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Research Article Phytochemical Screening of Root Extract of *Momordica boivinii* and Isolation of Two Steroids

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Abstract

Background and Objective: Momordica boivinii (M. bovinii) is one of the plant species widely used in traditional medicine in Sidama zone, Southern part of Ethiopia, for treatment of several human illnesses but their chemical constituents are not studied before. Therefore, the aim of the present study was to conduct phytochemical screening tests on extracts of the root of this plant species and also to isolate compounds from the extracts. Materials and Methods: The root material of M. boivinii was extracted with four solvent systems [n-hexane, dichloromethane, dichloromethane/methanol (50:50% by volume) and methanol] using maceration technique and sequential extraction approach. All the extracts were subjected to phytochemical screening tests following standard procedures reported in literatures. Finally, the dichloromethane/methanol (50:50% by volume) extract was subjected to column chromatographic separation. Results: The yields of the extracts were 1.1, 1.8, 7.5 and 5.4 g for n-hexane, dichloromethane, dichloromethane/methanol (50:50% by volume) and methanol, respectively. The preliminary phytochemical analysis revealed the presence of alkaloids, saponins, flavonoids, phenols, steroids, glycosides, terpenoids and tannins in the crude ectracts of dichloromethane and dichloromethane:methanol (50:50 by volume). The crude extract of n-hexane showed the presence of alkaloids, saponins, steroids, glycosides and terpenoids. However, flavonoids, phenols and tannins were not detected. On the other hand, the crude extract of methanol revealed the presence of secondary metabolites listed above except tannins. Column chromatographic separation of the dichloromethane/methanol (50:50% by volume) extract afforded two compounds (labeled as compound MB-1 and MBC-1). The structures of the compounds were elucidated to be stigmasterol and methyl ester of betulinic acid-3-trans-caffeate based spectroscopic (IR, ¹H-NMR and ¹³C-NMR) data and comparison with literature reports. This is the first report of isolation of the compounds from *M. boivinii*. Conclusion: The findings of the study validate the use of the plant in traditional medicine. However, biological activity test is recommended to get comprehensive information about the potential of the plant as source of modern drugs.

Key words: Momordica boivinii, sequential extraction, phytochemical screening, steroids, stigmasterol, betulinic acid-3-trans-caffeate

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Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

The genus *Momordica* consists of approximately 825 species of annual and perennial plants that are distributed in tropical areas such as East Africa, Caribbean and Southern America¹. The species in the genus of *Momordica* are widely used to treat several alignments in different parts of the world due to the presence of variety of phytoconstituents. Different plant parts (leaves, roots and fruits) of the species are used in traditional medicinal uses in areas where these parts are available in abundance. For instance, leaf decoction has been used as anti-fungal, anti-inflammatory, anti-malarial, anti-parasitic, anti-septic, anti-tumor, antidotal, antipyretic tonic, appetizing, antibilious, carminatives, digestive stimulant, febrifuge, menstrual stimulator, blood circulation, immunity, control fever, blood impurities, liver diseases, skin ailments, vermifuge, purgative, colic, topically for sores, wounds, infections, for worms and parasites; as an antiviral for measles, hepatitis, laxative and in addition, it has been effectively used to treat diabetes and cancer²⁻⁷.

One of the *Momordica* species that is widely found in East Africa is *Momordica boivinii* (*M. bovinii*) (Fig. 1). It is distributed in Botswana, Ethiopia, Kenya, KwaZulu-Natal, Malawi, Mozambique, Namibia, Northern Provinces, Somalia, Swaziland, Tanzania, Uganda, Zambia and Zimbabwe⁸. Though there are no scientific research reports (studies), it is well known that its different parts are widely used in traditional medicines for treatment of human diseases in countries where the plant is available. Its roots and leaves are used by Midzichenda tribes (Kenya) to treat spiritual ailments⁹. A report showed that stems and leaves of *M. boivinii* are used in the treatment of stomach problem¹⁰. Taking the plant with *Cissus faucicola* and other herbs has also been reported to be used in the treatment of oestrogen sufferings¹¹. Solution of aqueous extract of this plant has also been reported to show anti-HIV activity¹². In Ethiopia, the plant is also used in traditional medicine. For instance, its leaf and fruit are used to treat pneumonia¹³. Fresh roots and leaves are reported to be effective in the treatment of gonorrhea and intestinal parasites¹⁴. Its leaves is mixed with vegetables used as an appetite stimulant; fruit juice of *M. boivinii* mixed with *Allium sativum* (locally known as nech shunchurt) and chewing of its leaves also used to treat diabetes, diarrhoea, skin problem, hypertension, menstrual problems, abdominal pain, intestinal pain, abortion, breast cancer and anthrax (personal communication with healers).

A number of researchers reported biological activities of crude extracts from different parts of Momordica species^{13,15-22} as well as isolation of pure compounds²³⁻³⁴. Moreover, the biological/pharmacological properties of the different parts of these species are attributed to the presence of phytochemicals such as flavonoids, saponins, terpenoids, coumarins, alkaloids, proteins, cardiac glycosides, anthraquinones, anthocyanins, steroids and phenols in its leaves and fruits^{7,29,30,35-42}. To the best of authors knowledge, there are no reports on phytochemical study and isolation of compounds from roots and other parts of *M. boivinii*. Therefore, the aim of the present study was to carry out phytochemical screening on root extract of *M. boivinii* so as to detect its chemical composition and perhaps substantiate the traditional use of the plant. The phytochemical screening tests were carried out following standard procedures reported in literature.

