, cold, snakebite, reducing hyperglycemia in normal and
	Banana	diabetic, protected the gastric mucosa from aspirin-induced erosion,
		stimulating gastric and colonic mucosa
Caricaceae	Carica papaya L.	Skin blemishes, treating stomach ulcers, diphtheria
	Papaya	andconstipation, kidney failure and anticancer
Solanaceae	Capsicum frutescens L.	Skin diseases, headache and night blindness, tuberculosis, stomach-ache,
	Chili pepper	backache, cough and chest pain
Solanaceae	Capsicum annuum Linn.	Lethargic affections, atonic gout, dyspepsia, tympanitis,
	Var. grossam (Willd.) Sendt.	paralysis, malignent scarlet fever etc.
	Capsicum	
Solanaceae	Nicotiana tabacum Linn.	Eczema, itch, food poison, haemost, snake bite, toothache,
	Tambacco	spongy gums and wounds
Fabaceae	Bauhinia variegate	Antidysentery, antidiarrhea laxative, to cure
· abaca	Orchid tree	leucorrhoea and mumps and piles
Mimosaceae	Parkia roxburghii G. Don.	Chronic dysentery, piles and as laxative. Its most sought after
viiniosaceae	Kedaung	property is as anti-bacteria and is applied in traditional medicine
	Redading	for infections and stomach disorders
Daggara	7.0 m vaa muu T	
Poaceae	Zea mays L.	Diuretic, heart disease, hypertension and kidney stone and
~ 1 1	Corn	for lowering blood pressure
Convolvulaceae	Ipomeae batatas Sweet potato	Diarrhea, constipation and a good source of vitamin A and C
Moraceae	Artocarpus heterophyllus Lamk	Skin diseases and asthma and for healing stomach ulcers, diuretic.
	Jack fruit	
Poaceae	Bambusa tulda Roxb.	Chicken pox and other skin diseases; the paste of shoot is
	Jati	used haemostatic and antidote
Elaeagnaceae	Elaegnus latifolia Silverberry	Syphilis and blackfoot children
Myricaceae	Myrica nagi	Diarrhea, jaundice, scrofula, astringent, chronic inflammation of throat,
wyncaceae	Bayberry	leucorrhoea, uterine haemorrhage
Fahassas	Glycine max L. Merr.	Allergies diabetes, skin, eye diseases and dandruff problems
Fabaceae	· ·	Aftergles diabetes, skin, eye diseases and dandrun problems
G 1	Soyabean	
Fabaceae	Cicer arietinum Linn.	Lowering blood sugar levels, stomach disorders like dyspepsia, vomiting,
	Chickpea	indigestion, costiveness, diarrhea and dysentery, skin and scalp
		cleaning and moisturizing agent
Fabaceae	Phaseolus vulgaris Linn.	Bean-diuretic, hypotensive, resolvent, regulates blood sugar, water retention;
	Kidney Bean	albuminuria, especially of pregnancy; premenstrual tension

Table 1: Continue

E:	Botanical names	Mar. 3/4/
Family names	(common name)	Traditional uses
Fabaceae	Lens culinaris Medik. Masari	Diuretic, tonic, laxative and astringent to the bowels; useful in constipation and other intestinal affections
Fabaceae	Pisum sativum Linn.	Refrigerant, appetizer, fattening, laxative, alleviative of bile, phlegm and burning
rapaceae	Mattar	of the skin
Moringaceae	Moringa oleifera Lam.	Cardiac, antitumor, antipyretic, antiepileptic, antiinflammatory, antiulcer,
wormgaeeae	Drumstick tree	antispasmodic, diuretic, antihypertensive, cholesterol lowering, antioxidant,
	Di unisuck u ce	antidiabetic, hepatoprotective, antibacterial and antifungal activities
Brassicaceae	Raphanus sativus Linn.	Hypoglycemic potential coupled with antidiabetic efficacy, gastrointestinal (GI)
31 dassiede ode	Radish	prokinetic effects and strong antioxidant activity
Brassicaceae	Brassica campestris Linn.	Boils, migraine, muscular pain, skin disorder, bronchial disorder, cough, leprosy,
51 0551000000	Mustard	pneumonia, scabies, syphilis
Brassicaceae	Brassica nigra Koch	Rheumatism
	Rai	
Brassicaceae	Brassica oleracea Linn. Var.	Diuretic, laxative, stomachic and antihelminthic, digestive, tonic cholesterol-lowering
	White cabbage	anticancer, antifungal
Brassicaceae	Brassica rapa Linn.	Antiviral, antibacterial, anticancer activity and suppress thyroid function
	Cruciferous vegetables	•
Brassicaceae	Eruca sativa Mill.	Astringent, diuretic, digestive, emollient, tonic, depurative, laxative, rubefacient
	Rocket salads	and stimulant
Brassicaceae	Raphanus sativus Linn.	Intestinal parasites, carminative, diuretic, expectorant, laxative and stomachic,
	Rat-Tail Radish	asthma and other chest complaints
Chenopodiaceae	Chenopodium album Linn.	Apetite oleaginous, anthelmintic, laxative, diuretic, aphrodisiac, tonic, useful in
	White goosefoot	biliousness, abdominal pains, eye diseases, throat troubles, piles, diseases of the
		blood, the heart, the spleen, laxative, anthelmintic and cardiotonic
Campanulaceae	Pratia begonifolia	Urinary diseases, dissolving kidney and gall bladder stones
	Round leaf star creeper	
Amaranthaceae	Celosia cristatae L.	Diarrhea, urinary tract infection and excessive menstrual problem; leaf extract is
	Cock's comb	used in cuts and injuries
Brassicaceae	$Brassica\ oleracea$	Anti-cancer, especially in the prevention and treatment of gastric cancer, breast
	Brœcoli	cancer effect is the best
Brassicaceae	$Brassica\ oleraceae$	Asgoitrogens, anticancer and thyroid hormone production
	Linn. Var. capitat	
	Brussels sprouts	
Brassicaceae	$Brassica\ oleracea$	Cancer prevention, particularly with respect to the following types of cancer: bladder
	Cauliflower	cancer, breast cancer, colon cancer, prostate cancer and ovarian cancer
Cucurbitaceae	Cucurbita pepo	$Antirheumatic, demulcent, laxative, diuretic, nervine, taenifuge, good immune\ booster and the property of t$
	Pumpkin	also to treat kidney problems, prostate hyperplasia and intestinal parasites
Amaryllidaceae	Allium cepa	Colds, fever, allergy bacterial infection, congestion and sore throat, effectively
	Red Onion	treats an earache
Moraceae	Ficus hispida L.	Urinary bladder complains also applied on skin diseases and leprosy
	Creeping figs	
Malvaceae	Abelmoschus esculenus (Linn.)	$Diuretic, \ demulcent, \ stomachic, \ stimulant, \ cooling, \ tonic, \ carminative, \ aphrodisiac,$
	Lady's Finger	nerve tonic, stomachic, sweetens the breath and antiseptic properties
Leguminosae	0 , 0	Aromatic, cooling, laxative, skin and mucous membranes, relieve irritation of the skin
	Fenugreek	and alleviate swelling and pain
Chenopodiaceae	Spinacia oleraceae Linn.	Anti-oxidant, antiproliferative, antiinfammatory, antihistaminic, CNS depressant,
	Spinach	protection against gamma radiation, hepatoprotective etc
Cucurbitaceae	Cucumis sativus Linn.	Antihelminthic, cooling, diuretic and strengthening
	Khira	

Table 1: Continue

Family names	Botanical names (common name)	Traditional uses
Cucurbitaceae	Cucurbita maxima	Burns, scalds, inflammations, abscesses, boils and is remedy for migraine, neuralgiant
	Duch. ex Lam.	haemoptysis and hemorrhages
	Red Gourd	
Cucurbitaceae	L. acutangula M. Roem	Convulsion, cramps, fever, jaundice, madness, scabies,
	Kali tori	syphilis, tetanus, snake bite
Apiaceae	Coriandrum sativum Linn.	Carminative, expectorant, aromatic, narcotic, stimulant and stomachic
	Coriander	properties and the raw ones are chewed to stimulate the flow of gastric
		juices and sweeten the breath
Apiaceae	Foeniculum vulgare	Primarydysmenorrhea and carminative
	Fennel	
	Cyamopsis	Laxative, regular healthy digestive tract healthy
	tetragonoloba Linn.	Irritable Bowel syndrome (IBS), Crohn's disease and colitis
	Cluser bean	and also in reducing cholesterol
Apiaceae	Anethum graveolens Linn.	Mucosal protective, antimicrobial, antioxidant, antigastric irrition,
	Dill	stimulate milk production and change the estrous cycles, affects
		hypothalamo-gonadal axis in females and also affect male reproductive
		organ for enhancing male sexual behaviors
Amaranthaceae	Chenopodium album	Antiinflammatory, antispasmodic, antiallergic,
	White Goosefoot	antibacterial and antifungal remedies
Apiaceae	Centella asiatica	Asthma, ulcers, lepsory, lupus, vein diseases memory improvement,
	Centella	antidepressant, antibacterial, antifungal, psoriasis and anti-cancer agent
Cucurbitaceae	Momordica balsamina	Liver deficiencies, blood cleanser, ulcers of the stomach and duodenum,
	African cucumber	disorders. Used as a purgative, emetic, bitter stomachic and as a wash for fever
Amaranthaceae	Amaranthus spinosus	Astringent, diaphoretic, diuretic, emollient, febrifuge, purgative. Used to treat snak
	Spiny amaranth	bites, ulcerated mouths, vaginal discharges, nosebleeds, wounds
Amaranthaceae	Amaranthus hybridus	Intestinal bleeding, diarrhea, excessive menstruation
	Smooth amaranth	
Amaranthaceae	Amaranthus dubius	Alleviate stomach pains
	Spleen amaranth	
Acanthaceae	Justicia flava	Stomach ache, diarrhea, fevers, yaws, emetics and eye lotions
	Yellow justice	
Polygonaceae	Emex australis	Gastrointestinal disorders, colic, biliousness and dyspepsia
	Spiny emex	· · · · · · · · · · · · · · · · · · ·
Asteraceae	Galinsoga parviflora	Treating nettle stings
	Gallant soldier	
Portulacaceae	Portulaca oleracea	Dysentery, diarrhoea, haemorrhoids and enterorrhagia, antibacterial,
	Pigweed	antiscorbutic, depurative, diuretic, febrifuge
Fabaceae	Senna occidentalis	Stomach pains, biliousness, fevers, jaundice, ringworms,
	Coffeeweed	sore throats and edemas, abscesses and skin diseases
Chenopodiaceae	Chenopodium album	Anthelmintic, antiphlogistic, antirheumatic, mildly laxative,
•	Lamb's-quarters	odontalgic, bug bites, sunstroke and rheumatic joints
Pedaliaceae	Ceratotheca triloba	Stomach cramps, nausea, fever and diarrhea
	African foxglove	• / /
Mackinlayaceae	Centella asiatica	Inflammations, fevers, improves healing, immunity, improves
	Pennywort	the memory, wounds, chronic skin conditions, venereal diseases,
	v	malaria, varicose veins, ulcers and nervous disorders and senility
Cleomaceae	Cleome monophylla	Coughs, swellings and anthelmintic
	Spindle pod	

