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Effect of Different Durations of *Schistosoma mansoni* Infection on the Levels of Some Antioxidants in Mice

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Abstract: The levels of lipid peroxides, glutathione, vitamin C, vitamin E catalase and superoxide dismutase enzymes were measured in the livers of Schistosoma mansoni infected mice (CD strain weighing 20-25 g) at different durations post infection (1, 2, 4, 6 and 8 weeks). Moreover, the liver weight, body weight, liver to body weight and total protein were studied in the same animal groups. Control non-infected groups were run simultaneously with each infected mice group. The data obtained showed that lipid peroxides were elevated throughout the different durations of infection while glutathione decreased with infection. On the other hand, both vitamins C and E showed a reduction in the livers of mice during the different durations of infection. The activity of catalase showed an insignificant change after one and two weeks and a high significant decrease in the livers of four, six and eight weeks S. mansoni infected mice, while, superoxide dismutase significantly decreased one and two weeks post infection with a significant elevation four, six and eight weeks post infection. Moreover, a significant reduction was observed in body weight after four, six and eight weeks of infection accompanied with an elevation in liver weight only after six and eight weeks. Consequently, liver weight/body weight showed an elevation after four, six and eight weeks of infection. Finally, the protein content was significantly lower at one, six and eight weeks post infection with S. mansoni. It could thus be concluded that host-parasite association results in production of free radicals as a result of an oxidative stress where the parasites struggle to overcome the immune response of the host and changes in host liver antioxidants occur as a means to scavenge these radicals.

Key words: Antioxidants, schistosomiasis, free radicals

Introduction

Schistosomiasis ranks next to malaria regarding all the endemic parasitic diseases worldwide particularly in Egypt (El-Baz *et al.*, 2003). In parallel with massive research efforts in schistosomiasis over the past 30 years, persistent efforts have been made to understand the basis for infection (Van Nassauw *et al.*, 2001). The tropical parasite *Schistosoma mansoni* causes granulomatous inflammation after its eggs lodge in the hepatic portal capillaries. Previous studies indicate that the host's response involves the production of reactive oxygen species (Connors *et al.*, 1995; El-Sokkary *et al.*, 2002). Sayed and Williams (2004) reported that schistosomes use alternative electron donors and their variable resistance to overoxidation may reflect their presence in different cellular sites and emphasizes the significant differences in overall redox balance mechanisms between the parasite and the mammalian host.

The consequences of free radical generation in *S. mansoni* is still unknown (Facundo *et al.*, 2004). In some reports concerning *Schistosoma mansoni*, Sheweita *et al.* (2003) mentioned that high levels

of free radicals provide new evidence for organ damage since glutathione is decreased and lipid peroxides increased in different organs of hamsters infected with *S. hematobium*. In this manner. *S. mansoni* infection alters the hepatic levels of glutathione and activities of glutathione metabolizing enzymes and alterations may affect the capacity of the liver to detoxify or neutralize the toxic effect of endogenous and exogenous compounds.

From the previous information, it was of interest in the present study to investigate the levels of glutathione (GSH) vitamin E (VE), vitamin C (VC), superoxide dismutase (SOD), catalase and lipid peroxides, in infected mice groups at different stages of infection with *Schistosoma mansoni* in order to trace the changes that occur in these substances throughout the infection period. The objective of the study is to investigate the situation under the disease condition throughout the stages of parasite development, whether shortly after invasion or after egg deposition, since, substances expressed during the life cycle of the parasite such as free radicals may provide useful approaches as stage specific markers for development of drugs that may counteract infection attack. This may be achieved in the future by searching for a nutrient that can protect mitochondrial enzymes and/or stimulate enzymatic activity by elevating levels of substrates and cofactors; that can enhance antioxidant defences; that can scavenge free radicals and prevent oxidant production in mitochondria and finally that can repair mitochondrial membrane.

Materials and Methods

All chemicals used were of high analytical grade, products of Sigma (USA), Merck and Reidel (Germany), BDH (England) and Fluka (Switzerland) Chemical Co. Praziquantel used is a product of Egyptian International Pharmaceutical Industries Company (E.I.P.I.Co.).

The animals used were healthy male Swiss albino mice of CD strain, obtained from Theodor Bilharz Institute, ranging in weight from 20-25 g. They received fresh, deionized water and standard diet (El-Tkamol-Co.) *ad libitum*. Mice were divided into two main groups each divided into five subgroups.