MATERIALS AND METHODS

Chemicals: The n-hexane, dichloromethane and methanol were used for gradient extraction; ethyl acetate and n-hexane were used for column elution and pre-coated Thin Layer Chromatography (TLC) (silica gel, UV254) plates were



Fig. 1(a-c): Aerial part of *M. boivinii* Photo by Filippo T., 29 September, 2016

used for chromatographic analyses. The reagents such as Dragendorff's, dilute sodium hydroxide, ferric chloride, concentrated hydrochloric acid, concentrated sulphuric acid glacial acetic acid, acetic anhydride and chloroform were used to determine phytochemicals in the crude extracts. CDCl₃ was used as solvent for spectroscopic analysis. The chemicals used in this study were all of analytical grades and were purchased from Ranchem Co. Ltd. Agents in Addis Ababa, Ethiopia.

Equipments: Rotary evaporator (Heidolph, UK) for concentration of crude extracts, Grant (GLS 400) thermostatic bath shaker (for maceration of plant materials) was used. Oven (model N50L, GENLAB, WIDNES, ENGLAND), Analytical Balance ADAM (AFP-110L), UV chamber (Uvitec), ¹H-NMR, ¹³C-NMR and DEPT-135 were recorded using Bruker 400 MHz spectrometer for characterization of the isolated compounds. Infrared (IR) spectra were obtained from Perkin Elmer BX infrared spectrometer (400-4000 cm⁻¹).

Collection of plant material: The root of *M. boivinii* was collected in August 29, 2016 from Dale district, Sidama Zone, SNNPR region, Ethiopia. The area is about 317 km far from Addis Ababa (in Southern direction on the highway to Kenya). It is also 42 km from Hawassa University in the same direction. The plant (species) was authenticated by botanist Reta Regassa, Department of Biology, Hawassa College of Teachers' Education, Ethiopia and was given voucher number of MB/0034.

Preparation of plant specimen: The collected plant material (root) was chopped to small pieces and air-dried for 30 days without exposing to sun light and was then milled to suitable size for extraction using mortar and pestle.

Extraction: The roots of *M. boivinii* (500 g) were extracted sequentially using n-hexane, dichloromethane, dichloromethane/methanol (50:50% by volume) and methanol and maceration techniques. The mixture was subjected to continuous shaking for 48 h. The solution was filtered using Whatmann No.1 filter paper and the residual solvent in each gradient extract was removed using rotary evaporator under reduced pressure. The crude extracts of each solvent was dried, weighed and stored refrigerator till further analyses (phytochemical screening and chromatographic isolation). The percent yields of the extracts were calculated using the Eq.:

Percentage yield = $\frac{\text{Mass of the extract}}{\text{Mass of the plant material used for extraction}} \times 100$

Phytochemical screening tests: Phytochemical screening tests were carried out on the crude extract of n-hexane, dichloromethane, dichloromethane/methanol (50:50% by volume) and methanol using standard procedures reported in literature⁴³⁻⁴⁶ to detect the presence of some secondary metabolites namely steroids, terpenoids, saponins, flavonoids, tannins, alkaloids, phenols and glycosides.

Isolation and characterization of compounds: Among the above four crude extracts, the crude extract of dichloromethane/methanol (50:50% by volume) showed the best TLC profile. Then, 6.5 g of dichloromethane/methanol (50:50% by volume) extract was adsorbed onto silica gel (20 g) and subjected to column chromatographic isolation. The column was then eluted with n-hexane:ethyl acetate solvent system (with gradual increase in polarity). A total of 71 fractions (each 30 mL) were collected. The collected fractions were concentrated using rotary evaporator. The spots on the TLC plates were visualized using UV light (at 254 and 365 nm) followed by iodine vapor. Fractions were tested using TLC and those with the same TLC profiles were combined. The column chromatographic separation led to isolation of two compounds (compound MB-1 and MBC-1). The structural elucidations of the compounds were carried out based on data obtained from spectroscopic (UV, IR and NMR) data. All the spectroscopic analyses were done at the Department of Chemistry, Addis Ababa University, Ethiopia.

RESULTS AND DISCUSSION

All the crude extracts of *M. boivinii* were subjected to preliminary identification of phytoconstituents. The structural elucidation of the isolated compounds was performed based on the spectroscopic data (UV-Vis, FTIR and NMR) data and comparison with literature reports.

Masses of crude extracts: As discussed above, the extraction of the plant material (root) was carried out using different solvents (of different polarities) and in sequential extraction approach. It was started with n-hexane (least polar) and followed by dichloromethane and dichloromethane/methanol (50:50% by volume) (intermediate polarity) and methanol (the most polar). The amounts of extracts were 1.1, 1.8, 7.5 and 5.4 g for n-hexane, dichloromethane, dichloromethane/ methanol (50:50% by volume) and methanol, respectively (Table 1). The resulted amount of crude extract of dichloromethane/methanol was found to be higher than the other crude extracts.