Table 1: Continue

Family names	Botanical names (common name)	Traditional uses		
Commelinaceae	Commelina nudiflora Linn.	Febrifuge, rubefacient, diuretic, good blood coagulant, antifebrile and		
	Climbing dayflower	antidote, tonic for the heart, antifungal and antibacterial		
Caesalpinaceae	Cassia tora	Ageing, atherosclerosis, cancer, inflammatory, joint disease, asthma,		
	The Sickle Senna	diabetes and degenerative eye diseases		
Fabaceae	Bauhinia purpurea	Febrifugal, antidiarrhoeal, antidysenteric remedy and astringent		
	Butterfly tree			
Solanaceae	Solanum nigrum	Antitumorigenic, antioxidant, anti-inflammatory, hepatoprotective, diuretic		
	Black Nightshade	and antipyretic sedative, narcotic, tonic, laxative and appetite stimulant		
Solanaceae	$Physalis\ viscose$	Antioxidants, anti-inflammatory and anticancer		
	Gooseberry			
Portulacaceae	Portulaca oleracea	Antitumorigenic, antioxidant, anti-infammatory, anti-rheumatic,		
	Purslane	anti-dibetic, diuretic and wound healing		
Antioxidant defe	nse system 	Exogenous antioxidants, Principal dietary antioxidants from fruits		
Endogenous anti	oxidants enzymatic antioxidants	vegetables and grains		
Superoxide dis	mutase (SOD)	Vitamins: Vitamin C, Vitamin E		
Enzyme detoxify	ing superoxide radical (O2*-)	Trace elements-zinc, selenium		
Catalase (CAT)	and glutathione peroxidase (GI	Px) Carotenoids: β-carotenoid, lycopene, lutein, zeaxanthin		
Enzymes involve	d in the H <sub>2</sub> O <sub>2</sub> and GPx against	Phenolic acids: Chlorogenic acids, gallic acids, cafeic acids etc		
both $H_2O_2$ and $R_1$	OOH)	Flavonoids: Quercetin, kaempferol, myricetin		
Glutathione rec	ductase	Flavanols: Ptoanthocyanidin and pelagonidin		
Enzyme involved	in the regeneration of glutathione	Isoflavones: Genistein, daidzein and glycitein		
Thioredoxin re	ductase	Flavanones: Naringenin, eriodictyol and hesperetin		
Enzyme involved	in the protection against protein or	didation Flavones: Lutin and apigenin		
Glucose-6-phos	phate dehydrogenase			
•	in the regeneration of NADPH			
Non-enzymatic	antioxidants			

#### PROTECTIVE ACTION OF ANTIOXIDANTS

(principal intracellular reducing agents) Glutathione (GSH), uric acid, lipoic acid, NADPH, coenzyme Q, albumin, bilirubin

Increased consumption of fruits and vegetables has been associated with protection against various age-related diseases (Ames *et al.*, 1993; Steinberg, 1991). What dietary constituents are responsible for this association is not known, but well-characterized antioxidants, including vitamins C and E, or  $\beta$ -carotene, are often assumed to contribute to the observed protection (Ames, 1983; Buring and Hennekens, 1997; Gey *et al.*, 1991; Stahelin *et al.*, 1991; Steinberg, 1991; Willett, 1994). However, the results from intervention trials have not been conclusive regarding the protection following supplementation with such antioxidants (Hennekens *et al.*, 1996; Omenn *et al.*, 1996; Prieme *et al.*, 1997; Van Poppel *et al.*, 1995). Recent epidemiological evidence indicates that the putative beneficial effects of a high intake of fruits and vegetables on the risk of diseases of aging may not be exclusively due to these antioxidants (Hertog *et al.*, 1992; Knekt *et al.*, 1997), but other antioxidant phytochemicals contained in fruits and vegetables may be equally important (Table 2 and 3).

Table 3: Vegetables containing nutrients

Nutrients	Functions in the body/benefits	Vegetable sources
Vitamins		
Vitamin A	Helps maintain good vision (Necessary for night	Dark-green vegetables (such as spinach and turnip greens), orange
	vision), resistance to infections and supports growth	vegetables (such as carrots, pumpkin and sweet potato), orange
	and repair of body tissues. Also maintains integrity	flesh fruits (such as apricot, mango, orange, papaya, peach and
	of white and red blood cells, assists in	pineapple) and tomato
	immune reactions, helps maintain the stability	
	of cell membranes	
Vitamin C	Essential element in collagen formation (strengthens	Citrus fruits and their juices, berries, dark green vegetables
	blood vessels, forms scar tissue, is a matrix for bone	(spinach, asparagus, green peppers, brussel sprouts, broccoli,
	growth); an antioxidant; strengthens resistance	watercress, other greens), red and yellow peppers, tomatoes and
	to infections and improves absorption of iron	tomato juice, pineapple, cantaloupe, mangos, papaya and guava
Vitamin E	Helps maintain cell membrane, red blood cell	Vegetable oils such as olive, soybean, corn, cottonseed and
	integrity, protects vitamin A and fatty acids	safflower, nuts and nut butters, seeds, whole grains, wheat,
	from oxidation	wheat germ, brown rice, oatmeal, soybeans, sweet potatoes,
		legumes (beans, lentils, split peas) and dark leafy green
Vitamin K	Helps make factors that promote blood eletting	vegetables
vitaliiii K	Helps make factors that promote blood clotting	Nuts, lentils, green onions, crucifers (cabbage, broccoli) baked potato or sweet potato, banana and plantain, cooked dry beans
Folic acid	Part of accommos used in new cell synthesis	and leafy greens  Dark-green leafy vegetables (such as spinach, mustard greens
rone aciu	Part of coenzymes used in new cell synthesis  Essential for blood cell formation, protein	
		and romaine lettuce), legumes (cooked dry beans and peas, green
Fiber	metabolism and prevention of neural tube defects  Diabetes, heart disease	peas and seeds  Most fresh fruits and versetables, muts, beens and peas
rner Selected min	•	Most fresh fruits and vegetables, nuts, beans and peas
Magnesium	Involved in bone mineralization, the building of	Spinach, lentils, potato, banana, nuts, corn, cashews
8	protein, enzyme action, normal muscular	,, F,,,,,
	contraction and transmission of nerve impulses	
Calcium	Osteoporosis, muscular/skeletal, teeth, blood	Cooked vegetables (such as beans, tomatoes) peas,
	pressure	papaya, raisins, orange, pumpkin and cauliflower
Potassium	Hypertension (blood pressure) stroke,	Baked potato or sweet potato, banana, cooked dry beans, cooked
2 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	Arteriosclerosis	greens, dried fruits (such as apricots and prunes)
Manganese	Involved in the formation of bone, as well as in	Nuts, whole grain cereals, beans, rice, dried fruits, green leafy
	enzymes involved in amino acid, cholesterol	vegetables
	and carbohydrate metabolism	
Molybdenum	Important in a variety of enzyme systems	Legumes and grains
Copper	Supports healthy bones, muscles and blood	Legumes, nuts, seeds, raisins, whole grains
	vessels. Assists in iron absorption	
Boron	Bone health, prevention of osteoporosis	Non-citrus fruits, leafy vegetables
Calcium	The principal mineral of bones and teeth, also	Broccoli, chard and legumes
	involved in normal muscle contraction	
	(including heart muscle)	
Chromium	With insulin helps to convert carbohydrates	Broccoli, radish, onion, cabbage, cauliflower, garden pea, kale
J.m Jim dill	and fats into energy	and celery
Selenium	It is responsible for boosting the immunity of cancer	Crimini mushrooms, mustard seeds, broccoli, garlic, spinach and
Colonidii	patients and plays an important role in cellular	sunflower seeds
	function as it forms the integral part of an	Sumowol Secus
	important antioxidant enzyme	