The first main infected group was sacrificed one, two, four, six and eight weeks post infection with 100 cercariae of Egyptian *Schistosoma mansoni* strain.

The second main group served as control and was run simultaneously with each duration of the infected subgroups. The number of animals ranged from 6-10 mouse in each subdivision.

To obtain cercariae, 10-20 shedding *Biomphalaria alexandrina* snails were used to ensure bisexual infection and placed in a beaker containing 200 mL dechlorinated water. Then, the snails were exposed to sunlight at 8-9 am. Each mouse was infected subcutaneously by 100 cercariae.

Experimental Procedures

The liver was removed from the mouse, plotted with filter paper and weighted. The liver was homogenized in bidistilled water (5, 10 and 20% w/v) according to the parameter measured, using potter-Elvehjem homogenizer with Teflon pestle.

Estimation of Protein

Protein was estimated by the method of Bradford (1976). Bradford solution was added to of 5% homogenate and the developed blue colour was measured after 5 min at 595 nm in spectrometer (Novaspec, LKB, Sweden) against a blank containing water instead of the homogenate. The amount of protein was calculated from a standard curve using serial concentration of bovine sernm albumin (1-10 ug).

$Estimation\ of\ Lipid\ Peroxides$

Lipid peroxides were estimated by thiobarbituric acid reaction as described by Ohkawa *et al.* (1979) using saturated thiobarbituric acid (A) and trichloroacetic acid (20%) (B) solutions. The working

solution was prepared by mixing one volume of solution A with 3 volumes of solution B. 20% tissue homogenate was added to the working solution and mixed. The tubes were boiled, cooled and centrifuged at 3.000 rpm for 10 min. The developed colour was read against the blank at 532 nm in spectrometer and calculated as μ mole malonaldehyde per gram tissue.

Estimation of Glutathione

Glutathione was estimated by the method of Moron *et al.* (1979) using sodium phosphate buffer and dithiobisnitrobenzoic acid (DTNB). The developed colour was read against blank at 412 um within 5 min in spectrometer. The amount of glutathione was calculated as mg glutathione per gram tissue used from a standard curve plotted for serial concentrations of glutathione (5-100 µg).

Estimation of Vitamin C

Vitamin C was estimated by the method of Jagota and Dani (1982) using Folin-ciocalteu reagent (2.0 M concentration diluted 3-fold with double-distilled water) and TCA. The developed blue colour was read at 760 um after 10 min in spectrometer against a blank. The amount of ascorbic acid was calculated from a standard curve of vitamin C using 5-70 µg serial concentrations of the vitamin.

Estimation of Vitamin E

Vitamin E was estimated by the method of Baker and Frank (1968) using 7.7 m M α , α -Dipyridyl and 7.4 mM ferric chloride solution. The absorbance of colour was read for the test and standard against the blank at 460 nm.

Estimation of Superoxide Dismutase Activity

The activity of superoxide dismutase was estimated by the method of Nishikimi *et al.* (1972) using nitroblue tetrazolium, NADH and sodium pyrophosphate buffer. Increase in absorbance with time during the reaction was read in spectrometer at 560 nm.

Estimation of Catalase Enzyme

Catalase activity was assayed according to Lubinsky and Bewley (1979). The reaction for assaying catalase activity was initiated by adding 20 μ L 5% liver homogenate of concentration to 2.53 mL of the reaction mixture (sodium potassium phosphate buffer containing H_2O_2 . The disappearance of hydrogen peroxide was monitored by following the decease in absorbance at 230 um by spectrometer using molar extinction coefficient for hydrogen peroxide of 62.4 (Nelson and Kiesow, 1972).

Statistical Analyses

The analyses included the calculation of the mean value, standard deviation, standard error and t-values at level $p \le 0.05$ according to the method of Ronald *et al.* (1983). These determinations were calculated for both control and treated animals.

Results

From the result of Table 1 it is obvious that infected mice showed a highly significant increase in the levels of lipid peroxides with percentages 31.29, 40.0, 89.0 and 66.78% after one, two, six and eight weeks of infection, respectively, while a significant elevation was detected in the lipid peroxides after four weeks of infection with percentage change 8.69%. On the oher hand *Schistosoma mansoni* infection showed a highly significant decrease in liver glutathione of mice, with percentages 44.0, 19.88, 24.60, 32.46 and 40.0% after one, two, four, six and eight weeks of infection, respectively.