Phytochemical screening: Preliminary phytochemical screening tests were performed as per standard procedure, the various phytoconstituents on n-hexane, dichloromethane, dichloromethane/methanol (50:50% by volume) and methanol root extracts of *M. boivinii* following standard report. The results of the tests are presented in Table 2. Three secondary metabolites (flavonoids, phenols and tannins) were not detected in n-hexane extract. Tannins were also not detected in methanol extract. The other two extracts (dichloromethane extract and dichloromethane/ methanol (50:50% by volume extract) were found to contain all the secondary metabolites tested in the present study (Table 2).

Secondary metabolites are known to elicit several biological/pharmacological activities such as antiulcer, anti-inflammatory, antioxidant, cytotoxic, antitumor and antidepressant activities (e.g., phenols)⁴⁷, antimicrobial, cytotoxicity, anti-inflammatory, antitumor, oestrogenic, anti-allergic, antioxidant and vascular activities (e.g., flavonoids)47,48, astringents, against diarrhoea, as diuretics, against stomach and duodenal tumors and as anti-inflammatory, antiseptic, antioxidant and haemostatic pharmaceuticals (e.g., tannins)^{47,49}, antihypertensive effects antiarrhythmic effect, antimalarial activity and anticancer actions (e.g., alkaloids)^{47,50}, anticarcinogenic, antimalarial, anti-ulcer, hepaticidal, antimicrobial or diuretic activity and the sesquiterpenoid antimalarial drug artemisinin and anticancer (e.g., terpenoids and steroids)^{50,51}. There are also reports that show antimicrobial activities of extracts of Momoridca species. For instance, extracts obtained from different parts of Momordica species such as M. charantia have been found to show antimicrobial^{35,52-55}, antioxidant^{52,56,57} and antidiabetic^{58,59} activities. These biological/ pharmacological activities believed to be due to the above secondary metabolites^{25,60}. In the present study, the aforementioned secondary metabolites were detected in the root extracts of *M. boivinii*. This supports medicinal uses of the roots and also suggests the potentials of the different parts of the plant to be rich sources of compounds that could be

used as drug candidates. However, further biological activity test is needed on the root extracts and also on the extracts from other parts of the plant as such studies may lead to drug discovery and development.

Isolation and characterization of compounds: Two compounds (MB-1 and MBC-1) were isolated in the present study. The structures of these compounds are proposed based on the spectral data and comparison with literature reports.

Structural elucidation of compound MB-1: Compound MB-1 was obtained as a white amorphous solid (73.4 mg). Its R_f value was determined to be 0.4 in n-hexane:ethyl acetate (80:20%). Analysis of its IR spectrum (Appendix 1) revealed a broad band at 3435 cm⁻¹ indicating the presence of hydroxyl functional group. The strong band at 2924 cm⁻¹ represents C-H stretch of alkenes whereas the weak band around 2800 cm⁻¹ could be attributed to C-H stretching of methyl groups. The band at 1631 cm⁻¹ could be attributed to C=C bond. The observed data suggested that compound MB-1 could be an alcohol with at least one C=C double bond¹⁸.

In the ¹H-NMR spectrum (Appendix 2) of compound MB-1, the singlet peaks at δ 0.75, 0.81, 0.86, 0.94, 1.01 and 1.27 (Table 3) indicate the presence of protons of six methyl (-CH₃) groups whereas the peaks at δ 5.20 and 5.40 indicate the presence of olefinic protons in compound MB-1. The patterns of peaks in the ¹³C-NMR spectrum (Appendix 3) of compound MB-1 suggest that the compound could be stigmasterol-type compound. The peaks/signals at δ 140.80, 121.70, 139.60 and 117.30 could be assigned to C-5, C-6, C-22, C-23 double bonds, respectively, of stigmasterol-type compounds (Table 3). The

Table 1: Mass of extracted matter in each gradient extract

	Mass of crude	Yield
Solvent used for extraction	extract (g)	(%)
n-hexane	1.1	0.22
Dichloromethane	1.8	0.32
Dichloromethane/methanol (50:50% by volume)	7.5	1.50
Methanol	5.4	1.08

	Extracts					
Classes of						
phytochemicals	n-hexane	Dichloromethane	Dichloromethane/methanol (50:50% by volume)	Methanol		
Alkaloids	+	+	+	+		
Saponins	+	+	+	+		
Flavonoids	-	+	+	+		
Phenols	-	+	+	+		
Steroids	+	+	+	+		
Glycosides	+	+	+	+		
Terpenoids	+	+	+	+		
Tannins	-	+	+	-		