Table 3: Continue

Nutrients	Functions in the body/benefits	Vegetable sources
Zinc	Necessary to synthesize DNA, for wound healing,	Nuts, whole grains and legumes
	healthy growth and development of the body	
	during adolescence, childhood and pregnancy	
Polyphenols		
Anthocyanins	Act as antioxidants, the association between	Red cabbage, red onion, apricot, kale, broccoli etc.
	anthocyanins and oxidative stress appears to relate	
	to the ability of anthocyanins to reduce excitation	
	pressure and, hence, the potential for oxidative	
	damage	
Catechins		
Isoflavones	Isoflavones have an influence on the women's health	Soy-beans, garbanzo beans, chick peas, red clover and legumes
	during menopause and it is necessary to reduce the	
	risk of fibroids and andomertriosis	
Rutin	Ascular degeneration, vascular bleeding (bruising/	Green and yellow peppers, tomatoes, onions, broccoli and parsley
	capillary fragility, nose bleeds, periodontal bleeding,	
	varicose veins, hemorrhoids and aneurism	
Hesperidin	Hesperidin is effectively used as a supplemental	Leafy vegetables such as swiss chard, spinach, beets/beet greens,
	agent in the treatment protocols of complementary	parsley, lemon and rhubarb
	settings and also helps in reducing oedema or excess	
	swelling in the legs due to fluid accumulation. Its	
	deficiency has been linked to abnormal capillary	
	leakiness as well as pain in the extremities	
	causing aches, weakness and night leg cramps	
Quercetin	Quercetin has been found to be an effective	Vegetables, spices, seeds, nuts, mushrooms, celery, parsley and
	hyperthermia sensitizer, making it a potential	herbs
	adjunct therapeutic agent in the treatment of	
	various cancers that are sensitive to heat stress	
	(colon adenocarcinoma, prostate, ovarian, uterine	
Tannins	or cervical cancer, leukemia, melanoma and others)	Plack being and being white being For plant Union beneng
ranimis	Tannins have also been reported to anticarcinogenic	Black beans, red beans, white beans, Egg plant, Unripe banana
	and antimutagenic potentials of tannins may be	etc.
	related to their antioxidative property, also help to	
	accelerate blood clotting, reduce blood pressure, decrease the serum lipid level, produce liver	
	necrosis and modulate immunoresponses	
Lyannana	Act as strong antioxidant powers, with some studies	Tomato, dried parsley, basil, Asparagus, red cabbage and red
Lycopene	showing a reduced risk of cancer, cardiovascular	chilli
	disease and macular degeneration	Cinin
Lutein	Consuming lutein as part of the dietmight help to	Dark green leafy vegetables like spinach and kale, corn, Broccoli,
Lutem	prevent AMD (Age-related macular degeneration),	Papaya, green peas, Carrot, Brussels sprouts, Turnip greens,
	Reducing the risk of developing eye cataracts,	zucchini etc
	reduced risk of developing breast cancer	zuccimii eu
Cwestowanthins	Cryptoxanthin is an antioxidant and may help	Petals and flowers of plants in the genus Physalis, orange rind,
Orypoxanomis	prevent free radical damage to cells and DNA,	papaya
	as well as stimulate the repair of oxidative	papaya
	damage to DNA	
Ellagic acid	Ellagic acid has antiproliferative and antioxidant	Freeze-dried berries, pears, peaches, plums, grapes, apples, kiwi
magic aciu	properties, chemoprotective effect in cellular	and several nuts
	models by reducing oxidative stress	and soret at titus
-	moders by reducing oxidative stress	

Over the years, consumers have been paying more and more attention to the health and nutritional aspect of horticultural products. Having a diet rich in fruits and vegetables will be able to provide some protection against the common diseases such as cardiovascular diseases, cancers and other agerelated degenerative diseases (Scalzo et al., 2005). Evidence shows that free radicals are responsible for the damage of lipids, proteins and nucleic acid in cells could lead to these common diseases (Alothman et al., 2009). Recent studies showed that frequent consumption of fruits and vegetables can reduce the risk of stroke and cancer which is related to the antioxidant microconstituents contained on the plant parts. Different vegetables may exhibit different capacities due to the presence of different dietary antioxidants, such as vitamin C and E, carotenoids, flavanoids and other phenolic compounds (Saura-Calixto and Goni, 2006).

#### ANTIOXIDANT METABOLITES IN VEGETABLES

Antioxidants 'Antioxidants' are substances that neutralize free radicals or their actions (Sies, 1996). Nature has endowed each cell with adequate protective mechanisms against any harmful effects of free radicals: Superoxide Dismutase (SOD), glutathione peroxidase, glutathione reductase, thioredoxin, thiols and disulfide bonding are buffering systems in every cell. α-Tocopherol (vitamin E) is an essential nutrient which functions as a chain-breaking antioxidant which prevents the propagation of free radical reactions in all cell membranes in the human body. Ascorbic acid (vitamin C) is also part of the normal protecting mechanism. Other non-enzymatic antioxidants include carotenoids, flavonoids and related polyphenols, α-lipoic acid, glutathione etc., (Sies, 1996; Cadenas and Packer, 1996; Halliwell and Aruoma, 1993). Antioxidant compounds like phenolic acids, polyphenols and flavonoids scavenge free radicals such as peroxide, hydroperoxide or lipid peroxyl and thus inhibit the oxidative mechanisms that lead to degenerative diseases. There are a number of clinical studies suggesting that the antioxidants in fruits, vegetables, tea and red wine are the main factors for the observed efficacy of these foods in reducing the incidence of chronic diseases including heart diseases and some cancers (Table 2 and 3).

Vitamin C: Ascorbic acid (vitamin C), a powerful, water-soluble antioxidant as scavenger of ROS (Smirnoff, 2000). To prevent or at least alleviate deleterious effects caused by ROS. It has the abitlity to donate electrons in a number of enzymatic and non-enzymatic reactions. As a directly scavenge  ${}^{1}O_{2}$ ,  $O_{2}^{*-}$  and  ${}^{*}OH^{-}$  and regenerate tocopherol from tocopheroxyl radical, thus, providing membrane protection (Thomas *et al.*, 1992). Ascorbic acid also acts as co-factor of violaxanthin de-epoxidase, thus, sustaining dissipation of excess excitation energy (Smirnoff, 2000). Ascorbic acid plays an important role in minimizing the damage caused by oxidative process. This is performed by its synergistic action with other antioxidants (Smirnoff, 2005; Athar *et al.*, 2008). Ascorbate-Glutathione cycle, it plays a role in preserving the activities of enzymes that contain prostheritic transition metal ions (Noctor and Foyer, 1998). Both vitamins C and E show very apparent effects protecting the human body from free radicals and disease. These super vitamins have been shown to have positive effects on tumors activated by ultraviolet light (Table 3).

**Vitamin E:** Tocochromanols, known as vitamin E, are essential components of biological membranes, exclusively located in the plastid or thylakoid membranes (Munne-Bosch, 2005), where they have both antioxidant and non-antioxidant functions (Kagan, 1989). They occur in four different types or isomers, namely,  $\alpha$ ,  $\beta$ ,  $\gamma$  and  $\delta$ -tocopherols. All four types of tocopherols structurally consist of a chromanol head group attached to the phytyl tail, which together giving

vitamin E compounds amphipathic character (Kamal-Eldin and Appelqvist, 1996). Among others,  $\alpha$ -tocopherol with its three methyl substituents has the highest antioxidant activity of tocopherols (Kamal-Eldin and Appelqvist, 1996) and relative antioxidant activity of the tocopherol isomers in vivo is  $\alpha > \beta > \gamma > \delta$ , due to the methylation pattern and the amount of methyl groups attached to the phenolic ring (Blokhina et al., 2003).  $\alpha$ -tocopherols prevent the chain propagation step in lipid auto-oxidation and this makes it an effective free radical trap.  $\alpha$ -tocopherols found in green parts of plants scavenge lipid peroxy radicals through the concerted action of other antioxidants (Kiffin et al., 2006; Hare et al., 1998). Further, tocopherols are also known to protect lipids and other membrane components by physically quenching and chemically reacting with oxygen in chloroplasts, thus, protecting the structure and function of PSII (Igamberdiev and Hill, 2004). The main function of tocopherol lies in its fatty acyl chain-breaking activity, which scavenges Reactive Oxygen Species (ROS) resulting from photosynthesis, thus protecting polyunsaturated fatty acid chains (PUFAs) from lipid peroxidation. Increasing evidence suggests that in higher plants, vitamin E may play a protective role in cell membrane systems, thus maintaining the integrity and normal function of the photosynthetic apparatus (Havaux et al., 2005; Collin et al., 2008) (Table 3).

Vitamin K: This is a group of structurally similar, fat-soluble vitamins that the human body needs for posttranslational modification of certain proteins required for blood coagulation and in metabolic pathways in bone and other tissue. They are 2-methyl-1,4-naphthoquinone (3-) derivatives (Stafford, 2005). This group of vitamins includes two natural vitamers:vitamin  $K_1$  and vitamin  $K_2$ . Vitamin K<sub>1</sub>, also known as phylloquinone, phytomenadione, or phytonadione, is synthesized by plants and is found in highest amounts in green leafy vegetables because it is directly involved in photosynthesis (Newman et al., 2008). It may be thought of as the "plant form" of vitamin K. It is active in animals and may perform the classic functions of vitamin K in animals, including its activity in the production of blood clotting proteins. Animals may also convert it to vitamin K<sub>2</sub>. Vitamin K<sub>2</sub>, the main storage form in animals, has several subtypes, which differ in isoprenoid chain length. These vitamin K<sub>2</sub> homologs are called menaquinones and are characterized by the number of isoprenoid residues in their side chains. Vitamin K<sub>1</sub> is found chiefly in leafy green vegetables such as dandelion greens, spinach, swiss chard and Brassica (e.g., cabbage, kale, cauliflower, broccoli and brussels sprouts) and often the absorption is greater when accompanied by fats such as butter or oils (Table 3). Some vegetable oils, notably soybean, contain vitamin K, but at levels that would require relatively large calorific consumption to meet the USDA recommended levels (Weber, 2001).

Folate: Folic acid (also known as folate, vitamin M, vitamin B<sub>9</sub>, vitamin B<sub>c</sub> (orfolacin), pteroyl-L-glutamic acid and pteroyl-L-glutamate are forms of thewater-soluble vitamin B<sub>9</sub>. Folate is composed of the aromatic pteridine ring linked to para-aminobenzoic acid and one or more glutamate residues. Folic acid is itself not biologically active, but its biological importance is due to tetrahydrofolate and other derivatives after its conversion to dihydrofolic acid in the liver (Bailey and Ayling, 2009). It is present in a wide variety of foods, such as green-leafy vegetables and fruits (Table 3). They very rich in Leafy vegetables such as beets, corn, tomato, broccoli, brussels sprouts, romaine lettuce and bok choy and some of Asian vegetables (Houlihan et al., 2011; Hoffbrand and Weir, 2001). Folate is important for the synthesis of DNA, transfer RNA and the amino acids cysteine and methionine. DNA synthesis plays an important role in germ cell development and therefore, it is obvious that folate is important for reproduction. It has also been

reported that folic acid, the synthetic form of folate, effectively scavenges oxidizing free radicals and as such can be regarded as an antioxidant (Joshi *et al.*, 2001). Despite its water-soluble character, folic acid inhibits lipid peroxidation (LPO). Therefore, folic acid can protect bio-constituents such as cellular membranes or DNA from free radical damage (Joshi *et al.*, 2001). Only limited knowledge is available on the impact of dietary folate and synthetic folic acid on (sub) fertility.