Table 1: Estimation of lipid peroxides and glutathione in the liver of mice during different durations of Schistosoma mansoni infection

	Lipid peroxides	(µmol malonaldeh	yde/g tissue)	Glutathione mg/g tissue		
Durations post infection	Control X ±SE	Infected X ±SE	p<	Control $\overline{X} \pm SE$	Infected \overline{X} \pm SE	p<
One-week	22.85±0.10	30.0±0.133	0.001	1.925±0.045	1.075±0.05	0.001
Two-weeks	20.0±0.07	28.0 ± 0.07	0.001	2.108±0.02	1.688 ± 0.017	0.001
Four-weeks	23.0 ± 0.10	25.0±0.09	0.01	1.975±0.038	1.488 ± 0.02	0.001
Six-weeks	20.6±0.066	39.0 ± 0.07	0.001	1.625±0.037	1.097 ± 0.02	0.001
Eight-weeks	28.3±0.06	47.2±0.095	0.001	1.662 ± 0.03	0.997±0.025	0.001

Data are means ($\pm SE$) of lipid peroxides and glutathione determinations. Significant change as compared to control (p<0.001 : highly significant and p<0.01: significant)

Table 2: Determination of vitamin C and vitamin E in the liver of mice during different durations of Schistosoma mansoni infection

	Vitamin C μg/g tissue			Vitamin E mg/g tissue		
Durations post infection	Control X ±SE	Infected $\overline{\mathbf{x}} \pm \mathbf{SE}$	p<	Control X ±SE	Infected \(\overline{x} \pm SE \)	p<
One-week	269.45±7.0	182.29±6.58	0.001	10.50±1.36	4.08±0.50	0.001
Two-weeks	269.6±3.0	218.66±3.97	0.001	12.95±1.60	9.0±0.80	0.01
Four-weeks	253.75±7.50	193.4±6.0	0.001	6.04±0.56	3.68 ± 0.22	0.001
Six-weeks	234.8±5.0	164.95±3.26	0.001	3218 ± 0.30	2.395±0.16	0.01
Eight- weeks	186.35±2.0	173.6±4.28	0.001	2.966±0.44	1.87±0.26	0.01

Data are means (\pm SE) of vitamin C and vitamin E determinations. Significant change as compared to control (p<0.001: highly significant and p<0.01: significant)

It is obvious that liver vitamin C of infected mice showed a highly significant decrease after one, two, four, six and eight weeks post infection, with percentages 32.34, 18.90, 23.78, 29.70 and 6.80%, respectively Table 2. Also, vitamin E showed a highly significant decrease after one and four weeks of infection with percentages 61.0 and 39.0%, respectively and a significant reduction after two, six and eight weeks of infection with percentages 30.50, 25.57 and 36.95%, respectively as compared with clean control mice.

Table 3 shows that, an insignificant decrease of the catalase activity was detected after one and two weeks of infection, while a highly significant decrease of the liver enzyme activity was detected after four, six and eight weeks of the infection with percentages 18.42, 19.93 and 25.0%, respectively as compared with normal control mice. On the other hand superoxide dismutase enzyme showed a significant decrease in the liver of infected mice with percentages 10.67 and 5.57%, respectively after one and two weeks of the infection with *S. mansoni*. Also, the enzyme activity showed a highly significant increase after four, six and eight weeks of the infection giving percentages 11.85, 13.70 and 17.13%, respectively as compared with normal control mice.

It is obvious (Table 4) that an insignificant change is observed in the body weight after one and two weeks of infection, while a significant decrease was recorded with percentage 7.67, 4.66 and 15.80% after four six and eight weeks of infection with *S. mansoni*, respectively. Liver weight showed an insignificant change after one and four weeks of infection and a significant increase after two weeks of infection with percentage 4.0% while, a highly significant increase is recorded in liver weight after six and eight weeks of infection with percentages 25.60 and 22.0%, respectively. Liver weight/ body weight ratio showed an insignificant change after one and two weeks, while a slightly significant increase with percentage 4.90% was shown after four weeks of infection and a highly significant increase after six and eight weeks of infection with percentage 32.0 and 38.39% as compared with normal control.