+: Present, -: Not detected



Appendix 1: IR spectrum of compound MB-1

Carbon	¹³ C-NMR data	Reported ¹³ C-NMR	¹ H-NMR data of	Reported ¹ H-NMR	Nature
atoms	of compound MB-1	data of stigmasterol	compound MB-1	data of stigmasterol	of carbon
C-1	37.2	37.2			CH ₂
C-2	31.7	31.6			CH ₂
C-3	71.8	71.7	3.6 (tdd, 1H)	3.51(tdd, 1H)	CH
C-4	42.3	42.2			CH_2
C-5	140.8	140.8			C=C
C-6	121.7	126.6	5.40 (t, 1H)	5.31(t, 1H)	C=CH
C-7	31.5	31.6			CH ₂
C-8	29.7	31.8			СН
C-9	50.2	50.0			СН
C-10	36.3	36.2			C
C-11	24.3	21.1			CH ₂
C-12	39.6	39.6			CH ₂
C-13	42.3	42.1			С
C-14	56.8	56.8			CH
C-15	24.3	24.3			CH ₂
C-16	29.0	28.8			CH ₂
C-17	56.1	55.8			СН
C-18	11.8	12.2	1.27 (d, 3H)	1.03 (s, 3H)	CH₃
C-19	21.6	19.9	0.75 (d, 3H)	0.71 (s, 3H)	CH ₃
C-20	40.3	40.4-40.5			CH
C-21	22.0	21.0	1.01 (d, 3H)	0.91 (d, 3H)	CH₃
C-22	139.6	138.2	5.20 (m, 1H)	4.98 (m, 1H)	C=C
C-23	117.4	129.2-129.6	5.20 (m, 1H)	5.14 (m, 1H)	C=C
C-24	55.1	51.1-51.3			СН
C-25	31.7	31.9			CH
C-26	21.1	21.2	0.86 (d, 3H)	0.80 (d, 3H)	CH₃
C-27	19.4	19.1	0.94 (d, 3H)	0.82 (d, 3H)	CH₃
C-28	26.5	25.4-25.5			CH ₂
C-29	12.3	12.3-25.3	0.81 (t, 3H)	0.83 (t, 3H)	CH ₃











Appendix 4: IR spectrum of compound MBC-1 in CDCl₃



Fig. 2: Proposed structural of compound MB-1 (the same with that of stigmasterol)

peak 71.8 can also be assigned to methine at C-3 bearing a hydroxyl group⁶¹. The ¹³C-NMR (Appendix 3) showed that a total of 29 and 26 signals, respectively, that could be assigned to six methyl, nine methylene, eleven methine and three quaternary carbon atoms. Thus, the observed NMR data of compound MB-1 were found to be consistent with the reported NMR data of stigmasterol (or 24-Ethyl-cholesta-5, 22-dien-3beta-ol) (Fig. 2, Table 3)⁶²⁻⁶⁴.

Structural elucidation of MBC-1: A greenish crystalline compound (37.5 mg, labeled as MBC-1) was obtained from the column chromatographic separation that was eluted with hexane:ethyl acetate (94:6%) solvent system. Its R_{f} value was

determined to be 0.45 in n-hexane:ethyl acetate (90:10%) solvent system. Its IR spectrum (Appendix 4) showed a strong band around 3429 cm⁻¹ indicating the compound has an alcohol or alcohol -OH functional group. The strong band at 2926 cm⁻¹ represents C-H stretch of methylene whereas, the weak band (shoulder-type) around 2800 cm⁻¹ indicated the C-H stretching of methyl groups. The band at 1679 cm⁻¹ indicate the presence of a,b-unsaturated carbonyl functional group and bands around 1600 and 1500 cm⁻¹ (aromatic and olefin C=C bonds). The observed data suggested that compound MBC-1 could bear an alcohol/phenol functional group and also at least one C=C double bond and aromatic ring.

¹H-NMR spectrum (Appendix 5) showed symmetric doublet of doublet peaks at δ 6.4 and 7.6 indicating the presence of trans-olefinic hydrogen that is conjugated with aromatic ring. Moreover, peaks at 7.1, 7.0 and 6.9 could be attributed to three protons of tri-substituted aromatic/benzene ring. On the other hand, a singlet (and intense) peaks at δ 3.9 indicate the presence of methoxy group whereas a singlet peak at δ 4.8 indicates the presence of alcohol or phenol hydroxyl (-OH) group of an alcohol. The broad singlet peaks at δ 5.3 and 5.5 indicate the presence of germinal olefinic hydrogen atoms. The ¹H-NMR spectrum also revealed a broad range of signals (from δ 0.7-2.5) that could be attributed to methine, methylene and methyl protons