Fibers: Dietary fiber is largely composed of complex carbohydrates that are somewhat resistant to digestion. One major component of soluble fibers is pectin, which is largely composed of uronic acid residues such as galacturonic acid. Pectin and other soluble polysaccharides may undergo some metabolism in the small intestine and especially in the large intestine through bacterial enzymes, converting it to products that contribute to maintaining the colonic microflora, which is beneficial to digestion (Weisburger et al., 1993; Cummings et al., 1979; Holloway et al., 1983). Insoluble fiber like cellulose, found in plant cell walls, can aid in waste and toxin removal through several mechanisms (Weisburger et al., 1993). Dietary fibers in foods are also beneficial for good health. Physiological impacts of insufficient dietary fiber intake are constipation, increased risk of coronary heart disease and increased fluctuation of blood glucose and insulin levels (AACC, 2001; Jenkins et al., 1998). Including fruits and vegetables in the human diet may be beneficial, based on their dietary fiber content, with regard to some cancers (Weisburger et al., 1993; Harris and Ferguson, 1993) (Table 3).

## SECONDARY METABOLITES IN VEGETABLES IN FREE RADICAL SCAVENGING ACTIVITY

Secondary constituents are the remaining plant chemicals such as alkaloids (derived from amino acids), terpenes (a group of lipids) and phenolics (derived from carbohydrates). Antioxidants are secondary constituents or metabolites found naturally in the body and in plants such as fruits and vegetables. Plants produce a very impressive array of antioxidant compounds that includes carotenoids, flavonoids, cinnamic acids, benzoic acids, folic acid, ascorbic acid, tocopherols and tocotrienols to prevent oxidation of the susceptible substrate (Hollman, 2001). Common antioxidants include vitamin A, vitamin C, vitamin E and certain compounds called carotenoids (like lutein and beta-carotene) (Hayek, 2000). These plant-based dietary antioxidants are believed to have an important role in the maintenance of human health because our endogenous antioxidants provide insufficient protection against the constant and unavoidable challenge of reactive oxygen species (ROS; oxidants) (Fridovich, 1998).

#### PHENOLIC COMPOUNDS

Phenolic compounds comprise a large group of organic substances and flavonoids are an important subgroup. This group constitutes the majority of dietary antioxidants. There are four major groups of flavonoids, i.e., anthocyanins, flavones, flavonols and the isoflavonols. Anthocyanins, which are coloured flavonoids responsible for a wide range of colours in plants, can scavenge free radicals, particularly singlet oxygen due to their reversible oxidation-reduction properties. Flavonoids, mainly present as colouring pigments in plants also function as potent antioxidants at various levels (Sies, 1996; Cadenas and Packer, 1996). Flavonols include quercetin found in onion and to a lesser extent in French beans and broad beans. The isoflavonoids (isoflavones) are a group of flavonoids in which the position of one aromatic ring is shifted. Isoflavoids may also be responsible for the anticancer benefits of food prepared from soybeans. Food

sources that are especially rich in polyphenols include, among others, potato, plums, leafy vegetables, whole grain products and coffee (Souci et al., 2000) (Table 3). Polyphenols scavenge free radicals (R\*) possessing an unpaired electron either by donation of hydrogens or electrons, resulting in comparatively stable phenoxyl (PhO\*) radicals (neutral (PhO\*) or cationic (PhO+\*) molecules, respectively), which are stabilized by delocalization of unpaired electrons around the aromatic ring (Rice-Evans et al., 1996; Bouayed et al., 2011a, b). Polyphenols have been reported to be more efficient than vitamin C, vitamin E and carotenoids (concentration ranges between high micromolar and low millimolar in human plasma and organs) against oxidative stress at tissue levels (Scalbert et al., 2002; Manach et al., 2004; Yang et al., 2008; Pandey and Rizvi, 2009; Bouayed and Bohn, 2010; Bouayed et al., 2011b, c).

Carotenoids: Among the various natural pigments, carotenoids comprise a large family of more than 700 structures (Britton et al., 2004) and are synthesized in plants and other photosynthetic organisms, as well as in some non-photosynthetic bacteria, fungi, algae, yeasts and moulds (Stahl and Sies, 2003, 2005). Most carotenoids can be derived from a 40-carbon basal structure, which includes a polyene chain contains 3 to 15 conjugated double bonds. The pattern of conjugated double bonds in the polyene backbone of carotenoids determines their light absorbing properties and influences the antioxidant activity of carotenoids (Goodwin, 1980; Britton et al., 1998). The best documented antioxidant action of carotenoids is their ability to quench singlet oxygen (Paiva and Russell, 1999) via physical or chemical quenching (Stahl and Sies, 2003). Food items rich in carotenoids includes: corn, yellow pepper, apricots, spinach, pumpkin and sweet potato, tomatoes and carrots (Biehler et al. 2012). According to their chemical composition carotenoids are categorized as either carotenes or xanthophylls (oxocarotenoids) (Olson and Krinsky, 1995). While the carotene group, such as  $\beta$ -carotene,  $\alpha$ -carotene and lycopene, composed only of carbon and hydrogen atoms, xanthophylls, such as zeaxanthin, lutein,  $\alpha$  and  $\beta$ -cryptoxanthin, carry at least one oxygen atom (Stahl and Sies, 2005; Chaudhary et al., 2010).

Minerals: Some minerals are essential for human nutrition (e.g., Fe, Cu, Se and Zn), others such as Cr, Cd, Ni, As and Pb (Hartwig, 1995; Valko et al., 2006), possibly through the formation of Reactive Oxygen Species (ROS) (Rojas et al., 1999; Linder, 2001). The trace elements essentially act as cofactors for antioxidant enzymes involved in the destruction of toxic free radicals produced in the body as a normal consequence of the metabolic processes. Three key trace elements whose roles in antioxidant defence are gradually gaining attention are zinc, selenium and iron. Apparently, Fe, Cu, Zn and Se are necessary to maintain genetic stability and nutritional well-being of humans and animals (Rojas et al., 1999). Recent studies suggest that ultra-trace elements, such as As and Ni, may also play a role both in animal and human nutrition (Rojas et al., 1999). In the last 20 years, a substantial body of evidence has been accumulated to support the role of zinc as a cellular antioxidant (Powell, 2000). Although zinc does not react directly with ROS, a number of indirect mechanisms have been described (Powell, 2000; DiSilvestro, 2000). One of the ways in which zinc acts as an antioxidant is through the induction of the metallothioneins, a group of low-molecular-weight amino acid residues, the production of which is induced by zinc in many tissues including the liver, gut and kidney. For instance, cadmium is present in spinach and cauliflower, while lead is found in Brussels sprouts and Chinese beets (3).

Table 4: ROS scavenging and detoxifying enzymes and reactions catalyzed

Enzymes	EC No.	Reaction catalysed	Localization (a)
Superoxide dismutase (SOD)	1.15.1.1	$O_2^- + O_2^- + 2H^+ \leftrightarrow 2H_2O_2 + O_2$	Cyt, Chl, Mit, Per
Catalase (CAT)	1.11.1.6	$2H_2O_2 \leftrightarrow O_2 + 2H_2O$	Mit, Per, Gly
Glutathione peroxidase(GPX)	1.11.1.12	$2\text{GSH+PUFA-OOH} \leftrightarrow \!\! \text{GSSG+PUFA+2H}_2\text{O}$	Cyt, Chl, Mit, ER
Glutathione S-transferases (GST)	2.5.1.18	RX+GSH↔HX+R-S-GSH*	Apo, Cyt, Chl, Mit, Nuc
Phospholipid-hydroperoxide	1.11.1.9	$2\text{GSH+PUFA-OOH} \ (\text{H}_2\text{O}_2) {\leftrightarrow} \text{GSSG+2H}_2\text{O}^\dagger$	Cyt, Chl, Mit, ER
glutathione peroxidase			
Ascorbate peroxidase (APX)	1.11.1.11	$AA+H_2O_2 \leftrightarrow DHA+2H_2O$	Cyt, Chl, Mit, Per
Guaiacol type peroxidase (POX)	1.11.1.7	$Donor + H_2O_2 {\longleftrightarrow} oxidized\ donor + 2H_2O^\dagger$	CW, Cyt, Mit, Vac
Monodehydroascorbate	1.6.5.4	$NADH + 2MDHA \leftrightarrow NAD^{+} + 2AA$	Cyt, Chl, Mit
reductase (MDHAR)			
Dehydroascorbate reductase (DHAR)	1.8.5.1	$2\text{GSH+DHA}{\leftrightarrow}\text{GSSG+AA}$	Cyt, Chl, Mit
Glutathione reductase (GR)	1.6.4.2	NADPH+GSSG↔NADP++2GSH	Cyt, Chl, Mit, Per

<sup>\*</sup>R may be an aliphatic, aromatic or heterocyclic group, X may be a sulfate, nitrite or halide group; †Reaction with  $H_2O_2$  is slow, (a) Gechev *et al.* (2006) is used as reference for localization of enzymes. The abbreviations are: Apo, apoplast, Cyt, cytosol; Chl, chloroplasts; CW, cell wall; ER, endoplasmatic reticulum; Gly, glyoxysomes; Mit, mitochondria; Nuc, nucleus; Per, peroxisomes; Vac, vacuole

#### ENZYMATIC ANTIOXIDANTS IN VEGTABLES

Detoxification constitutes the second line of defence against the maleficent effects of ROS. Therefore, once formed, the ROS must be detoxified as effectively as possible to minimize damage (Moller, 2001). Efficient destruction of ROS requires the action of several antioxidant enzymes acting in synchronicity with each other (Noctor and Foyer, 1998). Plants have also developed very efficient enzymatic antioxidant scavenging system to protect themselves against destructive ROS reactions (Mittler et al., 2004). In the cell, toxic effects of ROS are counteracted by several scavenging and detoxifying enzymes, such as superoxide dismutase (SOD), catalase (CAT), Ascorbate Peroxidase (APX), glutathione peroxidases (GPX), glutathione-S-transferases (GST), monodehydroascorbate reductases (MDHAR), dehydroascorbate reductases (DHAR) and Glutathione Reductases (GR) (4). While some of these enzymes are entirely dedicated to ROS homeostasis, others are involved also in other processes related to control of development, redox regulation of target proteins and detoxification reactions (Gechev et al., 2006). In cells, different scavenging enzymes encoded by the ROS network can be found in almost every subcellular compartment. Besides, each cellular compartment contains more than one enzymatic activity that detoxifies a particular ROS (Mittler et al., 2004). For example, the cytosol contains at least three different enzymatic activities that scavenge H<sub>2</sub>O<sub>2</sub> (Suzuki and Mittler, 2006). It is clear that the presence of different enzymes in various cellular compartments disclosures their significant role in ROS detoxification for the survival of the plant (Mittler et al., 2004). In Table 4, ROS scavenging enzymes, their abbreviations, EC numbers, catalyzed reactions and cellular localizations are given.