Table 5 shows the level of total proteins in control and infected mice during different periods of infection with *S. mansoni*. It is observed that the level of total protein showed a significant decrease after one week of infection with percentage 4.72% and an insignificant change after two weeks of

Table 3: Estimation of catalase and superoxide dismutase enzyme activities in the liver of mice during different durations of Schistosoma mansoni infection

	Catalase µmol/n	ng proteins		Superoxide dismutase activity/mg protein		
Durations post infection	Control $\overline{\mathbf{x}} \pm \mathbf{SE}$	Infected \(\overline{x} \pm \)	p<	Control X ±SE	Infected $\overline{\mathbf{x}} \pm \mathbf{SE}$	p<
One-week	90.04±1.79	87.50±4.60	ns	331.896±6.60	296.466±6.10	0.001
Two-weeks	103.349±2.55	98.39±4.58	ns	338.14±6.77	319.305±6.50	0.01
Four-weeks	99.97±3.40	81.554±3.777	0.001	211.666±4.60	236.747±4.75	0.001
Six-weeks	89.195±3.58	71.418±3.15	0.001	178.59±5.0	203.08±5.20	0.001
Eight-weeks	85.535±4.59	64.077±2.60	0.001	156.269±5.70	183.04±6.94	0.001

Data are means (\pm SE) of catalase and superoxide dismutase determinations. Significant change as compared to control (p<0.001: highly significant, p<0.01: significant and ns: insignificant). Data are means (\pm SE) of catalase and superoxide dismutase determinations

Table 4: Body weight, liver weight and liver weight/body weight ratio of control and infected mice during different durations

	Control			Infected					
Durations									
post Infection	Body wt.	Liver wt.	L.t./B.T.×100%	Body wt.	Liver wt.	L.t/B.T.×100%	p<	p <	_p <
One-week	21.28±0.68	1.20±0.10	5.66±0.337	20.50±0.44	1.20±0.06	5.81±0.22	n.s		ns
Two-weeks	25.0±0.365	1.50±0.03	6.07±0.15	25.0±0.30	1.56±0.03	6.195±0.11		0.05	ns
Four-weeks	28.16±0.20	1.66±0.03	5.90±0.11	26.0±0.70	1.60±0.08	6.19±0.16	0.001	ns	0.05
Six-weeks	30.0±0.258	1.6±0.03	5.30±0.06	28.60±0.50	2.01±0.06	7.0 ± 0.11	0.01	0.001	0.001
Eight-weeks	31.66±0.66	2.0±0.049	6.40±0.07	26.65±0.55	2.44±0.04	8.857±0.29	0.001	0.001	0.001

Data are means (±SE) of body wt., liver wt. and liver wt./body wt. ratio. Significant change as compared to control (p<0.001: highly significant, p<0.01: significant, 0.05: slightly significant and ns: insignificant)

Table 5: Determination of total protein in the liver of control and infected mice during different durations post infection with Schistosoma mansoni

	Durations post in	Durations post infection							
	One week±SE	Two-weeks±SE	Four-weeks±SE	Six-weeks±SE	Eight-weeks±SE				
Control	147.40±2.90	143.66±3.36	137.26±2.60	143.20±2.30	148.60±1.55				
Infected	140.44±1.30	144.20±3.66	172.35 ± 1.38	126.12±1.227	124.80±1.10				
p<	0.01	ns	0.001	0.001	0.001				

Data are means (±SE) of total protein level. Significant change as compared to control (p<0.001: highly significant, p<0.01: significant and ns: insignificant) Total protein is expressed in mg protein/g tissue used

infection. After four weeks of infection the total protein showed a highly significant increase with percentage 25.50%, while after six and eight weeks of infection the total protein showed a highly significant decrease with percentage 11.90 and 16.0%, respectively.

Discussion

It was previously reported that the understanding of how schistosomes interact with their hosts, which is a highly complex strategy, may be one mechanism by which a suitable vaccine or chemotherapeutic drug could be developed (El-Ansary and Daihan, 2005). Previous authors reported that granuloma macrophages isolated from hepatic, intestinal and pulmonary lesions were found to release significant amounts of O_2 and H_2O_2 radicals (Shaheen *et al.*,1994). The oxidative processes which occur upon contact with *S. mansoni* eggs trapped in the liver seem to proceed uncontrolled, since the enzymatic activities involved in O_2 and H_2O_2 detoxification decrease drastically. Such events may be, at least in part, responsible for the pathology associated with schistosomiasis (Gharib *et al.*, 1999). The infection with *S.mansoni* not only triggers the production of reactive oxygen species, but it also leads to the alteration of the antioxidant defence mechanism (Pascal *et al.*, 2000).

The data obtained in the present study show that lipid peroxides were elevated by *S. mansoni* throughout the different durations of infection. It should be pointed out that fibrosis associated with the disease is stimulated by reactive end products of lipid peroxidation (Bedossa *et al.*, 1994; Soliman *et al.*, 2001). In addition, there is a positive correlation between collagen deposition during schistosomal infections and production of malonaldehyde by hepatic cells (Parola *et al.*, 1996).