Carbon	¹³ C-NMR data of	Reported ¹³ C-NMR data of	¹ H-NMR of compound	Reported ¹ H-NMR data of	Nature of
atoms	compound MBC-1 (ppm)	Betulinic acid-3-trans-caffeate ⁶⁵⁻⁶⁷	MBC-1 (ppm)	Betulinic acid-3-trans-caffeate ⁶⁵⁻⁶⁷	carbon
C-1	38.6	39.2	1.70 (m)	1.73(m)	CH ₂
C-2	23.9	24.6	1.66 (m)	1.68 (m)	CH ₂
C-3	78.4	81.2	4.2 (t)	4.57-5.1(t)	CH
C-4	37.5	38.3	-	-	С
C-5	56.3	55.7	0.90 (m)	0.91(m)	CH
C-6	19.0	19.0	1.52 (m)	1.53, 146 (m)	CH_2
C-7	33.0	35.1	1.4 (m)	1.39 (m)	CH_2
C-8	40.2	42.7	-	-	С
C-9	43.3	50.7	1.4 (m)	1.43 (m)	СН
C-10	31.9	34.6			С
C-11	22.1	21.8	1.13 (m)	1.12 (m)	CH ₂
C-12	22.1	26.4	1.26 (m)	1.74 (m)	CH ₂
C-13	36.9	39.0	2.38 (m)	239 (m)	CH
C-14	40.5	43.3			С
C-15	30.2	30.5	1.6 (m)	1.59 (m)	CH ₂
C-16	31.9	32.9	2.26 (m)	2.26 (m)	CH ₂
C-17	60.5	56.8	-	-	С
C-18	45.2	49.5	1.6 (m)	1.59 (m)	CH
C-19	43.3	47.1	3.5 (m)	3.0(m)	CH
C-20	147.8	150.0	-	-	С
C-21	32.6	31.4	-	1.93, 1.37(m)	CH ₂
C-22	37.0	37.9	1.92 (m)	1.93, 1.37 (m)	CH ₂
C-23	28.9	28.3	0.85 (s)	0.86 (s)	CH ₃
C-24	19.0	16.3	0.87 (s)	0.89(s)	CH ₃
C-25	20.3	16.9	0.90 (s)	0.93 (s)	CH ₃
C-26	16.30	16.4	0.96 (s)	0.97 (s)	CH ₃
C-27	14.1	14.9	0.96 (s)	0.97 (s)	CH_3
C-28	183.7	177.8	-	-	С
C-29	110.4	110.0	-	4.60, 4.57	C
C-30	20.4	19.6	1.6 (s)	1.68 (s)	CH₃
C-31	56.1	-	3.9 (s)	-	OCH ₃
1′	127.2	128.0	-	-	CH
2′	122.7	122.6	7.0 (s)	7.07 (d)	C
3′	115.9	115.8	-	-	С
4′	146.7	146.1	-	-	CH
5′	144.3	143.9	6.9 (d)	6.86 (d)	CH
6′	114.8	114.6	7.1 (d)	6.99 (dd)	С
7′	144.9	144.3	7.6 (d)	7.53 (d)	CH
8′	116.2	116.9	6.3 (d)	6.24 (d)	CH
9′	167.3	167.7			С

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Table 4: The ¹³C-NMR, DEPT-135 and ¹H-NMR data of compound MBC-1 and reported NMR data of betulinic acid-3-trans-caffeate⁶⁵⁻⁶⁷

of triterpene skeleton. ¹³C-NMR spectrum (Appendix 6) of compound MBC-1 suggested that the compound could be a triterpene with 31 carbons. Further investigation of ¹³C-NMR spectrum also revealed the characteristic signals for no-conjugated ester (δ 183.7), vinyl carbons (quaternary carbon at δ 147.8 and CH₂ at δ 110.4) as well as an oxygen-bearing methine group (δ 78.4) (Table 4). Additional aromatic moiety peaks were observed at δ 127.2, 122.7, 115.9, 146.7, 144.3 and 114.8. Comparison of the NMR spectral data of compound MBC-1 with data reported in literature⁶⁵⁻⁶⁷ suggested that the compound has similar skeleton with that of betulinic acid-3-trans-caffeate. The only difference between compound MBC-1 and betulinic acid-3-trans-caffeate was that in the spectrum of compound MBC-1, there is no band of carboxylic acid functional group (in the range of 2400-3400 cm⁻¹). Moreover, there was no peak around 12 ppm in the ¹H-NMR spectrum of compound MBC-1. This could indicate that, though it has similar skeleton with betulinic acid-3-trans-caffeate, compound MBC-1 has no carboxylic acid functional group or it is not betulinic acid-3-trans-caffeate. Instead, both ¹H-NMR and ¹³C-NMR showed the presence of methoxy group bonded to carbonyl functional group or presence of COOCH₃. Based on the spectral data and literature reports, compound MBC-1 is suggested to be methyl ester of betulinic acid-3-trans-caffeate (Fig. 3).





Fig. 3(a-b): Chemical structures of (a) Betulinic acid-3-transcaffeate and (b) Proposed structure of compound MBC-1

CONCLUSION

Phytochemical screening tests revealed that *M. boivinii* roots contain alkaloids, saponins, flavonoids, phenols, steroids, glycosides, terpenoids and tannins. However, further investigations are suggested to evaluate biological activities of root extracts and also extracts from its leaves and fruits. Moreover, two compounds (Stigmasterol and methylester of betulinic acid-3-trans caffeate) were identified and their structures were determined based on spectroscopic data and comparison of the data with literature reports. The results suggested the potential of the plant in discovery of new drugs for treatment of human diseases.

SIGNIFICANCE STATEMENT

The study conducted to analyze the phytoconstituents of *M. boivinii* roots and found that it contain some active compounds like alkaloids, saponins, flavonoids, phenols, steroids, glycosides, terpenoids and tannins which confirm the secondary metabolites presence within the plant root. This study would help other researchers in ameliorating the secondary metabolites in detail in future with their medical perspective. Thus, the best theory on it may be arrived at.