#### MECHANISM OF ANTIOXIDANT ACTION

The harmful action of ROS and free radicals is normally blocked by antioxidant substances, which scavenge the free radicals and detoxify the organism (Kumaran and Karunakaran, 2006). Antioxidants are compounds that can delay or inhibit the oxidation of lipid or other molecules by inhibiting the initiation or propagation of oxidizing chain reactions (Cakir *et al.*, 2006). All aerobic organisms have antioxidant defense systems (Cakir *et al.*, 2006). These systems are able to

scavenge free radicals and increase shelf life of processes foods by retarding the process of lipid peroxidation, the major cause of food and pharmaceutical deterioration (Halliwell, 1996; Gordon, 1996). Antioxidants can protect the human body from free radicals and ROS effects. They retard the progress of many chronic diseases as well as lipid peroxidation (Gulcin, 2007). Being enzymatic or non enzymatic species, antioxidant molecules are classified in different categories (Table 2). Both enzymatic (superoxide dismutase and catalase) and non-enzymatic (antocyanins and tochopherols) antioxidants are able to turn ROS into stable, harmless molecules. They are used in various applications such as pharmaceutical, food, cosmetic and chemical industries in order to act as preservative and extend efficiency and economical value of the products. Considering that antioxidant capacity of an organism is limited, excess production of various oxidizing compounds produces a condition called oxidative stress. Oxidative stress contributes to the pathogenesis of various diseases (e.g., cancers, neurodegenerative disorders and diabetes). Defense against oxidative stress through production of antioxidants and repair processes may constitute important allocations to somatic effort and is particularly relevant for species with low extrinsic mortality.

Antioxidants are major compounds that protect the quality of life by retarding the oxidation process through scavenging free radical produced during many natural events. Although their ultimate aim is removal of ROS, they may use different mechanism depending on their structure and site of action. Antioxidants are also able to act by up-regulating the expression of the genes encoding the antioxidant enzymes, repairing oxidative damage caused by radicals and increasing elimination of damaged molecules (Wood et al., 2006). The use of antioxidants in food industry is inevitable as they can increase shelf life and prevent oxidation. Synthetic antioxidants such as butylated hydroxytoluene (BHT), sodium benzoate and butylated hydroxyanisole (BHA) are widely used in food products. However, their use must be controlled due to possible hazards such as carcinogenicity and toxicity (Ito et al., 1983). Phenolic compounds from plant sources may act as antioxidants by scavenging lipid radicals. Over the last few years, an increasing interest in the search for naturally occurring antioxidants is ongoing. A large number of plants including fruits and vegetables are known as rich sources of antioxidant. The type of plant and its antioxidant activity depends entirely to the region that plant grows and natural vegetation present.

#### METHODS USED IN ANTIOXIDANT STUDIES: PRINCIPLE AND APPLICATIONS

A variety of *in-vitro* chemical methods are being used to determine the antioxidant activity of products and ingredients but questions regarding whether the results have any bearing on effectiveness in the human body are leading to development of additional methods that may be more appropriate for screening potential antioxidant ingredients. Several assays have been frequently used to estimate antioxidant capacities in fresh fruits and vegetables and their products and foods for clinical studies including 2,2-azinobis (3-ethyl-benzothiazoline-6-sulfonic acid) (ABTS) (Leong and Shui, 2002; Miller and Rice-Evans, 1997), 2,2-diphenyl-1-picrylhydrazyl (DPPH) (Brand-Williams et al., 1995; Gil et al., 2002), ferric reducing antioxidant power (FRAP) (Benzie and Strain, 1999; Guo et al., 2003; Jimenez-Escrig et al., 2001) and the oxygen radical absorption capacity (ORAC) (Cao et al., 1993; Ou et al., 2001; Prior et al., 2003). The ORAC assay is said to be more relevant because it utilizes a biologically relevant radical source (Prior et al., 2003). These techniques have shown different results among crop species and across laboratories. Ou et al. (2002) reported no correlation of antioxidant activity between the FRAP and ORAC techniques

among most of the 927 freeze-dried vegetable samples, whereas these methods revealed high correlation in blueberry fruit (Connor *et al.*, 2002). Similarly, Awika *et al.* (2003) observed high correlation between ABTS, DPPH and ORAC among sorghum and its products.

Various methods have been developed and applied in different systems but many available methods result in inconsistent results. There is no simple universal method by which antioxidant capacity can be assessed accurately and quantitatively. In this review article, the available methods are critically reviewed on the basis of the mechanisms and dynamics of antioxidant action and the methods are proposed to assess the capacity of radical scavenging and inhibition of lipid peroxidation both in vitro and in vivo. It is emphasized that the prevailing competition methods such as Oxygen Radical Absorption Capacity (ORAC) using a reference probe may be useful for assessing the capacity for scavenging free radicals but that such methods do not evaluate the characteristics of antioxidants and do not necessarily show the capacity to suppress the oxidation, that is, antioxidation. It is recommended that the capacity of antioxidant compounds and their mixtures for antioxidation should be assessed from their effect on the levels of plasma lipid peroxidation in vitro and biomarkers of oxidative stress in vivo.

#### ANTIOXIDANT ACTIVITY OF VEGETABLES

In search for sources of novel antioxidants and other important nutrients, a large number of plants have been extensively studied (Dasgupta and De, 2007; Indrayan *et al.*, 2005; Iqbal *et al.*, 2006; Joyeux *et al.*, 1995; Odhav *et al.*, 2007; Xin *et al.*, 2004) during last few years.

#### DPPH FREE RADICAL SCAVENGING ACTIVITY

According to their reducing/antioxidantive power the antioxidantive effect of these vegetables and leafy vegetables can be divided into four groups: (a) low (0-20%), moderate (20-40%), good (40-80%) and very good (80-100%).

The results summarized in Table 1 indicate that the vegetable interact with DPPH radicals and there by stabilize their hyper activity. The lowest groups of vegetables exhibiting antioxidant activity in the range of 0-20% are as follows: *Allium sativum*, *Capsicum annuum* Linn. Var. grossam (Willd.) *Momordica balsamina* L., *Solanum tuberosom* L. and *Spinacia oleraceae* Linn.

The average groups of vegetables exhibiting antioxidant activity in the range of 20-40% are as follows: Trigona foenum-graecum Linn., Lycopersicon esculentum, Rumex vesicarius Linn., Cucurbita pepo, Cucumis sativus, Rumex vesicarius Linn., Petroselinum crispum, Cucumis sativus and Raphanus sativus Linn. (Table 5).

The good groups of vegetables exhibiting antioxidant activity in the range of 40-80% are as follows: Brassica oleraceae Linn. Var. capitat, Anethum graveolens Linn., Brassica oleraceae Linn. Var. botrytis, Raphanus sativus Linn., Cucurbita maxima Duch., Lepidium meyenii, Cyamopsis tetragonoloba Linn., Lycopersicon esculentum, Capsicum annuum, Curcuma amada Roxb., Curcuma zedoaria, Macrolepiota mastoidea and Abelmoschus esculenus (Linn.) (Table 5).

The very good groups of vegetables exhibiting antioxidant activity in the range of 80-100% are as follows: Zinger officinale, Leucas aspera, Corchorus olitorius, Crotalaria ochroleuca, Solanum scabrum, Cleome gynandra, Achras Linn., Amaranthus sapotagangetica, Amaranthushybridus, Amaranthusspinosus, AsystasiaLeucas aspera, ochroleuca, SolanumCorchorus olitorius, Crotalariascabrum, Cleome gynandra, Achras sapota Linn., Amaranthus dubius, Asystasia gangetica, Amaranthus hybridus, Amaranthus spinosus, Bidens pilosa, Chenopodium album, Centella asiatica, Cleome monophylla, Ceratotheca triloba, Justicia flava, Momordica balsamina, Oxygonum sinuatum, Portulaca oleracea, Physalis viscose, Solanum nigru, Senna occidentalis, Olax psittacorum, Cassia tora, Ipomoea cairica, Leucas aspera, Commelina benghalensis (Table 5).

Vegetables with highest anticancer activity are Allium cepa, Curcuma amada, Curcuma zedoaria, Citrus limon, Brassica oleraceae, Lycopersicon esculentum etc. and the Cucumis sativus is with low level of anticancer activity (La Vecchia and Tavani, 1998; Nahak and Sahu, 2011; Kaur and Kapoor, 2004; Rao et al., 2004). Green leafy vegetables are rich sources of antioxidant vitamins (Gupta et al., 2005). The ascorbic acid, total carotene,  $\beta$ -carotene and total phenolic content of the green leafy vegetables, viz., Amaranthus sp., Centella asiatica, Murraya koenigii and Trigonella foenum graecum etc., are showing highest antioxidant activity among leafy vegetables (Gupta and Prakash, 2009). Spinach and kale are also rich sources of carotenoids and polyphenols. Spinach has an exceptionally high total polyphenol and flavonoid content. The high level of polyphenol acids and flavonoids in spinach leaves influences the high antioxidant activity (Ligor et al., 2012). Majority of leafy vegetables like Brassica, Coriandrum, Carrot, mint, Curry leaf, Radish, Methy contains phytochemicals which shows antioxidant activities include vitamins (A, C, E and K), carotenoid, terpinoid, flavonoid, polyphenols, saponins, enzymes and minerals. All these compounds prevent cancers and other diseases (Govind and Madhuri, 2011).