Results on glutathione content in infected mice livers revealed a highly significant reduction resulting from an oxidative stress due to schistosomiasis. Such depletion may be caused by increased cytotoxicity with H_2O_2 which leads to inhibition in glutathione reductase, the latter responsible for keeping glutathione in its reduced state (Harlan *et al.*, 1984). An interesting finding which coincides with the present data was shown by Yegen *et al.* (1990) that reduction of cellular GSH is accompanied by increased lipid peroxidation.

The present work was also extended to investigate the level of vitamin C in the different Schistosoma infected mice groups. It was found that this vitamin was significantly reduced to scavenge the free radicals produced by the parasite since peroxyl radicals are effectively trapped by ascorbate (Frei *et al.*, 1989).

Data obtained for vitamin E in infected mice livers revealed a significant reduction in the vitamin since the latter is required to protect biological membranes against oxidative degeneration. The administration of vitamin E in the diet shows a strong antioxidant activity against lipid peroxidation and also it protects hepatocytes against toxic injury (Nardini *et al.*, 1993; Sokol *et al.*, 1998). Reduction in the vitamin content due to oxidative stress was previously reported by Warren and Reed (1991).

In the present research, the activity of superoxide dismutase greatly declined one week post infection while an increase in the enzyme activity started from the 4th week till the end of the infection period. The decrease in SOD may result from production of H_2O_2 during oxidative metabolism as indicated by Pinteaux *et al.* (1996). Moreover the increase in enzyme activity may be an adaptive response to conditions of increased peroxidative stress in the liver. Previously, Shaheen *et al.* (1994) reported an increase in hepatic SOD in mice infected with *S. mansoni*. An interesting finding was reported by Sanz *et al.* (1996) who showed that cell defence mechanisms against oxidative toxicity increase in liver to suppress oxidative imbalance, thus SOD increases and GSH decreases.

Furthermore, the present data reveals a highly significant and progressive reduction in catalase activity which started four weeks post *S. mansoni* infection. In agreement with this, Gharib *et al.* (1999) showed that peroxide dismutation yields H_2O_2 which is detoxified by catalase resulting in decrease in its activity. In a recent study, Hanna *et al.* (2005) added that eosinophil peroxidase and its substrate H_2O_2 are released by inflammatory cells in the immediate vicinity of parasite eggs. Also in a more recent study El-Rigal *et al.* (2006) reported the inhibitory effect of *S.mansoni* on glutathione, vitamin C, vitamin E and catalase in infected mice while lipid peroxides increased.

With respect to body and liver weights, it was found that a significant reduction in body weight was recorded after four weeks of infection. These changes may occur due to the presence of the developing parasite worms and the initiation of egg deposition and also due to several metabolites released by the parasite which affect the host hepatic tissue (El-Marzouki and Amin, 1997). In a good connection to the present results, Ahmed and Gad (1995); Magalhaes *et al.* (1995); Fiore *et al.* (1996) and Soliman *et al.* (2001) found a reduction in body weight, an increase in liver weight and an increase in liver weight from the sixth week post-infection with *S. mansoni*, with initiation of schistosomal egg deposition.

On the other hand, data on total proteins recorded an increase after four weeks of infection then a significant decline after six and eight weeks. In hepatic fibrosis as a result of bilharzial infection, protein anabolism decreases while protein catabolism increases. Impairment in protein synthesis was previously supported by Coutinho *et al.* (2002) that malabsorption may be a contributing factor in decrease of protein synthesis through a defect in absorption of amino acids.

In conclusion, the objective of the present research was to study some aspects concerning host-parasite interaction. Previous studies have dealt with the production of free radicals in schistosomiasis, but only after egg deposition in liver and granuloma formation. No reports were cited concerning the trace of these elaborated substances throughout the developmental stages of the parasite in order to provide knowledge on the host-parasite complex interaction, to elucidate stage-specific markers and to develop a suitable target drug that can counteract the parasite at a certain stage of infection. The free radicals are mostly released by inflammatory cells in the vicinity of parasite eggs and the collagen fibers built around eggs create a barrier preventing released compounds from diffusing freely into surrounding areas. This allows the parasitized liver to cope with the threat posed by parasite eggs. This study may help to elucidate stage-specific bioactive markers which may help to develop novel intervention strategies that can modulate the host immune system.

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