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REFERENCES

- Joseph, J.K. and V.T. Antony, 2008. Ethnobotanical investigations in the genus *Momordica* L. in the Southern Western Ghats of India. Genet. Resour. Crop Evol., 55: 713-721.
- 2. Ojewole, J.A., S.O. Adewole and G. Olayiwola, 2005. Hypoglycaemic and hypotensive effects of *Momordica charantia* Linn (Cucurbitaceae) whole-plant aqueous extract in rats. Cardiovasc. J. South Afr., 17: 227-232.
- Ganesan, A., S. Natesan, R. Vellayutham, K. Manickam and N. Ramasamy, 2008. Anxiolytic, antidepressant and antiinflammatory activity of methanol extract of leaves of *Momordica charantia*Linn (Cucurbitaceae). Iran. J. Pharmacol. Therapeut., 7: 43-47.
- Bakare, R.I., O.A. Magbagbeola, O.W. Okunowo and M. Green, 2011. Antidiarrhoeal activity of aqueous leaf extract of *Momordica charantia* in rats. J. Pharmacogn. Phytother., 3: 1-7.
- 5. Nadkarni, K.M., 2007. Indian Materia Medica. Vol. 2, Popular Prakashan, Mumbai, India, Pages: 296.
- Satyavati, G.V., M.K. Raina and M. Sharma, 1976. Medicinal Plants of India. Vol. 1, Indian Council of Medical Research, New Delhi, India.
- Singh, J., E. Cumming, G. Manoharan, H. Kalasz and E. Adeghate, 2011. Suppl 2: Medicinal chemistry of the anti-diabetic effects of *Momordica charantia*: Active constituents and modes of actions. Open Med. Chem. J., 5: 70-77.
- 8. Jeffrey, C., 1978. *Momordica boivinii* Baill. [Family: Cucurbitaceae]. Royal Botanic Gardens, Kew, UK.
- Pakia, M. and J.A. Cooke, 2003. The ethnobotany of the Midzichenda tribes of the Coastal forest areas in Kenya. 2. Medicinal plant uses. S. Afr. J. Bot., 69: 382-395.
- Choi, C.W., S.B. Song, J.S. Oh and Y.H. Kim, 2015. Antiproliferation effects of selected Tanzania plants. Afr. J. Trad. Complement. Alternat. Med., 12: 96-102.
- Morris, B., 1996. Chewa Medical Botany: A Study of Herbalism in Southern Malawi. Vol. 2, LIT Verlag, Munster, Germany, pp: 82-83.
- 12. Anonymous, 2018. Medicinal plant compositions of matter and method of preparation-patents. US Patent No. 7, 306, 816. March 17, 2018, USA.
- 13. Eshetu, G.R., T.A. Dejene, L.B. Telila and D.F. Bekele, 2015. Ethnoveterinary medicinal plants: Preparation and application methods by traditional healers in selected districts of Southern Ethiopia. Vet. World, 8: 674-684.

- 14. Sintayehu, T.B., 2011. An ethnobotanical study of medicinal plants in Wondo genet natural forest and adjacent Kebeles, Sidama zone, SNNP region, Ethiopia. M.Sc. Thesis, Department of Biology, Addis Ababa University, Ethiopia.
- Rao, B.K., M.M. Kesavulu, R. Giri and C.A. Rao, 1999. Antidiabetic and hypolipidemic effects of *Momordica cymbalaria* Hook. fruit powder in alloxan-diabetic rats. J. Ethropharmacol., 67: 103-109.
- 16. Rao, B.K., M.M. Kesavulu and C. Apparao, 2001. Antihyperglycemic activity of *Momordica cymbalaria* in alloxan diabetic rats. J. Ethnopharmacol., 78: 67-71.
- 17. Kameswararao, B., M.M. Kesavulu and C. Apparao, 2003. Evaluation of antidiabetic effect of *Momordica cymbalaria* fruit in alloxan-diabetic rats. Fitoterapia, 74: 7-13.
- Faparusi, F., M.M. Bello-Akinosho, R.T. Oyede, A. Adewole, P.O. Bankole and F.F. Ali, 2012. Phytochemical screening and antibacterial activity of *Brillantaisia patula* leaf. Res. J. Phytochem., 6: 9-16.
- 19. Kirtikar, K.R. and B.D. Basu, 1975. Indian Medicinal Plants. Curcurbitaceae Dehradun. Vol. 3, Bishen Singh Mahindra Pal Singh, New Delhi, India, Pages: 1137.
- Koneri, R., R. Balaraman and C.D. Saraswati, 2006. Antiovulatory and abortifacient potential of the ethanolic extract of roots of *Momordica cymbalaria* Fenzl in rats. Indian J. Pharmacol., 38: 111-114.
- Sajjan, S., S.H. Chetana, P.M. Paarakh and A.B. Vedamurthy, 2010. Antimicrobial activity of *Momordica cymbalaria* Fenzl aerial parts extracts. Indian J. Nat. Prod. Res., 1: 296-300.
- Jeevanantham, P., S. Vincent, A. Balasubramaniam, B. Jayalakshmi and N.S. Kumar, 2011. Anti cancer activity of methanolic extract of aerial parts of *Momordica cymbalaria* Hook F. against *Ehrlich ascites* carcinoma in mice. J. Pharm. Sci., 3: 1408-1411.
- 23. Day, C., T. Cartwright, J. Provost and C.J. Bailey, 1990. Hypoglycaemic effect of *Momordica charantia* extracts. Planta Medica, 56: 426-429.
- 24. Jiang, Y., X.R. Peng, M.Y. Yu, L.S. Wan and G.L. Zhu *et al.*, 2016. Cucurbitane-type triterpenoids from the aerial parts of *Momordica charantia* L. Phytochem. Lett., 16: 164-168.
- Oragwa, L.N., O.O. Olajide, O.O. Efiom and S.K. Okwute, 2013. Didecanoate compound: Isolated from *Momordica charantia*Linn. seeds from Nigeria. Afr. J. Pure Applied Chem., 7: 375-381.
- Pornngarm, L., P. Pornsiri, S. Shugo and J. Kuguacin, 2013. A triterpenoid from *Momordica charantia* Linn: A comprehensive review of anticarcinogenic properties. InTech Open Sci., 13: 273-279.
- Takasaki, M., T. Konoshima, Y. Murata, M. Sugiura and H. Nishino *et al.*, 2003. Anticarcinogenic activity of natural sweeteners, cucurbitane glycosides, from *Momordica grosvenori*. Cancer Lett., 198: 37-42.