#### FRAR VALUE

The FRAP assay determines the capacity of antioxidants as reductants in a redox-linked colorimetric reaction of the reduction of Fe<sup>3+</sup>-2,4,6-tripyridyl-S-triazine to a blue-coloured Fe<sup>2+</sup> complex at low pH that is measured spectrophotometrically at 593 nm (Benzie and Strain, 1996). The extracts were incubated at room temperature with the FRAP reagent and the absorbance recorded after 1 h (Fasahat et al., 2012). The reducing power is expressed as µmol FeSO<sub>4</sub> g<sup>-1</sup>. Here in this review some of the vegetables showing very good antioxidant activity in terms of FRAP (FRAR (Mg<sup>-1</sup>) values and these are as follows: Daucus carota L. Var. sativa DC. (0.61), Cucumis sativus (1.69), Cucumis melo var. flexuosus Lagenaria siceraria (6.57), Luffa cylindrical L. (2.93), Luffa acutangula (L.) Roxb (2.58), Trichosanthes dioica Roxb. (3.78), Carica papaya L. (4.63), Beta vulgaris L. (5.45) Citrus limon L., Aegle marmelos Correa ex roxb. (6.36), Solanum melongena L. (3.69) and Musa paradisica L. (unripe) (13.18) (Table 5).

#### IC<sub>50</sub> VALUE

IC<sub>50</sub> value is defined as the concentration of substrate that 50% loss of DPPH\* activity and was calculated by linear regression method of plots of the percentage of antiradical activity against the concentration of the tested compounds. The results summarized in Table 3 and 4 indicate that the IC<sub>50</sub> values of *Brassica oleraceae* (Broccoli) with 7.53 mg mL<sup>-1</sup>, *Musa canvendish* (10.93 mg mL<sup>-1</sup>), *Zinger officinale* (1.8 mg mL<sup>-1</sup>) and *Phoenix dactylifera* (1.65 mg mL<sup>-1</sup>), *Apium nodiflorum* (0.07 mg mL<sup>-1</sup>), *Foeniculum vulgare* (2.75 mg mL<sup>-1</sup>), *Montia fontana* (1.49 mg mL<sup>-1</sup>) and *Anethum graveolens* Linn. (3.31 mg mL<sup>-1</sup>) showed significant lower IC<sub>50</sub> values which refer the good antioxidant potential sources (Table 5).

Table 5: Antioxidant activity of some vegetables

Botanical names with	Active components		
common names family	responsible for antioxidant activity	Antioxidant values	References
Daucus carota L.	Proteins, amino acids,	$210 (ORAC \ units \ 100 \ g^{-1})$	Chatatikun and
Var. sativa DC. carrot apiaceae	vitamin (A, B and C)	33.0%(DPPH)	Chiabchalard (2013),
		42.2%(ABTS)	Chanda $et\ al.\ (2013)$
		$0.61(\mathrm{FRAR}\ (\mathrm{Mg^{-1}})$	
Coriandrum sativum Linn.	Essential oil, vitamin C,	25.0 %(DPPH)	Gacche $et\ al.\ (2010)$
Coriander apiaceae	carotene, borneal,		
	limonene and $\alpha$ pinene		
Anethum graveolens	Carvone, limonene, $\alpha$ -phellandrene,	51.6 %(DPPH)	Gacche $et\ al.\ (2010)$
Linn.: dill apiaceae	dill ether and myristicin		
Caralluma tuberculata			
N.E. brown: chungah	Luteolin 4'-beta-D-glucopyranosyl-	$695.7(EC_{50})$	Khanzadi (2011)
Asclepiadaceae	(2>1)-alpha-L-rhamnopyranoside		
Brassica oleraceae			~
Linn. Var. capitat	Ascorbigen, vitamins (A,B and C),	42.4 %(DPPH)	Gacche <i>et al</i> . (2010)
Cabbage brassicaceae	sulphoraphane, dithiolthiones,		
	isothiocyanate		
Brassica oleraceae	Ascorbigen, vitamins (A,B and C),	67.2 %(DPPH)	Gacche <i>et al</i> . (2010)
Linn. Var. botrytis	sulphoraphane, dithiolthiones,		
Cauliflower Brassicaceae	isothiocyanate		
Brassica oleracea: Broccoli	Hydroxycinamic acid esters,	$880(ORAC \text{ units } 100 \text{ g}^{-1})$	Sosnowska et al. (2006)
Brassicaceae	1,2-disinapoylgentiobiose and	8.9 (μmol of Trolox g <sup>-1</sup> )	Ou et al. (2002)
1-sinapoyl-2-feruloylgentiobio		$7.52~{ m m}~{ m m}{ m L}^{-1}~({ m IC}_{50})$	and Piao et al. (2005)
Raphanus sativus Linn.	Vitamin C,	62.20%DPPH	Umamaheswari <i>et al.</i> (2012
Brassicaceae	anthocyanin, glucosinolates		
Cucurbita pepo: Zucchini	Alkaloids momordicine I,	12.19%	Hamissou <i>et al.</i> (2013)
Cucurbitaceae	momordicine II and cucurbitacine B		
Cucumis sativus: Cucumber	Luteolin-8-C-beta-D-glucopyranoside	60 ORAC units 100 g <sup>-1</sup> ),	Azeez <i>et al.</i> (2012)
Cucurbitaceae	(orientin), luteolin-6-C-beta-	28.32% (DPPH) and	
	Dglucopyranoside (isoorientin)	$178.2  ({ m IC}_{50}  { m mg mL}^{-1}),$	
		$1.69  (\mathrm{FRAR}  (\mathrm{Mg}^{-1})$	
Lactuca sativa: Lettuce	Vitamin (A,C, E, B1, B2, B3),	92.62% (DPPH)	Azeez et al. (2012)
Asteraceae	$\beta$ -carotene, malic acid,		
	linoic acid, fiber, pectin,		
	amino acids and minerals		
Cucumis melo var. flexuosus	Vitamin A, thiamin, riboflavin,	$22.23~(IC_{50}~mg~mL^{-1})$	Qusti et al. (2010)
Snake Cucumber	niacin, folate, ascorbic acid		
Cucurbitaceae	and proteins		
Lagenaria siceraria:	Vitamin-B complex, choline,	$1770  (ORAC  units  100  g^{-1})$	Chanda et $al.$ (2013)
Bottle Gourd Cucurbitaceae	vitamin-C, $\beta$ -carotene,	$6.57  (FRAR  (Mg^{-1})$	
	Cucurbitacins, fibres and polyphenol		
Luffa cylindrical L.	Luteolin-7-O-beta-D-glucuronide		
Chinese okra	methyl ester	980 (ORAC units $100 \text{ g}^{-1}$ )	Chanda $et\ al.\ (2013)$
Cucurbitaceae		$2.93  (FRAR  (Mg^{-1})$	Chanda $et\ al.\ (2013)$
Luffa acutangula (L.)	Carotene, fat, protein,	890 (ORAC units $100~\mathrm{g}^{-1}$ )	
Roxb: Silk Gourd	phytin, aminoacids, alanine,	$2.58  (FRAR  (Mg^{-1})$	
Cucurbitaceae	arginine, cystine, glutamicacid,		
	glycine, hydroxyproline,		
	leucine, lectin, serine,		
	tryptophan, pipecolic acid		

Tab	le 5:	Conti	nue

Botanical names with	Active components		
common names family	responsible for antioxidant activity	Antioxidant values	References
$Trichosanthes\ dioica\ Roxb$ :	Vitamin A, vitamin C,	840 (ORAC units $100~\mathrm{g}^{-1}$ )	Chanda $et\ al.\ (2013)$
Pointed gourd Cucurbitaceae	tannins, sapponin	$3.78  (FRAR  (Mg^{-1})$	
Cucurbita maxima Duch.:	Vitamin C and Phenols	51.6% (DPPH)	Gacche $et\ al.\ (2010)$
Bottle gourd Cucurbitaceae			
$Terminalia\ catappa\ { m L}.$	Alkaloid, Reducing sugar,	35.3% (DPPH)	Chyau et al. (2006)
Combretaceae	Resins, Steroids, Tannins, Saponins		
Carica papaya L.: Papaya	$\alpha$ -tocopherol, ascorbic acid,	$4.63  (FRAR  (Mg^{-1})$	Chanda $et\ al.\ (2013)$
Caricaceae	beta carotene, fl avonoids,		
	vitamin B1 and niacin		
Beta vulgaris L.: Beet	Vitamins (A, B and C),	$5.45  (FRAR  (Mg^{-1})$	Chanda $et\ al.\ (2013)$
Chenopodiaceae	iodine and betanin		
Spinacia oleraceae Linn.:	Vitamin (A, E and C),	20.4% (DPPH)	Gacche et al. $(2010)$
Spinach Chenopodiaceae	folic acids and minerals		
Lepidium meyenii: Maca	Alkaloids, protein, quercitin, saponins,	71.38% (DPPH)	Sandoval et al. (2002)
Cruciferae	sitosterols, steroids, stigmasterol,		
	tannins, uridine, vitamin		
	(B1, B12, C, E and K) and zinc		
Trigona foenum-graecum	Choline, trigonelline, saponin,	25.7 %(DPPH)	Gacche $et\ al.\ (2010)$
Linn.: Fenugreek Fabaceae	amino acids, vitamins and quercetin		
$Cyamops is\ tetragonoloba$	Flavonoids, tannins, glutamic,	44.6% (DPPH)	Gacche $et\ al.\ (2010)$
Linn.: Cluserbean Fabaceae	arginine, aspartic and leucine		
Bauhinia variegata Linn.	Protein, flavonoids, fatty oil-containing	36.01% (DPPH)	Sharma $et\ al.\ (2011)$
Mountain ebony Fabaceae	oleic acid, linoleic acid, palmitic		
	acid and stearic acid, dihydroxy		
	Kaempferol-3-glucoside,		
	lupeol and beta-sitosterol		
Portulaca oleracea Linn.:	Alkaloids, coumarins, flavonoids,	950.3 ( $EC_{50}$ )	Khattak (2011)
Purslane Portulacaceae	cardiac glycosides, anthraquinone		
	glycosides, alanine, catechol, saponins,		
	tannins, organic acids, glutathione,		
	glutamic acid and aspartic acid		
Citrus limon L.	Vitamin C, flavonoid, flavones,	$1.88  (FRAR  (Mg^{-1})$	Chanda <i>et al.</i> (2013)
Rutaceae	limonoid, limonene (terpenoid),		
	nobiletin, tangeretin, glucarate		
Aegle marmelos	Eugenol, lupeol, cineole,	6.36 (FRAR (Mg <sup>-1</sup> )	Chanda <i>et al.</i> (2013)
Correa ex roxb. Rutaceae	citronellal, aegenol, luvangetin,		
	psoralen and cuminaldehyde		
Lycopersicon esculentum:	Vitamins (A, B and C),	195 (ORAC units 100 g <sup>-1</sup> )	Azeez et al. (2012)
Tomato (ripe) Solanaceae	essential amino acids and lycopene	35.0% (DPPH)	,
Lycopersicon esculentum:	Vitamins (A, B and C),	41.10% (DPPH)	Azeez et al. (2012)
Tomato (unripe) Solanaceae	essential amino acids and lycopene	,/	- ()
Capsicum annuum:		42.92% (DPPH)	Azeez et al. (2012)
Green pepper Solanaceae			
Solanum tuberosom L.:	Solanine, solasodine, lutin,	300 (ORAC units 100 g <sup>-1</sup> )	Velioglu et al. (1998)
Potato Solanaceae	vitamin (A and C) and glucarate	80.0 % (DPPH)	6-4 01 41. (1000)
	·	·	Hamissou et al. (2013)
			111111111111111111111111111111111111111
Momordica charantia L. Cucurbitaceae	Linolenic and Palmitic acids, momrdin and vitamins	82.05%	Hamissou et al. (2013)

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Botanical names with	Active components		
common names family	responsible for antioxidant activity	Antioxidant values	References
Momordica balsamina L.	Alkaloids momordicine I,	8.60% (DPPH)	Chanda et al. $(2013)$
Cucurbitaceae	momordicine II and cucurbitacine B		
Solanum melongena L.:	Solasodine and glucarate	$400 (ORAC units 100 g^{-1})$	Chanda et al. (2013)
Egg plant Solanaceae		$3.69(FRAR (Mg^{-1})$	
Capsicum frutescens (ripe)	Carotene, vitamin C,	67.85 %(DPPH)	Azeez et al. (2012)
Solanaceae	glutathione, flavonoids		
Allium sativum: Garlic	Saponins, tannins, sulfurous	$1939 (ORAC units 100 g^{-1})$	Qusti et al. (2010)
Liliaceae	compounds, prostaglandins,	4.32%(DPPH) and	
	alkaloids, volatile oils and allicin	$24.17~(IC50~mg~mL^{-1})$	
Chenopodium album Linn.	Amino acids (leucin,	$454.7(EC_{50})$	Khattak (2011)
White goose foot Meliaceae	isoleucin, lycine), vitamin C		
Musa paradisica L. (unripe):		380 (ORAC units 100 g <sup>-1</sup> )	Chanda et al. (2013)
Banana Musaceae		$13.18  (\mathrm{FRAR}  (\mathrm{Mg}^{-1})$	
Moringa oleifera Lam.:	Alkaloids, flavonoids,	$1.75  (FRAR  (Mg^{-1}),$	Chanda et al. (2013)
Drumstick Moringaceae	anthocyanins, cosmetic oils	$376.0~(\mathrm{EC_{50}})$	and Khattak (2011)
	Leaf: Quercetin-3-0-glucoside,		
	quercetin-3-0-(6"-malonyl-glucoside),		
	kaempferol-3-0(6"-malonyl-glucoside),		
	3-caffeoylquinic and		
	5-caffeoylquinic acid		
Rumex vesicarius Linn.	Flavonoids (vitexin, isovitexin,	32.97% (DPPH)	El-Bakry <i>et al.</i> (2012)
Bladder-Dock Polygonaceae	orientin and isorientin), quinones,		
	carotenoids, vitamins-C proteins,		
	tannins, saponins, triterepenoids		
	and organic acids and minerals		
Zinger officinale: Ginger	Camhene, gingerol, zingiberene,	$1770  (ORAC  units  100  g^{-1})$	Azeez et al. (2012)
Zingiberaceae	borneol, cineol, cucumins and proteins	88.29% (DPPH)	
		and 1.8 (IC $_{50}~mg~mL^{-1})$	
Curcuma amada Roxb.:	Curcumin, phytosterol and	52.61% (DPPH)	Nahak and Sahu (2011)
Mango ginger Zingiberaceae	azulenogenic oil containing pinene,		
	camphor, curcumene and		
	ar-turmerone Caffeic, gentisic,		
	ferulic, gallic, cinnamic		
Curcuma zedoaria	Vitamin C and curcumin	63.27% (DPPH)	Nahak and Sahu (2011)
Zingiberaceae			
Allium cepa: Onion	Diallyl disulphide, allicin,	450 (ORAC units 100 g <sup>-1</sup> )	Velioglu et al. (1998)
Liliaceae	allin, quercetin antioxidant	90.2% DPPH	
	flavonoid, vitamin (C and E)		
Musa Cavendish: Banana	Vitamin C, vitamin E,	$10.93~((IC_{50}~mg~mL^{-1}~)$	Qusti et al. (2010)
Musacea	$\beta$ -carotene, gallocatechin		
	and dopamine		
Musa Cavendish:	linoleic acid	75.3% DPPH	Mokbel and Hashinaga
Banana peel Musacea			(2005)
Phoenix dactylifera	Phenolic compounds,	$1.65(\rm{IC_{50}\;mg\;mL^{-1}})$	Qusti et al. (2010)
Arecaceae	essential minerals, fiber and proteins		
Abelmoschus esculenus (Linn.)	Carotene, vitamins	43.8% (DPPH)	Gacche et al. $(2010)$
Lady's finger Malvaceae	(B and C) and amino acids		

Table 5: Contin	านe

Table 5: Continue  Botanical names with	Active components		
	•	Antioxidant values	References
common names family	responsible for antioxidant activity		
Nelumbo nucifera Gaertn.:	Alkaloids, vitamins,	92.8% (DPPH)	Choe et al. (2010)
Lotus leaf	quercetin flavonoid, tannic		
	acid and oligomeric procyanidines		
Hordeum vulgare leaf	Protein, minerals and enzymes	79.5% (DPPH)	Choe et al. (2010)
Leucas aspera: Thumbai	Alkaloids, flavonoids, terpenoids,	83.67%(DPPH)	Das <i>et al.</i> (2011) and
Lamiaceae	steroids, tannins, phlobatannins,		Rahman et al.
	saponins, glycosides, diterpenes,		(2008)
	tannins, saponins, sterols, oleic,		
	linoleic, palmitic, stearic and oleanolic		
Petroselinum crispum: Parsley	Flavonols, flavones,	40.1% (DPPH)	Kamel (2013),
Umbelliferae	flavanones, catechins,		Fatemeh $et\ al.\ (2011)$
	anthocyanidins and isoflavones		
Anethum graveolens: Dill	Tannins, terpenoids,	48.14% (DPPH)	Kamel (2013),
Umbelliferae	cardiac glycosides and flavonoids		Jana and Shekhawat (2010)
$Macrolepiota\ mastoidea$	Flavonoids, phenolic acids,	70.87% (DPPH)	Shirmila Jose and
(fr.) Singer: Wild Mosroom	fiber tannins and minerals.		Radhamany (2013)
Quercus infectoria: Olivier	Gallic acid and ellagic acid	$0.25~{\rm mg~mL^{-1}~(IC_{50})}$	Umachigi et al. (2013),
Fagaceae		$152.91 \; nmol \; g^{-1} \; (TAC)$	Umachigi et al. (2008)
Bambusa arundinacea:	Oxalic acid, amino acids,	51.41% (DPPH))	Sandhiya $et\ al.\ (2013)$
Spiny bamboo Poaceae	proteolytic enzyme, nuclease, urease		
Cassia sophera L.: Kasondi	Flavonoids, resins and phenols	$15.42~IC_{50}~(\mu g~mL^{-1})$	Rahman <i>et al</i> . (2008)
Caesalpiniaceae			
Corchorus olitorius:	Vitamin (B1, B2, C), carotinoids,	90.4% (DPPH)	Elias et al. (2012)
Jute mallow	calcium, potassium,		
Oceanopapayeraceae	proteins and dietary fibers		
Crotalaria ochroleuca:		86.9% (DPPH)	Elias <i>et al.</i> (2012)
Slender leaf Fabaceae			
Solanum scabrum:	Vitamin (A and C), riboflavin,		
Black nightshade Solanaceae	folic acid, carotenes, protein, iron,	92.8% (DPPH)	Elias et al. (2012),
	Iodine, Zinc and Selenium		Musyimi et al. (2012)
Cleome gynandra:	Alkaloids, flavonoids, tannins,	87.8% (DPPH)	Elias <i>et al.</i> (2012)
Spider plant Capparaceae	saponin, anthroquinons,		
	phlobatanins and steroids		
Amaranthus dubius:		78% (DPPH)	Odhav et al. (2007)
Bladder campion			
Amaranthaceae			
Asystasia gangetica:	Alkaloids, tannins,	84% (DPPH)	Odhav et al., 2007
Chinese violet Acanthaceae	steroidal aglycans,		
	saponins, flavonoids,		
	terpenoids and minerals.		
Amaranthus hybridus:	Polyphenols, carotenoids,	90% (DPPH)	Odhav et al. (2007)
Red amaranth	flavonoids, phenolics,	,	, ,
Amarantaceae	vitamins (C and E)		
Amaranthus spinosus:	Alkaloids, steroids, bita-steroids,	88% (DPPH)	Odhav et al. (2007)
Spiny amaranth	quercetin, betanin, amarathin,	` '	` /
Amaranthaceae	beta-cyanin and bita-carotene		
Bidens pilosa: Spanish needle	Aliphatics, flavonoids, terpenoids,	88% (DPPH)	Odhav et al. (2007)
Asteraceae	phenylpropanoids, aromatics,	Silva et al. (2011)	(====-/
	porphyrins and other compounds		