- 28. Thiruvengadam, M. and I.M. Chung, 2011. Establishment of an efficient *Agrobacterium tumefaciens*-mediated leaf disc transformation of spine gourd (*Momordica dioica* Roxb. ex Willd). Afr. J. Biotechnol., 10: 19337-19345.
- 29. Talukdar, S.N. and M.N. Hossain, 2014. Phytochemical, phytotherapeutical and pharmacological study of *Momordica dioica*. Evidence-Based Complement. Altern. Med. Vol. 2014. 10.1155/2014/806082.
- Jain, A., A. Nahata, S.R. Lodhi and A.K. Singhai, 2014. Effects of *Tephrosia purpurea* and *Momordica dioica* on streptozotocininduced diabetic nephropathy in rats. Biomed. Prevent. Nutr., 4: 383-389.
- Taylor, L., 2002. Bitter Melon (*Momordica Charantia*), Herbal Secrets of the Rainforest. 2nd Edn., Sage Press Inc., Austin Texas, USA., pp: 1-100.
- 32. Lin, K.W., S.C. Yang and C.N. Lin, 2011. Antioxidant constituents from the stems and fruits of *Momordica charantia*. Food Chem., 127: 609-614.
- Jung, K., D. Lee, J.S. Yu, H. Namgung, K.S. Kang and K.H. Kim, 2016. Protective effect and mechanism of action of saponins isolated from the seeds of gac (*Momordica cochinchinensis* Spreng.) against cisplatin-induced damage in LLC-PK1 kidney cells. Bioorg. Med. Chem. Lett., 26: 1466-1470.
- Ramalhete, C., D. Lopes, S. Mulhovo, J. Molnar, V.E. Rosario and M.J.U. Ferreira, 2010. New antimalarials with a triterpenic scaffold from *Momordica balsamina*. Bioorg. Med. Chem., 18: 5254-5260.
- 35. Annapoorani, C.A. and K. Manimegalai, 2013. Screening of medicinal plant *Momordica charantia* leaf for secondary metabolites. Int. J. Pharm. Res. Dev., 5: 1-6.
- Bharathi, L.K., H.S. Singh, S. Shivashankar and A.N. Ganeshamurthy, 2014. Characterization of a fertile backcross progeny derived from inter-specific hybrid of *Momordica dioica* and *M. subangulata* subsp. renigera and its implications on improvement of dioecious *Momordica* spp. Sci. Horticult., 172: 143-148.
- Kritikar, K.R. and B.D. Basu, 1999. Indian Medicinal Plants. Vol. 2, International Book Distributors, Dehradun, India, pp: 1129-1135.
- Thiruvengadam, M., K.T. Rekha, E.H. Kim, N. Praveen and I.M. Chung, 2013. Effect of exogenous polyamines enhances somatic embryogenesis via suspension cultures of spine gourd (*Momordica dioica* Roxb. ex. Willd.). Aust. J. Crop Sci., 7: 446-453.
- Sarma, D.S., A.V.S. Babu, K.R. Krishna and P.N. Basha, 2011. Phytochemical studies and biological activities on fruits of *Momordica cochinchinensis*. J. Chem. Pharm. Res., 3:875-881.
- 40. Behera, T.K., J.K. John and L.K. Bharathi, 2011. *Momordica*. In: Wild Crop Relatives: Genomic and Breeding Resources, Vegetables, Kole, C. (Ed.)., Springer-Verlag, Berlin, Heidelberg, pp: 217-246.