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Botanical names with	Active components		
common names family	responsible for antioxidant activity	Antioxidant values	References
Chenopodium album:	Flavonoids, lignins and tannins	82%(DPPH)	Odhav $et\ al.\ (2007)$ and
White goosefoot			Vermerris and Nicholson (2006
Amaranthaceae			
Centella asiatica: Centella	Asiatic acid, madecassic acid,	88%(DPPH)	Odhav et al. (2007),
Apiaceae	asiaticoside and madecassoside		Ling (2004) and Ling et al. (2000)
Cleome monophylla:	Flavonoids, alkaloids	84%(DPPH)	Odhav et al. (2007)
Spindle pod Capparidaceae	and triterpenoidal		
Ceratotheca triloba:	Mucilage, phenolic compounds,	84%(DPPH)	Odhav et al. (2007)
Wild foxglove Pedaliaceae	lipids, flavonoids, tannins,		
	saponins and fixed oils		
Emex australis: Double gee	Flavonoids,	78%(DPPH)	Odhav et al. (2007)
Polygonaceae	alkaloids and terpenoids		
Galinsoga parviflora:	Flavonoids, alkaloids steroids,	76%(DPPH)	Odhav et al. (2007)
Potato weed Asteraceae	terpenes and saponins		
Justicia flava: Yellow justicia	Flavonoids,	96%(DPPH)	Odhav et al. (2007)
Acanthaceae	alkaloids and triterpenoidal		
Momordica balsamina:	Resins, alkaloids, flavonoids,	94%(DPPH)	Odhav et al. (2007)
African cucumber	glycosides, steroids, terpenes,		
Cucurbitaceae	cardiac glycoside and saponins		
Oxygonum sinuatum:	Protein, dietary fibers and minerals.	92%(DPPH)	Odhav et al. (2007)
Dubbeltjie Polygonaceae			
Portulaca oleracea: Purslane	Vitamin (A, B and C),	96%(DPPH)	Odhav et al. (2007)
Portulacaceae	carotenoids, betalaine,		
	betacyanin and minerals		
Physalis viscose: Gooseberry	Carotenoids, vitamin C, minerals,	82%(DPPH)	Odhav et al. (2007)
Solanaceae	28-hydroxywithanolide, withanolides,		
	phygrine, kaempferol and quercetin		
Solanum nigrum:	Gallic acid, catechin, protocatechuic	92%(DPPH)	Odhav et al. (2007)
Black Nightshade	acid (PCA), cafieic acid, epicatechin,		
Solanaceae	rutin and naringenin		
Senna occidentalis:	Chrysarobin	82%(DPPH)	Odhav et al. (2007)
Negro coffee Fabaceae	(1,8-dihydroxy-3-methyl-9-anthrone)	` '	` ,
	and N-methylmorpholine		
Bauhinia purpurea:	Linolenic acid, oleic fatty	79.25%(DPPH)	Sahu <i>et al</i> . (2013)
Butterfly tree Fabaceae	acid and myristic acid.	, ,	, ,
Cassia tora: The Sickle	Chrysophanol, aloe-emodin,	83.76%(DPPH)	Sahu et al. (2013),
Senna Caesalpinaceae	rhein, glucose, 1-stachydnine,	(/	Vedpriya Arya
	amino acids, fatty acids, d-mannitol,		and Vaday (2011)
	β-sitosterol, myricyl alcohol, trigonellir	ne.	
	choline. Ononitol monohydrate	20,	
Ipomoea cairica	Alkaloids, tannins, phenolic	88.21%(DPPH)	Sahu <i>et al</i> . (2013)
Palmate-leaved	compounds, proteins and amino acid,	\ * * * * * * * * * * * * * * * * * *	
Morning Glory	terpinoids and sterols and saponins		
Convolvulaceae	terpinorus and seerois and supoinis		
Raphanus sativus Linn.:	Triterpenes, alklaoids,	22.06%DPPH	Umamaheswari et al. (2012)
Radish Brassicaceae	flavanoids, tannins,	22.00/eDilli	Smanareswarres ws. (2012)
Tradisti Di assicateae	saponins and coumarins		

Table 5: Continue

Botanical names with	Active components		
common names family	responsible for antioxidant activity	Antioxidant values	References
Commelina benghalensis:	Balsams, resin, tannins, 86.47%(DPPH)		Sahu <i>et al</i> . (2013)
Day Flower	flavonoids, phenols		
Commelinaceae	and volatile oils		
Leucas aspera: Thumba	Triterpenoids, oleanolic acid,	96.2%(DPPH)	Sahu <i>et al.</i> (2013)
Lamiaceae	ursolic acid and b-sitosterol, nicotine		
Sauropus androgynus:	Ascorbic acid, alpha-tocopherol	54.36%(DPPH)	Nahak and Sahu (2010)
Multi-vitamin	(vitamin E) and		
Euphorbiaceae	beta-carotene (vitamin A)		
Azadirachta indica: Neem	Alkaloids, flavonoids, terpinoids,	50.48%(DPPH)	Nahak and Sahu (2010)
Meliaceae	tannins, saponins and glycosides		
Ocimum basilicum:	Alkaloids, phenols, steroids,	70.76%(DPPH)	Nahak <i>et al.</i> (2011)
Sweet basil Lamiaceae	eugenol, eugenal, carvaled, methyl		
	chavicol, limatrol and Caryophylline		
Apium nodiflorum:	Limonene, p-cymene,	$0.07 (mg \ mL^{-1} \ EC_{50})$	Alpinar et al. $(2009)$
Fool's-water-cress	myristicine and $\beta$ -pinene		
Apiaceae			
Foeniculum vulgare: Fennel	Caffeic acid, p-Coumaric,	$2.75 ({ m mg~mL^{-1}EC_{50}})$	Alpinar et al. $(2009)$
Apiaceae	Ferulic acid-7-o glucoside,		
	Quercetin-7-o glucoside, Ferulic acid		
	1,5 Dicaffeoylquinic acid, Hesperidin,		
	Cinnamic acid, Rosmarenic acid,		
	Quercetin and Apigenin		
Montia Fontana: Blinks	Vitamin C, macro and micro	$1.49~(mg~mL^{-1}~EC_{50})$	Alpinar $et\ al.\ (2009)$
Montiaceae	minerals and organic acids		

#### ORAC VALUE

Based on widely reported rich antioxidant activities of vegetables, the scientists at United States Department of Agriculture (USDA) have developed a rating scale that measures the antioxidant content of various vegetables. The scale is called Oxygen Radical Absorbance Capacity (ORAC), which stands for oxygen radical absorbance capacity (Table 5). Ou et al. (2002) reported that green pepper, spinach, purple onion, broccoli, beet and cauliflower are the leading sources of antioxidant activities against the peroxyl radicals. Cao et al. (1996) reported that garlic had the highest antioxidant activity (µmol of Trolox equiv g<sup>-1</sup>) against peroxyl radicals Allium sativum (1939) followed by kale (1770), spinach (1260), Brussels sprouts, alfalfa sprouts, broccoli flowers, beets, red bell pepper, onion, corn, eggplant (980-390), cauliflower, potato, sweet potato, cabbage, leaf lettuce, string bean, carrot, yellow squash, iceberg lettuce, celery and cucumber (380-50). Kale had the highest antioxidant activity against hydroxyl radicals followed by Brussels sprouts, alfalfa sprouts, beets, spinach, broccoli flowers and the others. Lee et al. (2007) reported that among various cruciferous vegetables studied, red cabbage had the highest radical scavenging activity followed by Chinese white cabbage, green cabbage and mustard cabbage (Table 5).

# CORRELATION BETWEEN ANTIOXIDANT ACTIVITY AND TOTAL PHENOLIC CONTENT

It is generally believed that plants which are having more phenolic content show good antioxidant activity that is there is direct correlation between total phenol content and antioxidant activity (Qusti et al., 2010; Zhou and Yu, 2006). Typically phenols that possess antioxidant activity

are known to be phenolic acids and flavonoid, the major classes of phenolic compounds occurring widely in the kingdom especially in fruits and vegetables (Wojdylo et al., 2007). The amount of total phenols and antioxidanta are, by and large, with certain expections, within the range of values reported by several workers (Chu et al., 2000; Lee, 1992; Anese et al., 1999; Pellegrini et al., 2003) but they are lower than those reported by (Kaur and Kapoor, 2001, 2004; Kahkonen et al., 1999; Prior and Cao, 2000). It has been observed in some cases having no correlation between antioxidant activity and total phenolic content in extracts of some vegetables such as chick pea, drum stick, radish, mustard, purslane, white goose fruits, mountain abony, Caralluma, Carrot, Terminalia cattapa, Banana (unripe) determined by squared regression co-efficient (R2). These plants showed high phenol contents but comparatively low DPPH activity (Khattak, 2011; Chanda et al., 2013). Environmental factors such as differences in light, season, climate and temperature conditions on the one hand and production, optimal extraction factors and genotype on the other may have contributed to the differences in total phenols and antioxidant activities of various vegetables (Kalt, 2005; Rababah et al., 2010). It can be stated that phenolic content of the plant may be a good indicator of its antioxidant capacity (Govind and Madhuri, 2011).

#### CONCLUSION

Regardless of these advantages, unfortunately the majority of traditional vegetable plants are usually uncultivated and underutilized. It is critical to create awareness regarding diet related health benefits of these neglected precious crops. Further, with reference to food security there is a need to find out every potential source of safe and health-promoting nutrients. Findings of the study indicate that all the studied plants are excellent sources of micronutrients and free radical scavenging activity. The consumption of these traditional vegetables may have many beneficial health attributes. These data may also be helpful in allowing better food choices and improvement in nutritional and health status.

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