- 41. Anilakumar, K.R., G.P. Kumar and N. Ilaiyaraja, 2015. Nutritional, pharmacological and medicinal properties of *Momordica charantia*. Int. J. Nutr. Food Sci., 4: 73-83.
- Ahmed, I., M.S. Lakshni, M. Gillet, A. John and H. Raza, 2001. Hypotriglyceridemic and hypocholesterolemic effects of antidiabetic *Momordica charantia* (karela) fruit extract in streptozotocin-induced diabetic rats. Diabetes Res. Clin. Pract., 51: 155-161.
- 43. Tiwari, P., B. Kumar, M. Kaur, G. Kaur and H. Kaur, 2011. Phytochemical screening and extraction: A review. Internationale Pharmaceutica Sciencia, 1: 98-106.
- 44. Ganesh, S. and J.J. Vennila, 2011. Phytochemical analysis of *Acanthus ilicifolius* and *Avicennia officinalis* by GC-MS. Res. J. Phytochem., 5: 60-65.
- Okamoto, Y., A. Suzuki, K. Ueda, C. Ito and M. Itoigawa *et al.*, 2006. Anti-estrogenic activity of prenylated isoflavones from *Millettia pachycarpa*: Implications for pharmacophores and unique mechanisms. J. Health Sci., 52: 186-191.
- 46. Pooja, S. and G.M. Vidyasagar, 2016. Phytochemical screening for secondary metabolites of *Opuntia dillenii* Haw. J. Med. Plants, 4: 39-43.
- Saxena, M., J. Saxena, R. Nema, D. Singh and A. Gupta, 2013. Phytochemistry of medicinal plants. J. Pharmacogn. Phytochem., 1: 168-182.
- 48. Hodek, P., P. Trefil and A. Stiborova, 2002. Flavonoids potent and versatile biologically active compounds interacting with cytochrome P 450. Chem.-Biol. Int., 139: 1-21.
- 49. Dharmananda, S., 2003. Golinuts and the uses of tannins in Chinese Medicine. Proceedings of the Institute for Traditional Medicine, September 25, 1993, Portland, USA.
- 50. Nobori, T., K. Miura, D.J. Wu, A. Lois, K. Takabayashi and D.A. Carson, 1994. Deletions of the cyclin-dependent kinase-4 inhibitor gene in multiple human cancers. Nature, 368: 753-756.
- Just, M.J., M.C. Recio, R.M. Giner, M.J. Cuellar, S. Manez, A.R. Bilia and J.L. Rios, 1998. Anti-inflammatory activity of unusual lupane saponins from *Bupleurum fruticescens*. Planta Med., 64: 404-407.
- 52. Leelaprakash, G., J.C. Rose, B.M. Gowtham, P.K. Javvaji and S.A. Prasad, 2011. *In vitro* antimicrobial and antioxidant activity of *Momordica charantia* leaves. Pharmacophore, 2: 244-252.
- Majumdar, T., B. Deb, A. Das, A. Chakraborty and B.B. Goswami, 2011. Isolation of antimicrobially active compounds from the leaf of tribal edible plants of Tripura: *Momordica charantia* L. and *Paederia foetida* L. J. Nat. Prod. Plant Resour., 1: 108-116.
- 54. Mada, S.B., A. Garba, H.A. Mohammed, A. Muhammad, A.Olagunju and A.B. Muhammad, 2013. Antimicrobial activity and phytochemical screening of aqueous and ethanol extracts of *Momordica charantia* L. leaves. J. Med. Plants Res., 7: 579-586.

- 55. Abhinay, T., D. Vaibhav., G. Sayali and K. Sayali, 2014. Extraction of phytochemical components from the fruit of *Momordica charantia* and evaluation of its antimicrobial activity. Proceedings of the IRF International Conference, March 30, 2014, Pune, India.
- 56. Tan, S.P., C. Stathopoulos, S. Parks and P. Roach, 2014. An optimised aqueous extract of phenolic compounds from bitter melon with high antioxidant capacity. Antioxidants, 3:814-829.
- 57. Immaculate, A.C., L. Nivethini and V.T. Diana, 2015. Phytochemical screening and bioactivity of *Momordica charantia* L. Chem. Pharm. Res., 7: 970-975.
- Harinantenaina, L., M. Tanaka, S. Takaoka, M. Oda, O. Mogami, M. Uchida and Y. Asakawa, 2006. *Momordica charantia* constituents and antidiabetic screening of the isolated major compounds. Chem. Pharm. Bull., 54: 1017-1021.
- 59. Tongia, A., S.K. Tongia and M. Dave, 2004. Phytochemical determination and extraction of *Momordica charantia* fruit and its hypoglycemic potentiation of oral hypoglycemic drugs in diabetes mellitus (NIDDM). Indian J. Physiol. Pharmacol., 48: 241-244.
- 60. Daniel, P., U. Supe and M.G. Roymon, 2014. A review on phytochemical analysis of *Momordica charantia*. Int. J. Adv. Pharm. Biol. Chem., 3: 214-220.
- 61. Li, H., Z. Wang and Y. Liu, 2003. [Review in the studies on tannins activity of cancer prevention and anticancer]. Zhong Yao Cai, 26: 444-448, (In Chinese).
- 62. Pollock, J.R.A. and R.S. Stevem, 1965. Dictionary of Organic Compounds. 4th Edn., Eyre and Spottiswoode Ltd., UK.
- Kamboj, A. and A.K. Saluja, 2011. Isolation of stigmasterol and β-sitosterol from petroleum ether extract of aerial parts of *Ageratum conyzoides* (Asteraceae). Int. J. Pharm. Pharm. Sci., 3: 94-96.
- 64. Pateh, U.U., A.K. Haruna, M. Garba, I. Iliya, I.M. Sule, M.S. Abubakar and A.A. Ambi, 2009. Isolation of stigmasterol, β-sitosterol and 2-hydroxyhexadecanoic acid methyl ester from the rhizomes of *Stylochiton lancifolius* Pyer and Kotchy (Araceae). Niger. J. Pharm. Sci., 8: 19-25.
- Kim, K.H., S.U. Choi and K.R. Lee, 2010. Bioactivity-guided isolation of cytotoxic triterpenoids from the trunk of *Berberis koreana*. Bioorganic Med. Chem. Lett., 20: 1944-1947.
- Kim, J.H., J.C. Byun, A.K.R. Bandi, C.G. Hyun and N.H. Lee, 2009. Compounds with elastase inhibition and free radical scavenging activities from *Callistemon lanceolatus*. J. Med. Plant. Res., 3: 914-920.
- 67. Pan, H., L.N. Lundgren and R. Andersson, 1994. Triterpene caffeates from bark of *Betula pubescens*. Phytochemistry, 37: 795-799.